Biphasic influence of Miz1 on neural crest development by regulating cell survival and apical adhesion complex formation in the developing neural tube

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ABSTRACT Myc interacting zinc finger protein-1 (Miz1) is a transcription factor known to regulate cell cycle- and cell adhesion–related genes in cancer. Here we show that Miz1 also plays a critical role in neural crest development. In the chick, Miz1 is expressed throughout the neural plate and closing neural tube. Its morpholino-mediated knockdown affects neural crest precursor survival, leading to reduction of neural plate border and neural crest specifier genes Msx-1, Pax7, FoxD3, and Sox10. Of interest, Miz1 loss also causes marked reduction of adhesion molecules (N-cadherin, cadherin6B, and α1-catenin) with a concomitant increase of E-cadherin in the neural folds, likely leading to delayed and decreased neural crest emigration. Conversely, Miz1 overexpression results in up-regulation of cadherin6B and FoxD3 expression in the neural folds/neural tube, leading to premature neural crest emigration and increased number of migratory crest cells. Although Miz1 loss effects cell survival and proliferation throughout the neural plate, the neural progenitor marker Sox2 was unaffected, suggesting a neural crest–selective effect. The results suggest that Miz1 is important not only for survival of neural crest precursors, but also for maintenance of integrity of the neural folds and tube, via correct formation of the apical adhesion complex therein.

INTRODUCTION

The neural crest, a multipotent stem/progenitor cell population, arises within the developing nervous system of vertebrate embryos. After induction at the neural plate border, the specification of these cells is manifested by the expression of several neural crest–specifier genes, including FoxD3, Sox10, Ets-1, and Snail2, in the dorsal neural folds. Soon thereafter, neural crest cells undergo an epithelial-to-mesenchymal transition (EMT) to detach from the neural tube and begin migration to various and often distant destinations. Subsequently, they differentiate into multiple and varied cell types, such as neurons and glia of the peripheral nervous system and melanocytes, as well as facial bone and cartilage.

Adhesion molecules play a key role in both neural and neural crest development. N-cadherin (N-cad) expression is prominent in adherens junctions of the neural plate and neural tube, where it maintains apicobasal polarity, as well as adhesive integrity (Hatta and Takeichi, 1986; Nakagawa and Takeichi, 1995). N-cad is down-regulated from the dorsal aspect of the neural tube as neural crest cells emigrate from the neuroepithelium (Bronner-Fraser et al., 1992; Nakagawa and Takeichi, 1998; Cheung et al., 2005; Shoal et al., 2007). Cadherin 6B (Cad6B), on the other hand, is initially expressed in the dorsal neural folds but then repressed by the transcription factor Snail2 (Inoue et al., 1997; Nakagawa and Takeichi, 1998; Taneyhill et al., 2007; Strobl-Mazzulla and Bronner, 2012). The expression pattern of Cad6B suggests a role in formation and/or maintenance of the presumptive neural crest domain within the neural folds (Nakagawa and Takeichi, 1995). Cad6B is indeed essential for the de-epithelialization process of premigratory neural crest cells at the onset of EMT (Park and Gumbiner, 2010), and yet its down-regulation is required for completion of EMT and proper migration of neural crest cells (Coles et al., 2007). E-cadherin (E-cad) is expressed in the neuroepithelium throughout early neural development from the neural plate stage up to Hamburger and Hamilton (HH)
stage 11 in the chick (Dady et al., 2012). Although multiple studies have focused on the mechanisms of neural crest specification in the dorsal neural folds (Kerosuo and Bronner-Fraser, 2012), the link between specification and the adhesive integrity of the neural epithelium has not been clear.

