Our understanding of the organization of postsynaptic signaling systems at excitatory synapses has been aided by the identification of proteins in the postsynaptic density (PSD) fraction, a subcellular fraction enriched in structures with the morphology of PSDs. In this study, we have completed the identification of most major proteins in the PSD fraction with the use of an analytical method based on mass spectrometry coupled with searching of the protein sequence databases. At least one protein in each of 26 prominent protein bands from the PSD fraction has now been identified. We found 7 proteins not previously known to be constituents of the PSD fraction and 24 that had previously been associated with the PSD by other methods. The newly identified proteins include the heavy chain of myosin-Va (dilute myosin), a motor protein thought to be involved in vesicle trafficking, and the mammalian homolog of the yeast septin protein cdc10, which is important for bud formation in yeast. Both myosin-Va and cdc10 are threefold to fivefold enriched in the PSD fraction over brain homogenates. Immunocytochemical localization of myosin-Va in cultured hippocampal neurons shows that it partially colocalizes with PSD-95 at synapses and is also diffusely localized in cell bodies, dendrites, and axons. Cdc10 has a punctate distribution in cell bodies and dendrites, with some of the puncta colocalizing with PSD-95. The results support a role for myosin-Va in transport of materials into spines and for septins in the formation or maintenance of spines.

Key words: synaptic transmission; myosin; septins; vesicle transport; signal transduction; multiprotein complex

CNS synapses are elaborately organized sites of communication between neurons. Excitatory CNS synapses feature a prominent thickening at the cytoplasmic surface of the postsynaptic membrane termed the postsynaptic density (PSD). The PSD contains receptors with associated signaling and scaffolding proteins that organize signal transduction pathways near the postsynaptic membrane. Proposed functions for PSD proteins include regulation of adhesion between presynaptic and postsynaptic membranes (Siekevitz, 1985; Apperson et al., 1996), control of postsynaptic receptor clustering and function (Siekevitz, 1985; Sheng, 1996), and signal transduction in response to receptor activation (Kennedy, 1993). The PSD fraction is a subcellular fraction highly enriched in multiprotein structures derived from PSDs (Cotman et al., 1974; Cohen et al., 1977). Separation of proteins in this fraction by SDS-PAGE reveals ~15 major and 11 minor protein bands (Kennedy, 1997). The recent identification of signaling and scaffold proteins among the major bands in the PSD fraction confirms several of the proposed functions for PSDs (Kennedy, 1997, 1998; Ziff, 1997).

Signaling molecules that make up >1% of the total protein in the PSD fraction include the NR2A and NR2B subunits of the NMDA receptor (Moon et al., 1994), the α and β subunits of Ca^{2+}/calmodulin-dependent protein kinase II (CaMKII) (Kennedy et al., 1983; Kelly et al., 1984; Miller and Kennedy, 1985), and synGAP, a ras GTPase-activating protein phosphorylated by CaMKII (Chen et al., 1998). Other prominent PSD proteins are scaffold molecules, including the PSD-95 family (Cho et al., 1992; Kistner et al., 1993; Brennan et al., 1996; Kim et al., 1996; Lau et al., 1996), that link receptors to signaling proteins or to the cytoskeleton, thus helping organize the structure of PSDs (Kornau et al., 1995; Kim et al., 1995, 1998; Irie et al., 1997; Chen et al., 1998). Finally, densin-180, a putative adhesion protein with extracellular and intracellular protein binding domains, is a prominent component of the PSD fraction (Apperson et al., 1996).

Despite the recent progress in identifying PSD proteins, several protein bands in the PSD fraction have remained unidentified, and the functions of the PSD are as yet incompletely understood. In the present study, we used a new method based on mass spectrometry (Jensen et al., 1997) to rapidly and systematically identify proteins in the PSD fraction by comparing their tryptic peptide profiles with those of proteins in the protein sequence databases. In addition to many previously identified PSD proteins, we identified several proteins not previously known to be constituents of the PSD fraction. Two of these, myosin-Va (dilute myosin) and the septin protein cdc10, were selected for further study because they have the most intriguing potential functions. We report that both myosin-Va and cdc10 are enriched in the
PSD fraction and are among the more abundant proteins there. They both partially colocalize with PSD-95 at postsynaptic sites along dendrites in cultured hippocampal neurons.

MATERIALS AND METHODS

Isolation of the PSD fraction. PSD fractions were prepared from rat forebrains as previously described (Carlin et al., 1980; Cho et al., 1992). Synaptosomes were isolated from homogenates by differential and density gradient centrifugation and then extracted with 0.5% Triton X-100 for 15 min. The resulting “One-Triton” PSD fraction was pelleted by centrifugation at 60,800 × g for 45 min. A portion of the One-Triton fraction was extracted again either with 0.5% Triton X-100 for 15 min or with 3% N-lauryl-sarcosine for 10 min and then pelleted by centrifugation at 201,800 × g for 1 hr to obtain the “Two-Triton” PSD fraction or the “One-Triton plus Sarcosyl” PSD fraction, respectively. Protein concentrations were determined by a modified method of Lowry (Peterson, 1983).

