

## Supporting Information

### Multi-generation production of secondary organic aerosol from toluene photooxidation

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## Methods

We performed experiments to evaluate the contribution of multi-generation oxidation of toluene initiated by OH radicals to SOA and BrC formation in an environmental chamber. Monodispersed seed particles consisting of  $(\text{NH}_4)_2\text{SO}_4$ ,  $\text{NH}_4\text{HSO}_4$ , and  $\text{NaCl}$  were prepared to represent distinct particle acidity and interfacial electric fields and exposed to the oxidation products from OH-toluene reactions. The experiments of toluene oxidation were performed similarly as in our previous studies.<sup>1</sup> Commercially available gaseous  $\text{NH}_3$  (2000 ppm  $\text{NH}_3$  in  $\text{N}_2$ , Coastal Welding Supply Beaumont Texas) was injected into the chamber with a flow of 100 standard cubic centimeters per minute (sccm) for 2 min. 600  $\mu\text{L}$   $\text{H}_2\text{O}_2$  (35 wt%, Sigma-Aldrich) and 30  $\mu\text{L}$  toluene (99.5%, Sigma-Aldrich) were injected into a glass reservoir, which was then flushed into the chamber by a 10 slpm flow from a pure air generator (Aadco 737-11, Aadco Inc.) for 10 min. The gases were then mixed by a fan inside the chamber. Once desired concentrations were established, the black light lamps were turned on to initiate  $\text{H}_2\text{O}_2$  photolysis to generate OH radicals.

Aerosols were produced by a constant output atomizer (Model 3076, TSI), dried to RH of 2% by a Nafion drier (PD-070-18T-12SS, Perma Pure), charged by a  $^{210}\text{Po}$  bipolar charger, and analyzed by an integrated aerosol analytical system. A scanning mobility particle sizer (SMPS) consisting of a differential mobility analyzer (DMA, Model 3081, TSI) and condensation particle counter (CPC, Model 3760A, TSI) was used to select the initial particle size (100 nm) and to continuously measure the particle size growth factor,  $GF = D_p/D_0$ , where  $D_p$  is the diameter after exposure and  $D_0 = 100$  nm is the initial diameter. The initial particle concentration was maintained at  $1.5 \times 10^4 \text{ cm}^{-3}$ , which was measured by the CPC. An Aerosol Particle Mass Analyzer (APM, Model 3600, Kanomax) was used to measure the particle density throughout the experiments. The density for the SOA materials ( $\rho_{\text{SOA}}$ ) is calculated by,

$$\rho_{\text{SOA}} = \frac{\rho GF^3 - \rho_0}{GF^3 - 1}$$

where  $\rho$  and  $GF$  are measured particle density ( $\text{g cm}^{-3}$ ) and growth factor at  $t$  (min), respectively.  $\rho_0$  is the density of  $(\text{NH}_4)_2\text{SO}_4$  ( $1.77 \text{ g cm}^{-3}$ ),  $\text{NH}_4\text{HSO}_4$  ( $1.78 \text{ g cm}^{-3}$ ), or  $\text{NaCl}$  ( $2.16 \text{ g cm}^{-3}$ ). The uncertainty in the density measurements was estimated to be  $\pm 0.03 \text{ g cm}^{-3}$ .

To measure the optical properties of exposed particles, a 1-srpm flow from the chamber was diluted by 4-srpm  $\text{N}_2$  and introduced into a commercial integrating nephelometer (TSI 3563) and a home-built cavity ring-down spectrometer (CRDS) after 1.5 h of exposure. Light scattering ( $b_{\text{sca}}$ ) and extinction ( $b_{\text{ext}}$ ) at 532 nm were determined by the nephelometer and CRDS, respectively.<sup>2</sup> The nephelometer was calibrated by comparing measured  $b_{\text{sca}}$  with  $b_{\text{ext}}$  from CRDS when measuring ammonium sulfate particles with diameters of 100 nm, 150 nm, 200 nm, 250 nm, and 300 nm. The absorption coefficient ( $b_{\text{abs}}$ ) was determined from  $(b_{\text{ext}} - b_{\text{sca}})$ , and the single scattering albedo ( $SSA$ ) was calculated from  $b_{\text{sca}}/b_{\text{ext}}$ .