The Myc interacting zinc finger protein-1 (Miz1, also called ZBTB17) is best known for its role in transcriptional activation of cell cycle inhibitors (p15\textsuperscript{INK4A}, p21\textsuperscript{WAF1}) in cancer-associated studies (Peukert et al., 1997; Staller et al., 2001; Adhikary and Eilers, 2005; Phan et al., 2005; Weber et al., 2008). In addition, it also has been implicated in the regulation of cell adhesion by transcriptional activation of several adhesion-related genes in vitro (Gebhardt et al., 2006; Herkert et al., 2010). However, little is known about the expression pattern of Miz1 or molecular mechanisms underlying its role during embryonic development in vivo. In this study, we explore the possibility that Miz1 may represent a molecular link between neural crest specification at the neural plate border region and the adhesive changes that occur in the neuroepithelium. The results show that Miz1 is specifically expressed in the neural ectoderm and neural crest and that its loss of function leads to severe defects in survival of the neural crest precursor pool, alterations in the adhesive complex throughout the neuroepithelium, and delay/decrease in emigration.

RESULTS

Miz1 mRNA is expressed in the neural plate and migratory neural crest

As a first step in exploring the possible function of Miz1, we examined its expression pattern in the early chick embryo during stages of neural crest formation and onset of emigration. Initiation of Miz1 mRNA expression begins in the neural plate in the gastrulating embryo at HH5, after neural induction, as well as after the induction of the neural plate border region (Basch et al., 2006; Stuhlmiller and Garcia-Castro, 2012). It is highly expressed throughout the neural plate at HH5–7 and later enhanced in the closing neural folds during neurulation (HH7–8). Expression is strongest in the dorsalmost portion of the rising neural folds at HH8, as seen in transverse section (HH8\textsuperscript{′}). After neural tube closure at cranial levels (late HH8), Miz1 continues to be expressed throughout the neural tube, with the exception of the ventralmost aspect. Emigrating neural crest cells also express Miz1, and immunostaining with HNK-1 antibody confirms that the Miz1-positive cells are migratory neural crest cells (Figure 1).

Morpholino knockdown of Miz1 affects neural crest induction at the neural plate border

Induction of the neural crest-forming region begins in the HH3 gastrula (Basch et al., 2006; Stuhlmiller and Garcia-Castro, 2012), whereas expression of the neural plate border genes Msx-1 and Pax7 can be detected by in situ hybridization by late HH4 and is continued in the elevating neural folds throughout stages HH5–8 (Khudyakov and Bronner-Fraser, 2009). This is followed by expression of neural crest specifier genes like FoxD3 and Sox10 in the fully committed premigratory neural crest in the closing dorsal neural tube and emigrating cells at HH8–9 (Sauka-Spengler and Bronner-Fraser, 2008; Khudyakov and Bronner-Fraser, 2009). Because Miz1 is expressed in the neural plate during late gastrula stages beginning at HH5, we first asked whether its knockdown affects gene expression in the neural plate border region. The results reveal a decrease in expression of Msx-1 and Pax7 transcripts in the neural folds at HH7 (Figure 2, A and B), observed in 81% of the embryos (n = 16) after morpholino-mediated loss of Miz1 (Figure 2D). In contrast, the neural progenitor marker Sox2 expression does not appear to be diminished on the Miz1-treated versus control side (n = 12/12; Figure 2, F and G), although the neural tube itself appears a bit thinner.

To control for possible nonspecific effects of morpholino knockdown, we performed rescue experiments by coelectroporating Miz1 morpholino together with full-length chick Miz1, cloned into the chicken expression vector pciH2BRFP (Figure 2C). The results reveal a rescue of the expression of Msx-1 at the neural plate border at HH7 in the majority of electroporated embryos (n = 9; 67%; Figure 2E), confirming specificity of the effect.