Identification of proteins in the PSD fraction by mass spectrometry. Protein identification was performed by mass spectrometry combined with sequence database searches (Jensen et al., 1998). Protein bands cut from a Coomassie blue-stained polyacrylamide gel were rinsed, reduced, S-alkylated, and then incubated with trypsin (Shevchenko et al., 1996b). The resulting tryptic peptide mixtures were directly analyzed by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF-MS) (Jensen et al., 1996). Peptide mass spectra displayed up to 150 peptide ion signals for some of the larger proteins in the PSD fraction. The MALDI-TOF mass spectrometer (REFLEX, Bruker Daltonics, Bremen, Germany) was operated in the delayed extraction, positive ion mode. Samples were prepared according to the fast evaporation deposition method using o-cyano-4-hydroxycinnamic acid and nitrocellulose as the matrix (Vorm et al., 1994; Jensen et al., 1996). Ion signals produced by trypsin autodigestion peptides, which were present in all peptide mass spectra, were used for internal mass calibration. Peptide mass errors were typically in the range of 15–25 ppm.

Proteins were identified by searching databases with lists of tryptic peptide masses (peptide mass maps) generated from the proteins by MALDI-TOF MS. PeptideSearch or ProFound software (accessible on the Internet via http://www.protein.sdu.dk) was used to query nonredundant protein sequence databases with peptide mass data. Protein mixtures of up to five proteins were resolved by iterative database searches as described (Jensen et al., 1997).

Cloning of the cDNA encoding rat cdc10. Oligonucleotide primers were designed based on the nucleotide sequence of human cdc10 (hcde10; GenBank accession number X72008). First-strand cDNAs were prepared as described (Jensen et al., 1997). The Internet via http://www.protein.sdu.dk) was used to query nonredundant protein sequence databases with the complete coding region, so we used a strategy based on PCR to find the remaining coding sequence. The 5′ end of the rat cdc10 sequence was amplified by PCR with a sense primer (5′-GGCGGGCCTAGGTGGAAGATCCG-3′) and an antisense primer corresponding to hcde10 bp 422–441 (5′-CTGTTCATCTGTTGATTTG-3′). The product of the PCR reaction was ligated into the TA vector pCR2.1 and transformed into INVoF® cells (Invitrogen, San Diego, CA). The cultured neurons were sequenced on an automatic DNA sequencer in the Caltech DNA Microsequencing Facility.

We screened a ygt11 rat brain cDNA library (Clontech) for clones hybridizing to the cdc10 PCR product. A single 1041 bp clone and two copies of a 1116 bp clone were isolated and sequenced. The 5′ end of all three clones begins at an EcoRI site homologous to hcdc10 bp 140. The RI site that is absent in hcdc10. Neither of the cDNAs contained the complete coding region, so we used a strategy based on PCR to find the remaining coding sequence. The 5′ end of the rat cdc10 sequence was amplified by PCR with a sense primer (5′-GGCGGGCCTAGGTGGAAGATCCG-3′) and an antisense primer matching bp 1379–1399 of hcde10 (5′-AAGACGACGAACTGAAAC-3′). Each PCR reaction gave a single product of the appropriate size. The PCR products were ligated into the TA vector and sequenced in both directions. The portions of the reported sequence of rat cdc10 that were based on results of PCR amplification were derived from the sequences of at least four independent PCR products. The overlapping sequences were assembled with Sequencher software (Gene Codes Corp., Ann Arbor, MI) into a 1494 bp sequence containing the complete coding regions of rat cdc10.

Construction of fusion proteins and preparation of antibodies. A glutathione-S-transferase (GST):myosin-Va fusion construct was made by inserting a 2439 bp cDNA encoding amino acids 1042–1854 of mouse myosin-Va heavy chain (Seperack et al., 1995) into pGEX-5X-2 (Pharmacia Biotech, Piscataway, NJ). We prepared two GST:cdc10 fusion proteins in pGEX-5X-1. One contained base pairs 238–538 (encoding amino acids 49–148); the other contained base pairs 238–1352 (encoding amino acids 49–420). The proper orientation of the sequence for each construct was verified by restriction digestion and sequencing. Fusion proteins were produced in Escherichia coli DH5a. After transformation, the cells were grown at 30°C to an optical density of 0.5 at 600 nm, and expression of GST fusion proteins was induced by addition of 0.1 mM isopropyl-β-D-thiogalactopyranoside for 5 hr at 30°C. The cells were pelleted by centrifugation at 5000 × g for 10 min and resuspended in PBS plus protease inhibitors (20 mM sodium phosphate, pH 7.4, and 0.5% NP40 detergent, 1 mM EDTA, 1 mM EGTA, 20 μg/ml aprotinin, 5 μg/ml antipain, and 0.4 μg/ml pepstatin; 17 μg/ml of PMSF was added at each extraction step). The suspended cells were lysed by sonication [2 min, level 5, 50% cycle with a Branson (Danbury, CT) 450 Sonifier]. Triton X-100 was added to a final concentration of 1%, and the solution was stirred for 10 min. Lysates were cleared by centrifugation at 15,000 × g for 10 min. The supernatant was saved on ice, and the pellets were resuspended in PBS plus protease inhibitors. N-Laurylsarcosine was added to a final concentration of 1%, and the suspension was sonicated as above. Triton X-100 was then added again to a final concentration of 1%, and the lysate was cleared by centrifugation as above. The two supernatants were pooled, 2 ml of glutathione-conjugated agarose beads (Sigma, St. Louis, MO) was added, and the solution was rotated end over end at 4°C overnight at room temperature. The supernatant was removed, and the beads were washed three times with PBS. The GST fusion proteins were eluted overnight in 50 mM Tris, pH 8.0, 20 mM glutathione, and 1% Triton X-100, and the supernatant was removed after centrifugation to remove the beads.