An ion drift - chemical ionization mass spectrometer (ID-CIMS)<sup>44</sup> using the hydronium reagent ion ( $\text{H}_3\text{O}^+$ ) was employed to analyze gaseous concentrations with a sample flow rate of 0.5 slpm from the chamber. The sample line was heated to  $70^\circ\text{C}$  to reduce wall loss. Gas-phase concentrations from toluene oxidation (i.e., toluene and oxidation products) were quantified by ID-CIMS.<sup>3</sup> Briefly, the concentration of species A is determined by,

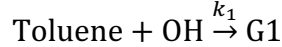
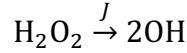
$$[A] = \frac{S_A}{k_{PTR} \Delta t S_{RI}}$$

where  $S_A$  and  $S_{RI}$  are the mass spectrum intensities for A and reagent ions, respectively.  $k_{PTR}$  is the proton-transfer rate constant between A and  $H_3O^+$ .<sup>4</sup>  $\Delta t = \frac{l}{U_i}$  is the retention time of ions, which is determined by the length of the drift tube  $l = 8$  cm and the ion drift velocity  $U_i$ . The ion drift velocity  $U_i$  is determined from:

$$U_i = u_0 \left( \frac{760 \text{ torr}}{p} \right) \left( \frac{T}{273 \text{ K}} \right) E$$

where  $u_0 = 2 \text{ cm}^2 \text{ V}^{-1} \text{ s}^{-1}$  is the reduced ionic mobility,<sup>5</sup>  $p = 5.0$  torr,  $T = 298$  K, and  $E = 37.5 \text{ V cm}^{-1}$  is the voltage gradient in the drift tube. Alternatively, the concentrations of toluene, glyoxal, and methylglyoxal were also calibrated by preparing their vapors with known mixing ratios in  $N_2$ .<sup>6</sup> In our CIMS configuration, formation of cluster ions was effectively suppressed, since a uniform electrical field in the drift tube decomposed weakly-bonded cluster ions. The detection limit (defined as 3 times of the ratio of signal to noise) for the oxidation products from toluene-OH reactions was estimated to be on the order of 50 ppt by the ID-CIMS.

In our experiments, the OH concentration,  $[OH]$ , was estimated using two methods. First, we considered the steady state from the reactions,



where  $J = 3.3 \times 10^{-5} \text{ s}^{-1}$  is the photolysis rate for  $H_2O_2$ ,<sup>6</sup> and  $k_1 = 5.5 \times 10^{-12} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$  is reaction rate between OH and toluene.<sup>7</sup> The steady-state OH concentration is expressed by,

$$[OH]_{ss} = \frac{J[H_2O_2]}{k_1[\text{Toluene}]}$$

where  $[H_2O_2]$  and  $[\text{Toluene}]$ , respectively, are the average concentrations of  $H_2O_2$  and toluene corrected for wall-loss. Alternatively, the concentration of OH was estimated from exponential fitting of the measured decay of toluene concentration,  $[\text{toluene}] = [\text{toluene}]_o e^{-k_2 t}$ ,

$$[OH] = \frac{k_2}{k_1}$$

where  $[\text{toluene}]_o$  is the initial toluene concentration and  $k_2$  is the fitted pseudo first-order rate constant for the toluene-OH reaction. The calculated concentrations of  $[OH]_{ss}$  ( $3 \times 10^6 \text{ molecules cm}^{-3}$ ) agreed within 90% of that derived from exponential fitting ( $2.7 \times 10^6 \text{ molecules cm}^{-3}$ ).

While the oxidation products from OH-toluene oxidations were directly detected by ID-CIMS, we also estimated loss of vapors to the chamber wall by the first-order wall-loss coefficient  $k_w$  ( $\text{s}^{-1}$ ) by considering gas-phase transport within the chamber according to Zhang *et al.*:<sup>8</sup>

$$k_w = \left( \frac{A}{V} \right) \frac{\alpha_w \bar{c}}{4 + \frac{\pi}{2} \left( \frac{\alpha_w \bar{c}}{\sqrt{k_e D g}} \right)}$$

where  $A/V$  is the surface to volume ratio of the chamber (equal to  $6/L$  for a square chamber with  $L = 1$  m),  $\alpha_w$  is the mass accommodation coefficient of vapors onto Teflon chamber walls at RH = 90%,  $\bar{c} = \sqrt{\frac{8k_B T}{m}}$  is the mean thermal speed of the molecules, and  $k_e$  is the coefficient of eddy diffusion.  $k_e$  is estimated to be 0.5 from the loss rate of particles,<sup>9</sup>

$$k_e = \frac{(\frac{\pi L \beta_0}{12})^2}{D_b}$$

where  $\beta_0$  is the fractional loss rate of particles,

$$\beta_0 = \frac{dN}{Ndt} = \frac{15000 \text{ cm}^{-3} - 7500 \text{ cm}^{-3}}{15000 \text{ cm}^{-3} \times 7200 \text{ s}} = 1.4 \times 10^{-5} \text{ s}^{-1}$$

$D_b = 6.75 \times 10^{-10} \text{ m}^2/\text{s}$  is the Brownian diffusion coefficient for particles.  $D_g$  is the gas-phase diffusion coefficient. For  $\text{NH}_3$ , the mass accommodation coefficient of vapors onto the chamber wall is 0.05.<sup>10</sup> Gas-phase diffusion constant  $D_g$  is  $1.98 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$ . The mean thermal speed  $\bar{c}$  is  $603 \text{ m s}^{-1}$ . The first-order wall-loss coefficient of  $\text{NH}_3$  is calculated to be  $1.2 \times 10^{-2} \text{ s}^{-1}$ .