Loss and gain of Miz1 affect neural crest marker expression at the onset of emigration

Because Miz1 is continuously expressed during neurulation and neural crest emigration, we asked whether its knockdown also affects neural crest gene expression within the dorsal neural tube. The results show that morpholino-mediated loss of Miz1 results in a decrease in the expression of the neural crest–specifier genes FoxD3, Sox10, and Ets-1 in the dorsal neuroepithelium before neural crest emigration. There are fewer FoxD3-expressing neural crest precursors, and the dorsal neural tube appears thinner on the morpholino (MO)-treated side compared with the contralateral control side (Figure 3, A–C). The diminution of neural crest markers was seen in 84% of the embryos after in situ hybridization (n = 32; Figure 3E). These findings were verified using quantitative PCR (qPCR) of neural tubes from HH8 embryos, which show significantly decreased expression of relevant neural crest markers (Figure 3F).
We next examined embryos at HH10–12, using Sox10 in situ hybridization, as well as HNK immunostaining, to determine whether the effects of loss or gain of Miz1 persisted or there was recovery with time. The results suggest that the delay of neural crest cell emigration after Miz1 loss has long-lasting effects on neural crest migration, with fewer migrating neural crest cells even at HH12 (Figure 4, A and B). Of interest, we found that the effect on neural crest emigration may be a later and separate effect from the decrease in the size of the presumptive neural crest domain in the neural folds at HH7. We show that Miz1 knockdown at HH8, well after neural crest specification at the neural plate border is complete, still results in a diminution in the number of migrating neural crest cells at HH11 (Figure 4C). Conversely, overexpression of Miz1 induced premature emigration of neural crest cells from the neural tube, as well as an overall increase in the number of neural crest cells at these later stages (Figures 4, D and E).

**Loss of Miz1 induces cell death and decreased proliferation**

The observed thinning of the neural tube and reduction in size of the presumptive neural crest domain after loss of Miz1 (Figures 2, A′ and G′, and 3A′) led us to ask whether there might be alterations in cell survival and/or proliferation. The results show that loss of Miz1 induced a dramatic increase in apoptosis at HH7 (n = 6/6) as assayed by caspase immunostaining. This was further quantified in transverse sections, revealing a fivefold increase in cell death in the neural plate (NP; average 4.97, SEM 0.82, n = 3) and a sixfold increase in the neural crest domain (NCD) at the neural plate border (average 5.88, SEM 2.19, n = 3) when compared with the contralateral side of the same embryo injected with control MO and normalized to 1 (Figure 5A). We also detected a decrease in the rate of proliferation on the Miz1-morphant side in the neural plate and neural crest domain of 0.6-fold compared with the control MO side normalized to 1 (Figure 5B; NP average 0.67, SEM 0.055; NCD average 0.63, SEM 0.043, n = 3).

**Miz1 regulates cadherins in the developing neural tube**

Miz1 expression has been associated with regulation of adhesion-related molecules in the epidermis (Gebhardt et al., 2006). To test whether a similar situation is in play during neural crest development, we investigated whether knockdown of Miz1 affects cell adhesion, by analyzing expression of three cadherins, N-cad, E-cad, and Cad68, present in the developing neural tube. Immunostaining revealed a reduction of N-cad on the Miz1-morphant side throughout the neural plate at HH7 (n = 4/5) and the closing neural tube at HH8 (n = 3/4; Figure 6A). Similarly, expression Cad68 was reduced on the Miz1-morphant neural folds at HH7 (n = 6/6), as well as in the dorsal neural tube at HH8 (n = 3/3; Figure 6B). Conversely, we detected an increase of E-cad in the neural plate at HH7 (n = 5/6), as well as in the dorsal neural tube at HH8 (n = 3/4) on the Miz1-knockdown side.
compared with the contralateral control MO-injected side (Figure 6C). Double immunostaining confirmed that E-cad expression indeed is increased concomitant with a decrease in Cad6B in the dorsal neural tube at HH8 (Figure 6D).

Overexpression of Miz1 resulted in loss of N-cad protein on the Miz1 side at HH7 and HH8 (n = 5/5; Figure 6E) and an increase in Cad6B expression compared with the contralateral side that was injected with the empty control vector (n = 4/5; Figure 6F). We failed to detect a consistent change in the expression of E-cad after Miz1 overexpression, with the exception of a slight decrease in a portion of the neural tube visible in cross section (n = 5).