The eluted fusion proteins (~10 μg/injection) were injected into Swiss–Webster mice to generate polyclonal ascites fluid (Ou et al., 1993). The specificity of the antibodies was tested by immunoblotting against both purified fusion proteins and brain homogenates. Two mice produced antibodies specific for myosin-Va (antibodies DBI-B and DBI-C). Three mice inoculated with the cdc10 47–149 construct produced antibodies specific for cdc10 (antibodies N1, D1, and D2), as did three mice inoculated with the cdc10 47–420 fusion protein (antibodies L1, L2, and L3). Ascites fluids were partially purified by 50% ammonium sulfate precipitation overnight at 4°C, concentrated, and then dialyzed into 0.1 M Tris, pH 7.5. The concentration of IgG was estimated by comparison with mouse IgG standards on Coomassie blue-stained gels (DBI-C, 2 μg/ml, L2, 1.5 μg/ml).

Immunocytochemical labeling of dissociated hippocampal neurons. Cultures of hippocampal neurons from embryonic day 18 rats were grown on poly-D-lysine- and laminin-coated coverslips at a density of ~200/mm² (Brewer et al., 1993; Apperson et al., 1996). After 3–5 weeks in vitro, the coverslips were removed from the culture media, washed in HBSS with 10 mM HEPES, pH 7.4, and fixed with ~20°C methanol for 20 min. The fixed cells were rehydrated with HBBS and incubated in preblock buffer (20 mM phosphate buffer, pH 7.4, 5% normal goat serum, 0.05% Triton X-100, and 450 mM NaCl) for 1 hr at 4°C. They were then incubated overnight at 4°C with either DB1-C antibodies at a 1:400 dilution or L2 antibodies at a 1:200 dilution. Polyclonal rabbit antibodies against PSD-95 (Cho et al., 1992) were added at a dilution of 1:50. The cultures were washed in preblock solution, and Cy3-conjugated goat anti-mouse secondary antibodies and FITC-conjugated goat anti-rabbit antibodies were added at a dilution of 1:100 in preblock buffer and incubated for 1 hr at room temperature. Coverslips were washed once in preblock buffer and twice in PBS and then post-fixed for 10 min with 2% paraformaldehyde. The cultures were then rinsed with PBS with 0.1 mM sodium bicarbonate, pH 9.2, and then mounted on slides with 90% glycerol, 4% n-propyl gallate, and 0.1 mM sodium bicarbonate, pH 9.2. The immunostained cells were viewed using a Zeiss (Thornwood, NY) LSM310 fluorescence laser-scanning confocal microscope. A 63× oil
immersion objective was used at electronic zoom factors from 1 to 2. Images were scanned for 64 sec using contrast settings from 320 to 410 and brightness settings from 9600 to 9700. Images were aligned and colorized with Adobe (Mountain View, CA) Photoshop without adjusting the original data.

For control immunolabeling, the DB1-C and L2 antisera were preab-sorbed with their respective antigens before application to cultures. Each antiserum was mixed with the appropriate GST fusion protein at a ratio of 1 mol of IgG to 3 mol of antigen.

**Immunohistochemistry.** Forebrain homogenate, synaptosomes, and detergent-extracted PSD fractions were separated by SDS-PAGE under reducing conditions and electrophoretically transferred to nitrocellulose membranes. The membranes were blocked at least 2 hr in 5% nonfat milk in TBST (10 mM Tris, pH 7.5, 200 mM NaCl, and 0.2% Tween 20) and incubated with DB1-C antibodies (diluted 1:1500) or L2 antibodies (diluted 1:1000) for 5 hr. Bound antibodies were detected by the alkaline phosphatase method using secondary antibodies purchased from Boehringer Mannheim (Indianapolis, IN).

**RESULTS**

**Strategy for identification of proteins in the PSD fraction by MALDI-TOF mass spectrometry**

To identify proteins in the PSD fraction, the Two-Triton PSD fraction (30 μg) was fractionated by SDS-PAGE, and proteins were stained with Coomassie blue. The gel was photographed, and 26 individual protein bands were coded for study. Individual protein bands were then excised and digested with trypsin in the gel (Rosenfeld et al., 1992; Jeno et al., 1995; Shevchenko et al., 1996a). The mass-to-charge ratio of the peptides released from the gel was measured by MALDI-TOF MS with high mass accuracy (Jensen et al., 1996). The complete set of peptide masses from each protein band was then compared with the tryptic peptide masses predicted for each protein in a comprehensive nonredundant database (Fig. 1).