The average concentration of a gas species (A) is estimated by,

$$\overline{[A]} = \frac{1 - e^{-k_w \Delta t_i}}{k_w \Delta t_i} [A]_0$$

For initial injection of 400 ppb  $\text{NH}_3$ , the average concentration of  $\text{NH}_3$  was estimated to be 19 ppb. The first-order wall-loss rate of  $\text{NO}_x$  was measured to be  $2.5 \times 10^{-3} \text{ h}^{-1}$ , leading to negligible wall-loss.<sup>11</sup>

To analyze the particle-phase chemical composition by thermal desorption - ion drift - chemical ionization mass spectrometer (TD-ID-CIMS),<sup>12</sup> seed particles after 20 min of exposure to photooxidation were collected for 2 hours by a platinum filament in a 2.5 slpm flow from the reaction chamber, and the analytes were evaporated by heating the filament and detected by ID-CIMS using  $\text{H}_3\text{O}^+$  as the reagent ions.

The measured particle growth at  $t = 90$  mins was used to derive the uptake coefficient of methylglyoxal/glyoxal from,

$$\gamma = \frac{4 \times FR \times k_1}{\bar{c} \times S}$$

where  $FR$  is the percent contributions of condensable organic species from toluene-OH oxidation to the particle-phase mass intensity (Table S2).  $k_1$  is first-order uptake rate constant of condensable oxidized organics COO (glyoxal, methylglyoxal,  $\gamma$ -dicarbonyls, organic acids, and LVO), which is calculated from the measured size-growth of seed particles,

$$k_1 = \frac{\pi \times (D_p^3 - D_0^3) \times \rho \times N \times N_A}{6 \times MW \times [\text{COO}] \times \Delta t}$$

where  $D_p$  and  $D_0$  are the final and initial diameters of the particles during the time-period of  $\Delta t = 90$  min, respectively,  $\rho$  is the density of total SOA ( $1.6 \text{ g cm}^{-3}$  at RH = 90% and  $1.45 \text{ g cm}^{-3}$  at RH = 10%),  $N = 15000 \text{ cm}^{-3}$  is the measured particle number concentration,  $N_A$  is Avogadro constant, MW is the molar weight, and  $[\text{COO}]$  is the average gas-phase concentration.  $\bar{c}$  is the mean thermal speed of A.  $S$  is the average surface-area of aerosols ( $\text{cm}^2 \text{ cm}^{-3}$ ) at  $\Delta t = 90$  min,

$$S = \frac{\pi \int_{t=0}^{90 \text{ min}} D^2 \times HGf^2 \times N \times dt}{\Delta t}$$

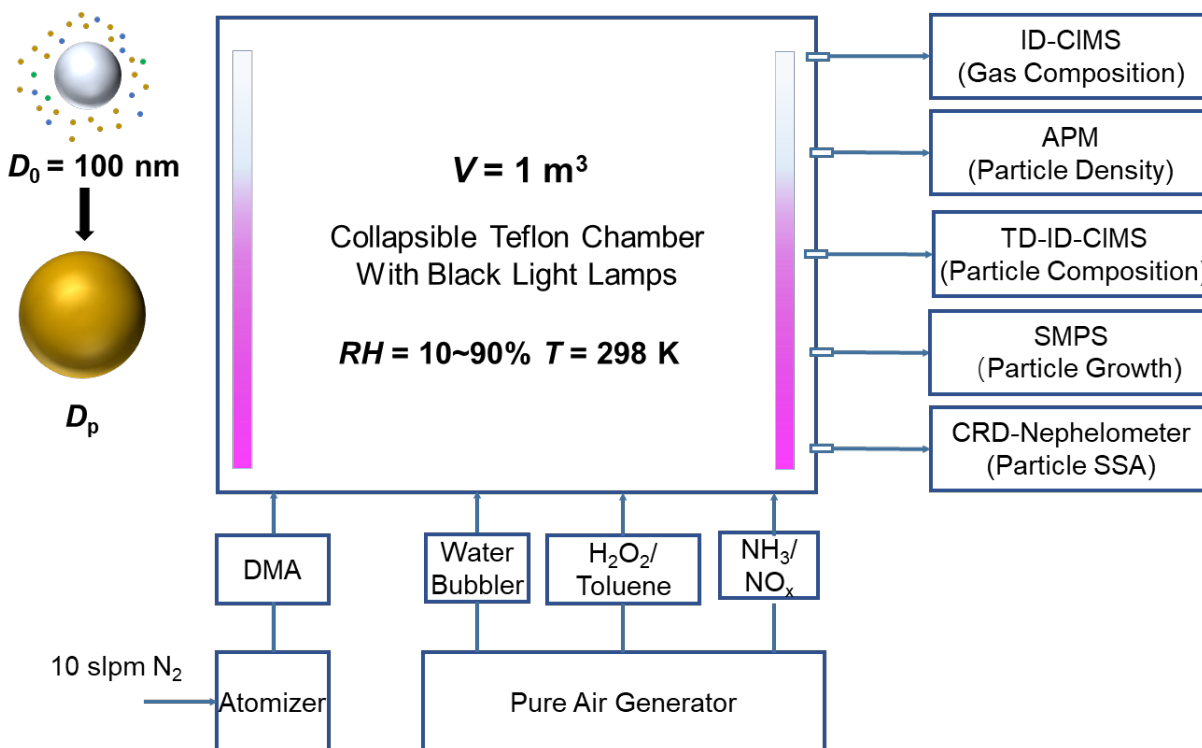
where  $D$  is the time-dependent diameter and  $HGf$  is the hygroscopic growth factor.<sup>13</sup> Results of the derived  $\gamma$  values are summarized in Table 1. The error bars of the uptake coefficients were estimated from the uncertainties for the parameters in Table S6 (i.e., 1  $\sigma$  of at least three repeated measurements for each parameter).