To analyze changes caused by Miz1 knockdown at the mRNA level, we performed qPCR analysis of HH8 embryos. The results revealed a significant reduction in the levels of Cad6B transcripts on the Miz1 morpholino-treated side (t test, p = 0.018), consistent with the immunostaining results. In contrast, there was no statistically significant change in the transcript levels of either N-cad or E-cad (Figure 6G). To further study effects on adhesion caused by Miz1 knockdown, we examined mRNA levels in the dorsal neural tubes of HH8 embryos of other adhesion molecules, α1-catenin or β1-integrin, previously shown to be Miz1 targets in other cell types (Gebhardt et al., 2006). Our results show a statistically significant reduction in α1-catenin transcripts on the Miz1 morpholino-treated side (Figure 6G, t test, p = 0.041), whereas levels of β1-integrin were unchanged compared with the control side (average 1.04, SEM = 0.081).

**DISCUSSION**

Here we show that Miz1 is involved in the regulation of survival of developing neural tube cells and formation of apical adhesion complexes therein (Figures 5 and 6). Analysis of the expression of neural crest marker genes in the neural plate border and dorsal neural tube revealed a marked decrease in the size of the presumptive neural crest domain (Figure 2) at HH7. Of interest, neural induction appears to proceed normally after loss of Miz1, since no changes were noted in expression of the neural marker Sox2 within the neural stem cell precursor domain in the neural plate, although there was a general thinning of the neuroepithelium consistent with the cell survival defect on both the neural crest and neural cells (Figure 2, F and G). In contrast, we noted a dramatic loss of neural plate border and neural crest-specifier genes, reflecting a diminution in the size of the neural crest domain, after knockdown of Miz1 (Figures 2 and 3). The reduction in number of FoxD3- and Sox10-expressing premigratory neural crest cells led to a delay in emigration and decrease in total number of migrating neural crest cells (Figure 4). Thus loss of neural crest cells after Miz1 knockdown may at least partially be caused by a decrease in the neural crest precursor pool caused by a critical role for Miz1 in cell survival (Figure 5). We also detected substantial changes in the adhesive properties of the entire neuroepithelium (Figure 6). The fact that levels of Cad6B transcripts and protein were significantly changed after gain or loss of Miz1 may suggest a connection between Miz1 and Cad6B that specifically affects the formation/maintenance of the neural crest domain and the transcription of...
FIGURE 4: Miz is required for the proper migration of neural crest cells. (A) In situ hybridization of Sox10 reveals that the depletion of neural crest cells caused by Miz1 knockdown at HH4 does not recover at later stages (HH12). (B) Embryos treated with CoMO on one side have normal Sox10 expression. (A’, A”) Transverse sections of Miz1 MO–treated embryos (left) show a significant loss of migrating neural crest cells compared with the contralateral control side as assayed by Sox10 expression and HNK immunostaining. (C) Electroporation of the Miz1 MO at HH8, into the closing neural tube well after neural crest specification is complete, still causes delayed neural crest emigration, as seen by Sox10 in situ hybridization of a transverse midbrain section from a HH10 embryo. (D) Overexpression of Miz1 at HH4 causes an increase in the numbers of migratory neural crest cells, as shown by Sox10 in situ hybridization in a late HH9 embryo. The equidistant purple bars on both sides demonstrate the difference between the longer lateral migration length seen on the Miz1 overexpression side vs. the control side treated with the empty pCAG vector. (E, E’, and E”) Overexpression of Miz1 increases the overall amount of the migratory neural crest cells, as shown by Sox10 in situ hybridization and HNK immunostaining in a late HH10 embryo. The green asterisk marks the Miz1 MO–treated and the red asterisk marks the Miz1 overexpression side.