A protein from the PSD fraction was considered identified when the spectrum of its measured peptide masses met the previously established criteria for positive identification of proteins using MALDI-TOF MS and automated database searching (Jensen et al., 1997). First, to distinguish a valid match from a false positive, a minimum of five measured peptide masses must match tryptic peptide masses calculated for an individual protein in the database, with <50 ppm deviation in mass between measured and calculated values. Second, the peptides identified by these matches must provide at least 15% sequence coverage of the identified protein. Other criteria are also considered, such as the percentage of the total number of observed peaks that can be assigned to a putative match and the similarity in molecular weight of the unknown protein to the putative match. It is important to note that multiple proteins comigrating in a single band can be identified by removing the peptide masses assigned to the first identified protein from the complete list of masses and using the remaining list of masses to rescan the database (Jensen et al., 1997).

For this study, we searched the database for proteins with a mass range of 0–700 kDa for large proteins (apparent Mr >200 kDa by SDS-PAGE) and 0–300 kDa for the smaller proteins, with no constraint on species of origin. After the initial identification of a protein, a “second pass search” was conducted. In this step, incomplete tryptic cleavage and peptide modifications that may alter the peptide masses, such as oxidized methionine or S-acrylamidocysteine, were calculated for the putatively identified protein and compared with the measured masses. The modified peptides identified in the second pass search were added to the list generated in the first pass search to increase the number of matching peptides and sequence coverage.

**Identification of myosin-Va (dilute myosin)**

Figure 2 illustrates the peptide mass map of a previously uniden-tified protein band in the PSD fraction containing proteins of apparent mass 190 kDa. The complete set of peptide masses from this band was found to contain peptides from two proteins. Thirty-six of the measured peptide masses matched theoretical tryptic peptide masses calculated for the heavy chain of the unconventional myosin, myosin-Va, (also called dilute myosin; accession number Q99104), a protein with a predicted mass of 215 kDa (Table 1). The matching peptides cover 451 of 1853 amino acids, or 24% of the myosin-Va sequence. The peptide masses exclude a match with the homologous protein myosin-Vb, illustrating the power of this technique to unambiguously identify a protein isoform. The peptide masses assigned to myosin-Va were then removed from the mass spectrum list, and the database was queried with the remaining masses. Twenty-three of the remaining peptides were assigned to the abundant PSD protein αII-spectrin (accession number X90845), also called fodrin (Glenney et al., 1982). Myosin-Va migrates faster than full-length αII-spectrin. However, apparently because αII-spectrin is a particularly abundant protein in the Two-Triton PSD fraction, peptides from αII-spectrin were detected in the digest of the myosin-Va band as well as in digests of several other protein bands of lower molecular weight.

**Identification of cdc10**

Figure 3 illustrates the peptide mass map of a previously uniden-tified PSD protein band in the PSD fraction containing a protein of apparent mass 45 kDa. Eleven peptide masses matched the theoretical peptide masses calculated for the mouse septin protein.
cdc10 (accession number O55131), a protein with a predicted mass of 48–50 kDa. The matching peptides cover 136 of 436 amino acids, or 31% of the sequence (Table 2). Septins are a family of proteins that form membrane-associated filaments that recruit other proteins to establish specialized domains (Field and Kellogg, 1999). They play well established roles in formation of the yeast bud site and neck constriction before cytokinesis (see Discussion).

Because no cdc10 ortholog had yet been isolated from rat, we screened a Agt11 rat brain cDNA library with a fragment of cdc10 obtained by PCR amplification from first-strand adult rat cDNAs. We isolated two identical 1116 bp clones and one 1041 bp clone, the sequence of which was entirely contained in the longer clones. The clones contained all of the coding region for rat cdc10 except 144 bp at the 5′ end and 50 bp at the 3′ end. The sequences of these regions were obtained after their amplification by PCR (see Materials and Methods), and the complete 1494 bp sequence was assembled and submitted to GenBank (accession number AF142759). The 1310 bp continuous open reading frame contains a potential alternative initiation codon 57 bp after the first. The DNA sequence of the second potential initiation codon better fits the sequence defined by Kozak (1989, 1997). However, initiation of translation may potentially occur at either start site to give proteins of either 419 or 436 aa, with predicted masses of 48.6 or 50.5 kDa. The deduced amino acid sequence of rat cdc10 is >99% identical to that of mouse, with only a Gly to Ser substitution at residue 216. The sequences of the mouse cdc10 peptides that were matched in the MALDI-TOF MS experiments are identical to those of rat cdc10.

