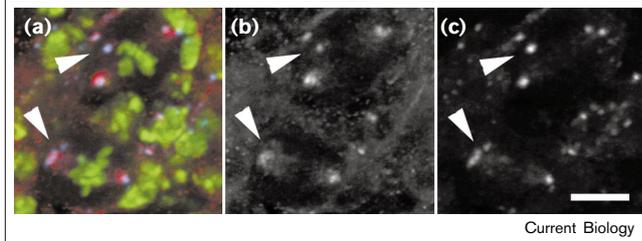


## The SCF ubiquitin ligase protein Slimb regulates centrosome duplication in *Drosophila*

Edward J. Wojcik, David M. Glover and Thomas S. Hays

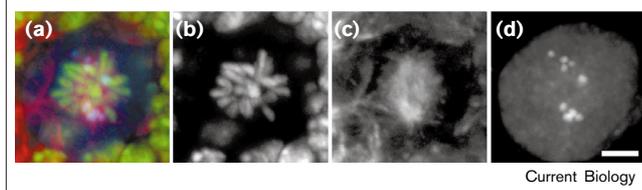
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Figure S1



Off-axis centrosomes in the *slimb<sup>crd</sup>* mutant contain ASP. (a) Adjacent metaphase *slimb<sup>crd</sup>* mutant neuroblasts (arrowheads) triple-labeled for DNA (green),  $\gamma$ -tubulin (blue) and ASP (red). The ASP channel from (a) is shown alone in (b). The lower arrowhead points to the ASP signal at a cluster of off-axis centrosomes; the upper arrowhead points to an off-axis extra centrosome. (c) The  $\gamma$ -tubulin channel from (a). The scale bar represents 5  $\mu$ m.

Figure S2



Polyloid *slimb<sup>crd</sup>* neuroblasts can be seen with bipolar spindles and excess centrosomes. (a) Merged Z-series of an aberrant *slimb<sup>crd</sup>* mutant polyloid neuroblast fluorescently labeled for DNA (green), tubulin (red) and CP190 (blue). (b–d) The DNA, tubulin and centrosome channels, respectively, from (a). The scale bar represents 5  $\mu$ m.

### Supplementary materials and methods

#### Fly stocks

Animals that are trans-heterozygous for *slimb<sup>crd</sup>* and a previously identified recessive lethal *slimb* allele, *slimb<sup>PZ00295</sup>* [S1], failed to survive beyond mid-larval development. Cells in the central nervous system of these larvae displayed a mitotic phenotype indistinguishable from that of *slimb<sup>crd</sup>* hemizygotes (not shown). The *slimb<sup>crd</sup>* homozygotes die at the larval–pupal boundary; animals hemizygous for *slimb<sup>crd</sup>* exhibited lethality at an earlier stage of the third-larval instar, suggesting that the *P* allele is hypomorphic. Furthermore, the *slimb<sup>PZ00295</sup>* mutation was both viable and fertile in combination with the *slimb<sup>crd</sup>* revertant chromosomes. Amorphic *slimb* alleles have previously been shown to result in embryonic lethality [S1].

#### Immunofluorescence and confocal microscopy

Whole-mount larval brains were prepared for immunofluorescence as described [S2]. Triple-labeled specimens were generated by using combinations of the following antibodies: anti- $\gamma$ -tubulin (Sigma, DM1A at 1:200); anti-CP190 (at 1:500; [S3]); anti-CNN (at 1:500 [S4]);

and anti-ASP (at 1:200, [S5]). The fluorescent DNA stains ToPro-3 or Sytox Green (both at 1:2000; Molecular Probes) were interchangeably used as required. Fluorescently conjugated secondary antibodies used were anti-mouse Texas Red conjugate (Jackson ImmunoResearch Labs) and an anti-rabbit Alexa-488 conjugate (Molecular Probes). Z-series data were collected on a Bio Rad MRC 1024 confocal system. Sequential image acquisition prevented bleed-through artifacts from both double- and triple-labeled samples. Projections of Z-series were created with NIH image, and final images adjusted for printing using Adobe PhotoShop.

#### Labeling of S-phase cells in larval brains

We followed the procedure of Mills *et al.* [S6] with the following modifications. Wild-type and mutant brains were isolated intact and carried through the entire protocol as whole mounts. Both wild-type and mutant brains from each replicate experiment were performed simultaneously and in the same buffers. Replicating DNA was labeled *in situ* with Alexa 488 dUTP (Molecular probes). Samples from several independent experiments were analyzed by confocal microscopy as above.

### Supplementary references

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