FIGURE 5: Loss of Miz1 induces apoptosis and defects proliferation in the whole neuroepithelium at HH7. (A) Immunostaining of caspase 3 shows increased apoptosis on the morphant side visible in the whole mount and transverse section of an HH7 embryo. When quantified, Miz1 Mo causes a fivefold increase in apoptosis in the neural plate (NP) and an almost sixfold increase in the neural crest domain (NCD) in the lateral neural folds as compared with the control MO–injected side, which was normalized to 1. (B) Immunostaining of the mitosis marker phospho–histone H3 shows decreased proliferation on the Miz1 MO side. The proliferation in the neural plate, as well as in the neural crest domain, was only 0.6-fold vs. the contralateral side, which was normalized to 1.
they comprise adherens junctions that control the apicobasal polarity of epithelial cells by tight attachment to the actin cytoskeleton. α-Catenins are linkers between the cadherin/β-catenin (or plagoglobin) complexes and actin-binding proteins such as vinculin, talin, and α-actinin (Taneyhill, 2008). We observed a decrease in the expression of α1-catenin mRNA as well as Cad6B in the HH8 embryos after Miz1 knockdown, whereas the mRNA levels of Ncad and Ecad were not changed (Figure 6). In line with our results, in vitro overexpression of Cad6B in L cells induces increased expression of α1-catenin (Nakagawa and Takeichi, 1995). Similarly, overexpression of another

![Image](image1.png)

**FIGURE 6**: Miz1 regulates the apical adhesion complexes in the entire neuroepithelium. (A) Loss of Miz1 induces a loss of N-cad in the neural folds at HH7 and the neural tube at HH8, as shown by immunostaining. (B) Loss of Miz1 induces a loss of Cad6B in the lateral neural crest domain of the neural folds at HH7 and the dorsal neural tube at HH8, as shown by immunostaining. (C) Loss of Miz1 induces an increase of E-cad expression in the entire neural folds at HH7 and in the dorsal neural tube at HH8 (white arrow), as shown by immunostaining. (D) Double immunostaining shows decreased expression of Cad6B in the dorsal neural fold (white arrow) with concomitant increase in E-cad. (E) Overexpression of Miz1-H2BRFP decreases N-cad protein expression in the apical side of the neural folds. (F) Overexpression of Miz1-H2BRFP increases the expression of Cad6B in the neural crest domain in the lateral neural folds. (G) qPCR data show a decrease in the mRNA levels of Cad6B and a previously shown Miz1 target, α1-catenin, which binds to cadherins and stabilizes adherens junctions. No statistically different change in the N-cadherin or E-cadherin transcript levels was observed. The green asterisk marks the Miz1 MO-treated and the red asterisk marks the Miz1 overexpression side.

the neural crest markers within. Finally, the finding that loss of Miz1 at the time of neural tube closure still causes defects in neural crest emigration (Figure 4C) suggests that Miz1 at least partially alters the EMT process independent of its effects on neural crest survival and adhesion at the neural plate border. It is interesting to note that as the neural folds elevate, the intensity of Miz1 expression increases at the dorsal tips of the closing neural tube (Figure 1, HH8). This further suggests a potentially important function at this time that may be distinct from its earlier role.

In cancer cells and keratinocyte cultures, Miz1 regulates the expression of several adhesion-related genes and is required for maintenance of the proper polarized structure of the epidermis (Gebhardt et al., 2006; Herkert et al., 2010). Similarly, we noted significant changes in expression and distribution of cadherins in the developing neuroepithelium (Figure 6). Loss of Miz1 caused a reduction in N-cad protein and a concomitant increase in E-cad levels in the apical adhesion complex in the neural plate. This N-cad to E-cad switch is similar to that seen after Sip1 knockdown (Rogers et al., 2013). Moreover, we found that Cad6B was decreased in the prospective “neural crest domain” at HH7 due to a loss of Miz1 and increased after its overexpression. Expression of Cad6B in the developing neural tube is essential for proper neural crest development (Nakagawa and Takeichi, 1995; Park and Gumbiner, 2010). Overall, either loss or gain of Miz1 alters the balance of the three cadherins expressed in the neuroepithelium. Together with the observed alterations in neural crest marker genes, this suggests that their correct balance is key for proper formation of the neural crest domain and subsequent emigration from the neural tube.