**Myosin-Va and cdc10 are enriched in the PSD fraction**

One criteria that we have used to assess the specificity of association of a protein with the PSD fraction is its enrichment in the PSD fraction compared with forebrain homogenate and synapto- somes (Cho et al., 1992; Apperson et al., 1996). We raised antisera against fusion proteins containing GST and C-terminal sequences of myosin-Va or cdc10 as described in Materials and Methods. The antisera recognize protein bands of the correct molecular weights on immunoblots of forebrain homogenates (Fig. 4A,B). The doublet of ~48–50 kDa recognized by the antibody against cdc10 may arise from initiation at both of the two potential initiation sites in the cdc10 message. Immunoblots of forebrain homogenates (FBH), synapto- somes, and One-Triton and Two-Triton PSD fractions with anti-myosin-Va revealed a single band of 190 kDa in each fraction, with myosin-Va enriched approximately threefold in the PSD fractions compared with FBH (Fig. 4A). Similar immunoblots with anti-cdc10 revealed that it is enriched approximately fivefold in the PSD fractions compared with FBH (Fig. 4B). Some cdc10 remains in the sarcosyl-treated PSD fraction, although it is significantly diminished compared with the One-Triton and Two-Triton PSD fractions. The fold enrichment of these two proteins in the PSD fraction is not as large as that of PSD-95 (Cho et al., 1992), densin-180 (Apperson et al., 1996), or the NR2B subunit of the NMDA receptor (Moon et al., 1994), each of which is enriched ~10- to 30-fold in the PSD fraction compared with FBH, and each of which is localized nearly exclusively at synapses in dendrites of cultured hippocampal neurons (Kornau et al., 1995; Apperson et al., 1996). The threefold to fivefold enrichments of myosin-Va and cdc10 in the PSD fraction suggest that they may be concentrated at the PSD, but that significant portions of the two proteins are also located elsewhere in neurons or glia.

**Myosin-Va and cdc10 are located at synapses in cultures of dissociated hippocampal neurons**

We investigated the subcellular location of myosin-Va and cdc10 by fluorescence immunocytochemistry. Dissociated hippocampal neurons plated at embryonic day 18 (E18) and grown in culture for 3–5 weeks were double-stained with antibodies against either myosin-Va or cdc10 and the PSD protein PSD-95 as described in Materials and Methods. High-resolution imaging with the laser scanning confocal microscope revealed that both myosin-Va and cdc10 partially colocalize with PSD-95 at discrete sites on dendrites (Fig. 5A,B). The myosin-Va and cdc10 antibodies also stain the neuronal cell body. Diffuse myosin-Va staining is visible throughout dendritic shafts and in axons, which form a network of fine processes around the thicker dendrites. Staining for cdc10 is...
also visible in the shafts, but only as discrete puncta that do not colocalize with PSD-95. Thus neither myosin-Va nor cdc10 is exclusively confined to synapses.

Control cultures were stained with primary antibodies against myosin-Va or cdc10 that had been preabsorbed with the appropriate antigen as described in Materials and Methods. Under these conditions, almost no staining was visible; therefore Figure 5 accurately represents the subcellular distributions of the two proteins.

Additional proteins identified in the Two-Triton PSD fraction

Twenty-six protein bands of the Two-Triton PSD fraction were analyzed as above by tryptic digestion followed by MALDI-TOF MS and an automated search of the database. Thirty-one individual proteins, including myosin-Va and cdc10, were identified and assigned to specific protein bands (Fig. 6A,B).

Several signaling proteins previously identified in the PSD fraction by microsequencing or by other biochemical methods were also found in this study (Fig. 6B): NR1 and NR2B subunits of the NMDA-type glutamate receptor (Moon et al., 1994); the α and β subunits of CaMKII (Kennedy et al., 1983; Kelly et al., 1984; Miller and Kennedy, 1985); synGAP, a synapse-specific Ras GTPase-activating protein (Chen et al., 1998; Kim et al., 1998); citron, a target for Rac GTPases (Zhang et al., 1999); and an insulin receptor tyrosine kinase 53 kDa substrate protein (Yeh et al., 1996; Abbott et al., 1999). Each of these proteins has previously been localized by immunocytochemistry to synapses in dissociated hippocampal neurons (Kennedy et al., 1990; Kornau et al., 1995; Chen et al., 1998; Kennedy, 1998; Kim et al., 1998; Abbott et al., 1999; Zhang et al., 1999).

Similarly, we identified scaffold and cytoskeletal proteins that have been identified previously in the PSD fraction by other methods. These proteins include scaffold proteins PSD-95 (Cho et al., 1992; Kistner et al., 1993; Kornau et al., 1995) and homer (Brakeman et al., 1997; Kato et al., 1997, 1998; Naisbitt et al., 1999; Tu et al., 1998, 1999) and cytoskeletal proteins, α-actinin (Wyszynski et al., 1997, 1998), bassoon (tom Dieck et al., 1998), αII- and β-spectrin (Carlin et al., 1983), β-actin (Kelly and Cot-

