An apertureless near-field microscope for fluorescence imaging

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We describe an apertureless near field microscope for imaging fluorescent samples. Optical contrast is generated by exploiting fluorescent quenching near a metallized atomic force microscope tip. This microscope has been used to image fluorescent latex beads with subdiffraction limit resolution. The use of fluorescence allows us to prove that the contrast mechanism is indeed spectroscopic in origin.

The resolving ability of optical microscopy with far field optics is limited by the diffraction of light. Near field scanning optical microscopy allows one to take optical images with resolution below the diffraction limit. To surpass this limit, apertureless near field scanning microscopes (ANSOM) were proposed and demonstrated, with apparent resolutions as low as 1 nm. ANSOM techniques involve the use of an oscillating sharp probe, which is scanned over the sample. The probe perturbs an incident laser beam, either by introducing phase shifts in the electric field or by periodic occlusion of the sample. Ac detection techniques are used to discriminate light scattered by near field interactions from the far field contribution. However, for these single wavelength microscopes the contrast mechanism is still not completely understood and on occasion the images are contaminated by topographical artifacts. Recently a fluorescence ANSOM microscope (FANSOM) was demonstrated which uses the principle of a two photon excitation and electric field enhancement near the tip.

Here we report the results of a new FANSOM microscope designed to image fluorescent samples with single photon excitation. We exploit the fact that fluorescent molecules transfer energy nonradiatively to proximate metal or semiconductor surfaces. This quenching effect has been studied for many years and may limit the effectiveness of microscopes using electric field enhancement as a contrast mechanism. The ability to use single photon excitation allows greater flexibility in the choice of laser source and reduces the peak power through the objective and on the sample. The use of fluorescence facilitates interpretation of the images and provides compelling evidence that the contrast mechanism is indeed optical in origin. Specific spectroscopic effects such as photobleaching are observed, showing that there is no height contamination in the optical image. As with ANSOM, the resolution is determined by the radius of curvature of the probe. AFM tips with 5 nm radius of curvature are now commercially available, and carbon nanotube AFM tips have been demonstrated. It is thus reasonable to expect that the ultimate resolution of FANSOM will be in the 1–5 nm range. There are a number of important possible applications of FANSOM, including imaging of single DNA molecules for optical mapping and fluorescent in situ hybridization, spectroscopy of nanoparticles and quantum well devices, and the possible extension to single molecule Raman imaging.

We used our FANSOM to image fluorescent latex particles with subdiffraction limit resolution and to measure fluorescence photobleaching, proving that the contrast mechanism is due to a near-field effect and is optical in nature. The experimental setup is shown in Fig. 1. It consists of a tapping mode atomic force microscope (AFM) on top of an inverted optical microscope, both homemade. The sample is scanned under the tapping AFM tip (resonant frequency ~250 kHz), while a green HeNe (543.5 nm, Uniphase 1674P) laser beam is focused to a diffraction limited spot on the sample plane.
the tip via a microscope objective. Commercial silicon AFM tips (nanosensors) were coated with a 60 nm layer of gold by thermal vacuum deposition. Photons emitted by fluorescence are collected by the same objective (Olympus Plan Apo Chromat 60×, 1.4 NA) and imaged onto a photon counting avalanche photodiode (EG&G SPCM 100), whose output is in turn processed by a gated photon counter (Stanford Research Systems SR400) to implement a lock-in detection scheme. The gating was triggered by the position of the AFM cantilever such that the counts measured while the tip was closest to the sample were subtracted from the counts measured when the tip was farthest from the sample. This effectively selects a 100 Hz frequency window centered on the 250 kHz resonant frequency of the cantilever and allows discrimination of the far-field emission from the photon suppression due to fluorescence quenching at the near field of the tip. The position of the AFM cantilever was measured on a split photodiode using a standard optical lever scheme. The split photodiode signal was used as part of a feedback loop to maintain constant tapping amplitude, and was also used with a phase locked loop to trigger the photon counter gates.

During image acquisition, the sample was scanned in the xy plane and two signals were collected: the photon counter difference output (optical) signal and the AFM z-feedback (topographical) signal. This allows the simultaneous acquisition of topographical (AFM), and near field optical fluorescence (FANSOM) images. Far-field optical images using the total number of photons per pixel were also taken during a separate scan. Our samples consisted of 60 nm fluorescent latex beads (Nile Red, Interfacial Dynamics Corp.) deposited on an RCA cleaned glass cover slip. The laser beam had a total power of 20 nW focused into a 500 nm diameter spot. We used a set of bandpass filters in front of the avalanche photodiode that selected out the region between 580 and 620 nm, excluding both the 543.5 nm excitation light and the 670 nm diode laser used for the AFM feedback.

Figure 2 shows two sets of images collected from a 1 μm² field of view. Fluorescent beads are seen using AFM, FANSOM, and far field imaging. In one image [Fig. 2(a)], there is also a nonfluorescing contaminant particle that is not visible in the corresponding FANSOM [Fig. 2(b)] or far field [Fig. 2(c)] images. We estimated the resolution of the FANSOM microscope by tak-
ing a vertical line slice through the center of the beads and measuring the full width at half maximum of the peak, finding 95 and 120 nm. Given that the bead has a 60 nm diameter, we can estimate the resolution of the FANSOM to be between roughly 30 nm, well below the diffraction limit of 260 nm. Although this method of determining the resolution does not strictly adhere to the classical Rayleigh criterion, it does show unambiguously that the resolution surpasses the diffraction limit. Control images taken with the tip removed show only the expected far-field image, with no signal in the FANSOM image. Control images taken with the green laser blocked show no FANSOM image, only the normal AFM image.

We verified that the FANSOM image is not contaminated with height information by observing photobleaching of the bead. Repeated images of the same bead showed a gradual reduction in the level of the FANSOM signal, while the AFM image remained unchanged. We also took images with 100 nm beads excited by a 60 nW laser beam. By positioning the AFM tip directly above a single bead, we were able to observe the near-field signal as a function of time. We measured the signal in this case as the height of a peak in the power spectrum of the output of the avalanche photodiode. These approach curves show that the modulation depth of the signal is greater than 50% for 60 nm beads when gold coated tips were used, and increases as the size of the feature is reduced.

In conclusion, we have demonstrated subdiffraction limit near field fluorescence imaging with an apertureless probe. We have verified the near field effect and demonstrated that the images are free from topographical artifacts. In future work, we plan to test the ultimate sensitivity of the FANSOM by imaging single dye molecules and biological samples.

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