Another protein involved in cell–cell adhesion and previously associated with Miz1 (Gebhardt et al., 2006) is α1-catenin (also called αE-catenin). It is expressed apically throughout the neuroepithelium, as well as in the migrating neural crest cells (Nakagawa and Takeichi, 1995), and thus shares an overlapping expression pattern with Miz1. Cadherins form complexes with intracellular catenins and together they comprise adherens junctions that control the apicobasal polarity of epithelial cells by tight attachment to the actin cytoskeleton. α-Catenins are linkers between the cadherin/β-catenin (or plagoglobin) complexes and actin-binding proteins such as vinculin, talin, and α-actinin (Taneyhill, 2008). We observed a decrease in the expression of α1-catenin mRNA as well as Cad6B in the HH8 embryos after Miz1 knockdown, whereas the mRNA levels of Ncad and Ecad were not changed (Figure 6). In line with our results, in vitro overexpression of Cad6B in L cells induces increased expression of α1-catenin (Nakagawa and Takeichi, 1995). Similarly, overexpression of another
neurospheres (Kerosuo et al., 2008). For example, a Miz1/c-Myb complex regulates neural stem cell self-renewal in embryonic tissues and massive apoptosis, as well as a lack of p57kip, in the whole embryo (Adhikary et al., 2003). However, we speculate that this function is separate from the regulation of the adhesion complexes, since the apoptosis and proliferation defect was seen throughout the neural plate, but loss of Miz1 selectively affected neural crest marker expression. Whereas neural crest marker expression was compromised, neural specification proceeded normally, as shown by the normal size of the Sox2-expressing neural stem cell domain (Figure 2G). This suggests that Miz1 selectively affects neural crest precursors. This may not be surprising, given that Miz1 comes on at HH5, well after neural induction is complete, suggesting that it may have a specific role in the survival of developing neural crest cells. Intriguingly, sumoylation of Miz1 in cancer cells by interaction with Arf disrupts the transcriptional activator complex formed by Miz1 and nucleophosmin, which causes repression of multiple genes involved in cell adhesion and signal transduction. This results in weakening of cell–cell and cell–matrix interactions and induces apoptosis (Herkert et al., 2010). Thus the adhesion changes caused by loss of Miz1 may be a primary defect that secondarily induces apoptosis in the neuroepithelium.

Miz1 regulates differentiation in the epidermal stem cell niche in vivo by coordinating the exit of epidermal stem cells from their niche and thus regulating the balance of keratinocyte differentiation in a manner that involves both adhesion-related changes and cell cycle regulation (Gebhardt et al., 2006). In addition to adhesion changes, we also noted decreased proliferation on the Miz1-morphant side of the neural plate (Figure 5B). Miz1 is known to regulate transcription levels of the cell cycle inhibitors—for example, p15ink4a and p21waf1 (Adhikary and Eilers, 2005)—which may explain what occurs in the neuroepithelium. Another possibility is that the extensive apoptosis we observe may be secondary due to the compromised proliferation capacity of the neuroepithelium. Finally, it is intriguing to speculate that Miz1 may act together with N-Myc in the developing neural tube. Consistent with this possibility, N-Myc has been shown to interact with Miz1 in cell lines of the neural crest–derived cancer neuroblastoma (Akter et al., 2011). Alternatively, Miz1 may function as an independent transcriptional activator at these stages or form a repressive complex with another partner. In addition to c-Myb, Miz1 has been shown to interact with TopBP1, BCL6, Zbtb4, and p14Arf (Adhikary and Eilers, 2005; Phan et al., 2005; Liu et al., 2006; Weber et al., 2008; Herkert et al., 2010).