### Table 1. Myosin-Va peptides identified from the MALDI-TOF peptide mass map shown in Figure 2

<table>
<thead>
<tr>
<th>Measured mass</th>
<th>Calculated mass</th>
<th>Myosin-Va residues</th>
<th>Amino acid sequencea</th>
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<tr>
<td>2480.246</td>
<td>2480.197</td>
<td>587–609</td>
<td>(K)MLPEFLFDDEKAIPTSATSSGR(T)</td>
</tr>
<tr>
<td>2513.372</td>
<td>2513.270</td>
<td>1468–1487</td>
<td>(K)EKDFQGMLEYKREDEQKLVK(N)</td>
</tr>
<tr>
<td>2553.202</td>
<td>2553.229</td>
<td>1332–1353</td>
<td>(R)SHENAEALRGEIQSKEENR(Q)</td>
</tr>
<tr>
<td>2630.273</td>
<td>2630.382</td>
<td>30–51</td>
<td>(K)DJKPKGDKVLILLSLEEGKDLEYR(L)</td>
</tr>
</tbody>
</table>

a M*, Methionine sulfoxide; C, S-acrylamidocysteine.
man, 1978; Matus et al., 1982; Adam and Matus, 1996), tubulin (Kelly and Cotman, 1978), a brain-specific intermediate filament protein termed \( \alpha \)-internexin, (Suzuki et al., 1997), and neurofilaments M and L (Kelly and Cotman, 1978). We identified one cytoskeletal protein that has not previously been reported in the PSD fraction, plectin (Fig. 6A). Plectin is a 300 kDa protein that associates with intermediate filaments, actin, and tubulin (Wiche et al., 1991). We have not verified its location at synapses.

We identified a glycolytic enzyme, glyceraldehyde-3-phosphate dehydrogenase (G3PDH), previously reported to be in synaptosomes and in the PSD fraction (Rogalski-Wilk and Cohen, 1997; Wu et al., 1997; Moon et al., 1998). G3PDH binds to F-actin and may be anchored to the PSD via this interaction (Rogalski-Wilk and Cohen, 1997). We found Hsc-70, a constitutively expressed form of the 70 kDa heat shock protein family (Kiang and Tsokos, 1998), that has previously been found in the PSD fraction (Suzuki et al., 1999) and was recently shown by immunocytochemistry to be present at synaptic junctions (S. N. Baek, I. S. Park, I. Jin, L. T. Schenker, M. B. Kennedy, and I. S. Moon, unpublished results).

Additional proteins identified in this study, but not previously reported in the PSD fraction (Fig. 6A), include contactin, a glycosylphosphatidylinositol-linked glycoprotein of the Ig superfamily (Reid et al., 1994; Langnaese et al., 1998), and KIAA0378, a protein of unknown function encoded by an open reading frame deposited in the human genomic database (Nagase et al., 1997). Some of the proteins that we detected in this study are known to be, or appear to be, contaminants of the PSD fraction in the sense that they are not present at the postsynaptic site in fixed tissue. These include glial fibrillary acidic protein (Matus et al., 1980), which is not expressed in neurons, and synapsin, which is located principally in the presynaptic terminal (De Camilli et al., 1983). Synapsin may cofractionate with the PSD fraction by virtue of its affinity for CaM kinase II (Benfenati et al., 1992). The other presynaptic protein found in the PSD fraction, bassoon (tom Dieck et al., 1998), may also cofractionate anomalously with the PSD, or, more interestingly, it may be bound to junctional proteins in the PSD fraction that span the synaptic cleft in vivo. We found a group of mitochondrial proteins, including two inner membrane proteins, the ATP/ADP carrier (Fiore et al., 1998),

Table 2. CDC10 peptides identified from the MALDI-TOF peptide mass map shown in Figure 3

<table>
<thead>
<tr>
<th>Measured mass</th>
<th>Calculated mass</th>
<th>CDC10 residues</th>
<th>Amino acid Sequence$^a$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1243.545</td>
<td>1243.559</td>
<td>137–146</td>
<td>(K)FEDYLNASES(V)</td>
</tr>
<tr>
<td>1361.576</td>
<td>1361.648</td>
<td>333–343</td>
<td>(K)SPLAQM*EEERR(E)</td>
</tr>
<tr>
<td>1523.622</td>
<td>1523.687</td>
<td>298–309</td>
<td>(K)DVTTNYHENEYR(S)</td>
</tr>
<tr>
<td>1686.778</td>
<td>1686.841</td>
<td>47–62</td>
<td>(R)GFEFTLM*VGESGLGK(S)</td>
</tr>
<tr>
<td>1708.763</td>
<td>1708.821</td>
<td>195–208</td>
<td>(K)ADTLTPEEC*QFKK(Q)</td>
</tr>
<tr>
<td>1792.785</td>
<td>1792.825</td>
<td>402–415</td>
<td>(R)QFEEEKANWEOARQ(R)</td>
</tr>
<tr>
<td>1842.890</td>
<td>1842.942</td>
<td>46–62</td>
<td>(K)RGFEFTLM*VGESGLGK(S)</td>
</tr>
<tr>
<td>1953.961</td>
<td>1953.982</td>
<td>25–41</td>
<td>(K)NLEGYVGAFNPOYR(K)</td>
</tr>
<tr>
<td>2082.036</td>
<td>2082.077</td>
<td>25–42</td>
<td>(K)NLEGYVGAFPOYR(K)</td>
</tr>
<tr>
<td>2097.988</td>
<td>2097.986</td>
<td>221–237</td>
<td>(K)YEPEDEEEEEENKLV(K)</td>
</tr>
<tr>
<td>2607.417</td>
<td>2607.309</td>
<td>63–85</td>
<td>(K)STLINSFLTDLVESPEYPGPSR(I)</td>
</tr>
</tbody>
</table>

$^a$ M*, Methionine sulfoxide; C*, S-acrylamidocysteine.
thick gels loaded with ~30 μg of protein per lane. The protein composition of postsynaptic densities may vary in different brain regions depending on the predominant neuronal cell type. The mass spectrometric method is sensitive enough that the protein compositions of PSD fractions isolated from particular brain regions, or after defined electrophysiological manipulations, could be analyzed and compared in their entirety. The amount of tissue needed for such experiments would be determined only by that required for a clean sucrose density fractionation during the preparation of the PSD fraction. The method cannot, however, be used to measure the precise stoichiometric relationships among proteins in a complex, because the sizes of the peptide mass peaks obtained in the mass scan are determined in part by the extent to which individual peptides can be volatilized.

Myosin-Va is slightly enriched in the PSD fraction (Fig. 4A) and located in synapses, dendrites, axons, and cell bodies of cultured hippocampal neurons (Fig. 5A). Expression of myosin-Va mRNA in the brain has been demonstrated by in situ hybridization (Mercer et al., 1991). Evidence is accumulating that myosin-Va functions as a motor that transports membrane vesicles along actin filaments (Brown, 1999). In one study, myosin-Va was shown to transport a population of vesicles derived from the endoplasmic reticulum (ER) (Tabb et al., 1998). Myosin-Va is also associated with presynaptic vesicles (Prekeris and Terrian, 1997; Evans et al., 1998). Function-blocking antibodies against myosin-Va completely inhibited the motility of these vesicles in vitro (Evans et al., 1998). Thus, in the presynaptic terminal, myosin-Va may be responsible for docking or transporting synaptic vesicles.

The importance of myosin-Va in dendrites is demonstrated in mice with “dilute” mutations that prevent the expression of myosin-Va and lead to severe neurological deficits culminating in death at ~3 weeks of age. The brains of the mutant mice appear normal on a gross level (Mercer et al., 1991). However, two recent studies demonstrate postsynaptic defects in dendritic spines of their Purkinje cells. In wild-type mice, branches of the smooth ER extend into the shaft of Purkinje neuron spines. Mice with dilute lethal (dl) and dilute-opisthotonos (dop) mutations of myosin-Va are missing the ER in spines of Purkinje cells (Dekker-Ohno et al., 1996; Takagishi et al., 1996), suggesting that defects in transport or anchoring of ER-derived organelles within the spine might contribute to the phenotype associated with the mutations. Myosin-Va mutations in humans have been linked to Griscelli syndrome (Pastural et al., 1997), characterized by albinism, immune deficits, and seizures followed by death in the first decade. The phenotype displayed in these patients is reminiscent of the phenotype of myosin-Va-deficient mice and further demonstrates the importance of myosin-Va in the nervous system.

Myosin-Va was identified in the “first pass search” with peptides derived from the band at 190 kDa (Fig. 6A), indicating that it is likely the most abundant protein in this band. Its abundance in the PSD fraction supports the idea that it may transport vesicles or proteins into spines and PSDs of forebrain neurons. Dendritic spines contain actin filaments that extend through the neck of the spine to the PSD and appear to make contact with the spine apparatus (Fifkova and Delay, 1982; Matus et al., 1982; Cohen et al., 1985; Morales and Fifkova, 1989). These filaments may provide a substrate for transport of postsynaptic proteins and organelles to the synapse by myosin-Va (Brown, 1999). Myosin-Va interacts directly with CaMKII (Costa et al., 1999), which is also an abundant PSD protein, and is phosphorylated by it (Coelho and Larson, 1993). Thus, activation of CaMKII by Ca2+ influx through NMDA receptors could potentially modulate myosin-Va...
motor activity at the synapse. Naisbitt et al. (2000) report that the PSD-95/GKAP (guanylate kinase-associated protein) complex interacts with a light chain that is shared by dynein and myosin V, providing additional evidence that myosin V may be a motor protein in the postsynaptic spine.

The second PSD protein that we studied in detail is the mammalian homolog of cdc10, a member of the septin family. We show that it partially colocalizes with PSD-95 at synapses in cultured hippocampal neurons (Fig. 5B). Cdc10 and the other septins were first identified as proteins that form heterooligomeric filaments that encircle the yeast bud neck (Byers and Goetsch, 1976; Haarer and Pringle, 1987; Ford and Pringle, 1991; Kim et al., 1991). Temperature-sensitive inactivation of any of the septins in yeast results in loss of neck filaments and causes cell cycle arrest and defects in bud growth and cytokinesis (Field and Kellogg, 1999). In the absence of neck filaments, several kinases, and enzymes involved in cytokinesis fail to localize properly to the neck region, indicating that septins may form a scaffold for the assembly of protein complexes (Flescher et al., 1993; Chant et al., 1995; Field and Kellogg, 1999).