Similar to activation of Miz1 in the epidermis by transforming growth factor-β (TGFβ) signaling (Gebhardt et al., 2006, 2007), TGFβ family members bone morphogenetic protein 4/7 (BMP4/7) are expressed at the neural plate border and critical for neural crest specification in this region. Later BMP4 is expressed in the tips of the dorsal neural folds at early HH8 and is a major neural crest inducer (Liem et al., 1995). Given that BMP4 activates expression of Cad6B and RhoB and its expression is crucial for neural crest EMT (Liu and Jessell, 1998; Park and Gumbiner, 2010), Miz1 may represent a transcriptional link between BMP signaling and effects on adhesion molecules and cell survival in the developing neural tube.

In addition to its role as a transcription factor, there is some evidence that Miz1 may play other intracellular roles. In a hematocyte cell line, for example, Miz1 is largely cytoplasmic and associated with microtubules and cholesterol metabolism; it reacts to cytoskeletal changes and translocates into the nucleus only upon microtubule depolymerization (Ziegelbauer et al., 2001). Thus it is possible that the effects of Miz1 loss observed on cell adhesion molecules occur by intracellular rather than transcriptional mechanisms.

In summary, our results reveal for the first time a critical role for the transcriptional regulator Miz1 during neural crest development. Miz1 affects gene expression at two levels of the neural crest gene regulatory network—at the neural plate border/neural folds and in the emigrating bona fide neural crest cells. This leads to long-lasting alterations in migrating neural crest cell numbers. Our results reveal changes in adhesion molecules and survival of the cells throughout the entire neuroepithelium that ultimately affect the ability of the neural crest cells to properly form and emigrate from the neural tube.

MATERIALS AND METHODS

In situ hybridization

Embryos were fixed with 4% paraformaldehyde, washed with phosphate-buffered saline/0.1% Tween, dehydrated in MeOH, and stored at −20°C. The avian Miz1 probe was made by using chEST672h7 (www.chick.manchester.ac.uk), and the Sox10, FoxD3, Msx-1, and Sox2 probes were made by cloning respective genes to DNA vectors from reverse transcription (RT) PCR products made by using chicken whole-embryo cDNA as template. Whole-mount in situ hybridization was performed as described (Acloque et al., 2008). The digoxigenin-conjugated RNA probes were visualized by using anti-dig-AP antibody (1:2000; 11093274910; Roche Diagnostics, Mannheim, Germany) and 4-nitro blue tetrazolium chloride/5-bromo-4-chloro-3’-indolylphosphate p-toluidine (11383213001 and 11383221001; Roche Diagnostics). Embryos were sectioned at 12–20 μm.

Morpholino knockdown/overexpression and electroporation of chicken embryos

Fluorescein isothiocyanate (FITC)–conjugated morpholinos were purchased from Gene Tools (Philomath, OR). The Miz1 translation-blocking morpholino (AACTGGGACAGCTGCTGCAACAGG) was targeted to the S’ untranslated region (UTR) 78–54 base pairs before
the ATG, and a control morpholino was designed to ensure lack of non-specific effects from electroporation (CTCGCATGAAGAAAACACGGGAGCACACA). The MOs were diluted to 1.5 mM concentration and electroporated together with an empty pGAG vector as carrier DNA (1 μg/μl). All the electroporations presented in this study were performed at gastrula stage (HH4) unless otherwise stated. The Miz1 morpholino/vector DNA was either injected on one side of the embryo alone with the non-electroporated side serving as an internal control or as two-sided injections with control morpholino on the contralateral side. The electroporation was carried out as previously described (Sauka-Spengler and Barendbaum, 2008). Briefly, the chicken embryos were collected on Whatman filter papers and electroporated at HH4 by using 5.3 V and five pulses (50 mA/100 mA) and incubated on individual Petri dishes (Falcon 1008 35 × 10 mm) in thin albumin until they reached the desired stage. The HH8 electroporation was performed by injecting the MO into the neural tube of four- to six-somite embryos and electroporated by using 20 V and five pulses (50 mA/100 mA).