Septin homologs have been identified in Drosophila and mammals. They are generally associated with membranes in places where the membrane is undergoing remodeling, such as the site of bud emergence (Flescher et al., 1993; Chant et al., 1995), and extensions of neuronal growth cones (Neufeld and Rubin, 1994; Fares et al., 1995). Cdc10 was recently found to be associated with the exocyst complex in neurons (Hsu et al., 1998), which may be involved in vesicle fusion at the plasma membrane. The association of septins with a complex involved in membrane fusion may indicate that cdc10 plays a role in adding membrane to developing

Figure 5. Immunocytochemical localization of myosin-Va and cdc10 in cultures of dissociated hippocampal neurons. Hippocampal neurons dissociated at E18 were grown in culture for 28 d and then fixed and double-immunostained as described in Materials and Methods. Images of the two fluorophors were colorized and combined (left). At right are the single images of the boxed regions. A, Immunocytochemical localization of myosin-Va and PSD-95. Red indicates Cy3 staining of myosin V, and green indicates FITC staining of PSD-95. Regions of overlap are yellow. Myosin-Va is distributed throughout cell bodies, dendrites, and axons and appears concentrated at synapses, where it colocalizes with PSD-95. B, Immunocytochemical localization of cdc10 and PSD-95. Red indicates Cy3 staining of cdc10, and green indicates FITC staining of PSD-95. CDC10 has a punctate distribution throughout the soma and dendrites. Some, but not all, of the cdc10 in dendrites colocalizes with PSD-95 at synapses. Scale bars, 10 μm.
neuronal processes at sites that could include synapses. Thus, in the PSD, septins may form a cytoskeletal structure for the assembly or addition of proteins at the postsynaptic membrane. The septin scaffold might then persist as part of the PSD in mature synapses. Septin polymerization is regulated by several signaling pathways during cytokinesis in yeast. Therefore, septin polymerization at the synapse may be dynamically regulated in response to synaptic signals.

The identification of α-actinin-1 in the PSD fraction illustrates the sensitivity of the mass spectrometric method. Four forms of human α-actinin are known (α-actinins 1–4). The molecular masses of the peptides from the α-actinin band unambiguously identify the isoform in the PSD fraction as a homolog of human α-actinin-1. Wyszynski et al. (1997, 1998) identified human α-actinin-2 as an interactor with the NMDA receptor subunit NR1 in a yeast two-hybrid screen. Using antibodies against α-actinin-2, they showed that it is concentrated in dendritic spines in hippocampal neurons. Human α-actinin-2 is 79% identical to human α-actinin-1 in amino acid sequence, and all known functional domains are conserved between the two proteins. Our data suggest that the rat form of α-actinin that is present at postsynaptic sites is most homologous to human α-actinin-1 and not α-actinin-2. However, α-actinin-1 and α-actinin-2 are likely to be functionally identical.

We did not find densin-180, which was previously identified in the PSD fraction by microsequencing and localizes at the synapse (Apperson et al., 1996). The presence of densin-180 may have been obscured in this study, because it comigrates with the NR2B subunit of the NMDA receptor (Moon et al., 1994; Apperson et al., 1996) and with citron (Zhang et al., 1999). In addition, densin-180 is extensively glycosylated (Apperson et al., 1996). The altered masses of tryptic peptides containing glycosyl groups may have precluded their identification by mass spectrometry.

Notably, several proteins originally identified by yeast two-hybrid screens for proteins that interact with known PSD proteins were not found in the PSD fraction in our study, including shank (Naisbitt et al., 1999), yotaio (Lin et al., 1998), and GKA (Kim et al., 1997; Naisbitt et al., 1997). Two homologs of PSD-95, SAP102 (Muller et al., 1996) and Chapsyn-110/PSD-93 (Brenman et al., 1996; Kim et al., 1996), were also not identified here, although they have been reported to be enriched in the postsynaptic density. Our inability to identify these proteins may reflect their relatively low abundance in the PSD fraction, although it is also possible that some of these proteins comigrate with more abundant PSD proteins and thus are difficult to detect by mass spectrometry.

It is important to note that low abundance of a protein in the PSD fraction does not necessarily indicate that it is absent from the PSD in vivo. It is quite likely that the association of some proteins with the native PSD is disrupted by extraction with Triton X-100 during the purification of the PSD fraction. Thus, additional methods, such as high-resolution immunolocalization, will be needed to ascertain the full protein composition of the postsynaptic lattice. Nonetheless, the identification of proteins in the PSD fraction has been a useful first step in understanding the organization of signaling molecules at the postsynaptic membrane.

REFERENCES


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Figure 6. Proteins identified in the PSD fraction by MALDI-TOF MS. Thirty micrograms of protein from the Two-Triton PSD fraction were subjected to SDS-PAGE and stained with Coomassie blue. Individual protein bands were isolated, and the proteins in each band were identified by MALDI-TOF mass spectrometry as described in Materials and Methods. The positions of molecular weight standards are shown at left. A, Proteins identified for the first time in this study as constituents of the PSD fraction. The presence of myosin-Va and cdc10 (bold, underlined) at synaptic sites was verified in this study. B, Proteins identified in this study that were previously identified in the PSD fraction by other methods.


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