qPCR
mRNA was isolated from HH8 embryos by using the Ambion RNAqueous-Micro Kit (Life Technologies, Grand Island, NY) by individually collecting neural tube halves from the Miz1 MO–treated and contralateral control MO–treated sides, respectively. The control embryos were treated with control MO on both sides. The results are shown as the relative expression fold of the treated side versus the control side and were analyzed using the ∆∆CT method (Livak and Schmittgen, 2001). Results from three to eight individual embryos were pooled and are shown as average values. The error bars represent SEM. Primers were designed to target an exon–exon boundary, and their amplification rate was verified as linear within the margin of ±10% amplification rate change between points in the logaritically diluted cDNA standard curve. The following primers were used: GapdhFwd, ATCTACTTCTCCACCCCGT; GapdhRev, AGCCACA-CCTCTCTGATGAG; Sox10Fwd, AGGTTGGAAGAGTTGTCCCC; FoxD3Fwd, TCTGCTGGCTGTTCATCAGCA; NcadFwd, ACCAGGAAGAGTTCATCAGCAAC; Cad6BFwd, ACCAGAACAGCACACAG; Cad6BRev, GAGTATGTTGCTGCTTACA-CCT; Cdc42Fwd, CAGAGACCTTACGTTACG; and Cdc42Rev, CATCCGGCGTCCTACCTG.

Immunostaining
The following primary antibodies against the indicated epitopes were used: Pax7, HNK-1 (3HS), Ncad (MNCD2), and Cad6β from the Developmental Studies Hybridoma Bank (University of Iowa, Iowa City, IA) were used in a 1:10 dilution, as well as Ecad (0.25 μg/ml; 610181; BD Biosciences, San Jose, CA), the mitosis marker phos- pho–histone H3 for detection of proliferation (1:500; 06-570; Upstate, Millipore, Billerica, MA), and caspase 3 for detection of apoptosis (1:300; AB385; R&D Systems, Minneapolis, MN). Embryos were fixed in 4% paraformaldehyde without saline for 15 min at room temperature, washed in 0.5 M Tris-buffered saline (TBS) buffer, pH 7.5, and blocked in TBS with 10 mM CaCl2, 1.5M NaCl, 0.5% Triton, 1% dimethyl sulfoxide, and 10% goat serum. The antibodies were diluted in the blocking buffer, and whole-mount embryos were stained by incubation for 2 d in +4°C (or on sections overnight in some cases) and by using the respective Alexa secondary antibodies (Life Technologies).

Rescue experiments and overexpression
The avian Miz1 coding sequence was cloned by using the predicted Ensemble sequence ENSGALT00000039813 to make the forward primer (TTTT TTCGAA ttaagctgccagcagc) with a BstBI site and the expressed sequence tag sequence ENSGALT0000005815 for the reverse primer (TTTT GCTAGC tttaccgctttcacaag) added with an Nhel site. The full-length PCR product was cloned into the avian expression vector PsplH2RFP with the pGAG promoter and the nuclear red florescent protein (RFP) reporter gene separated by an internal ribosome entry site. The rescues were performed by injecting Miz1 morpholino together with the full-length chicken Miz1 H2RFP (1 μg/μl) on one side of the embryo with control morpholino together with the empty pGAG vector on the contralateral side. The Miz1 expression sequence starts from the coding region (ATG) and is not affected by the Miz1 morpholino, which targets the 5’ UTR upstream of the coding sequence. For overexpression, 3 μg/μl Miz1 H2RFP in EB buffer (10mM Tris-HCl, pH 8.5) was injected and electroporated on one side of the embryo and while introducing 3 μg/μl of the empty pGAG vector into the contralateral side. All electroporations were performed at HH4.

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