## References

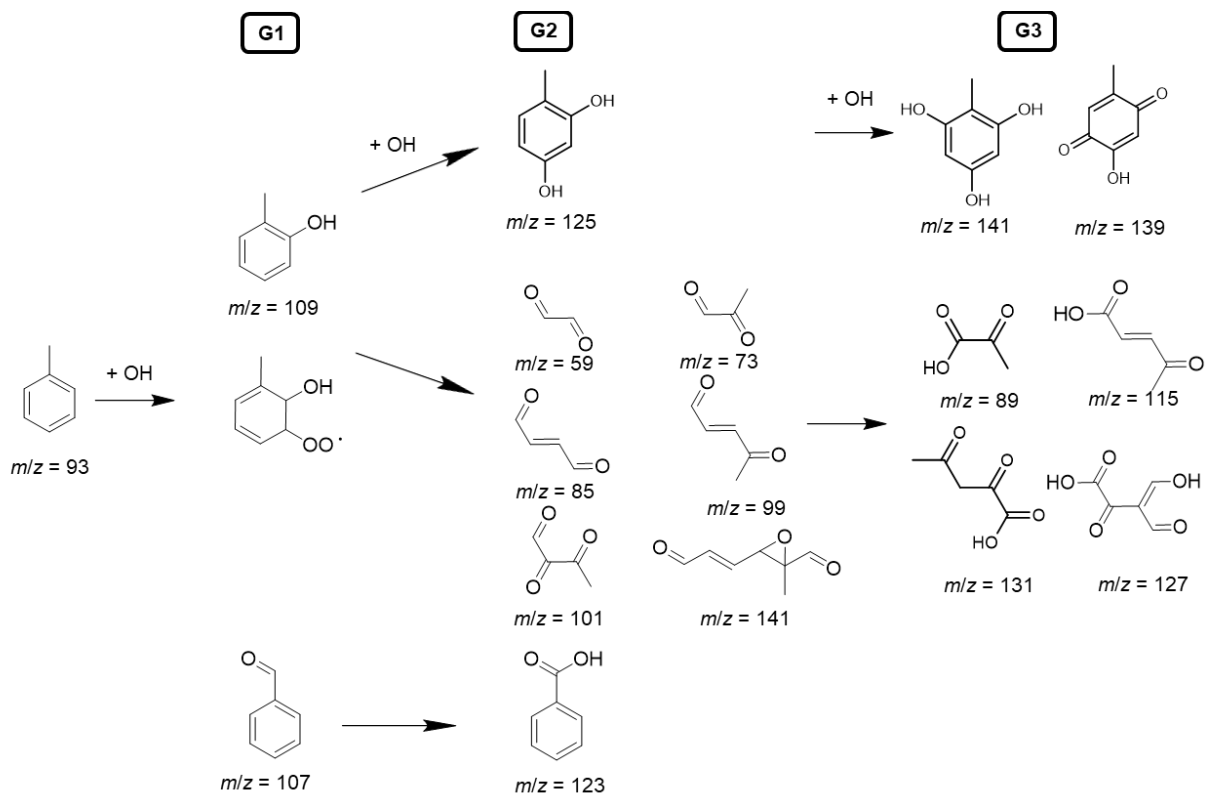
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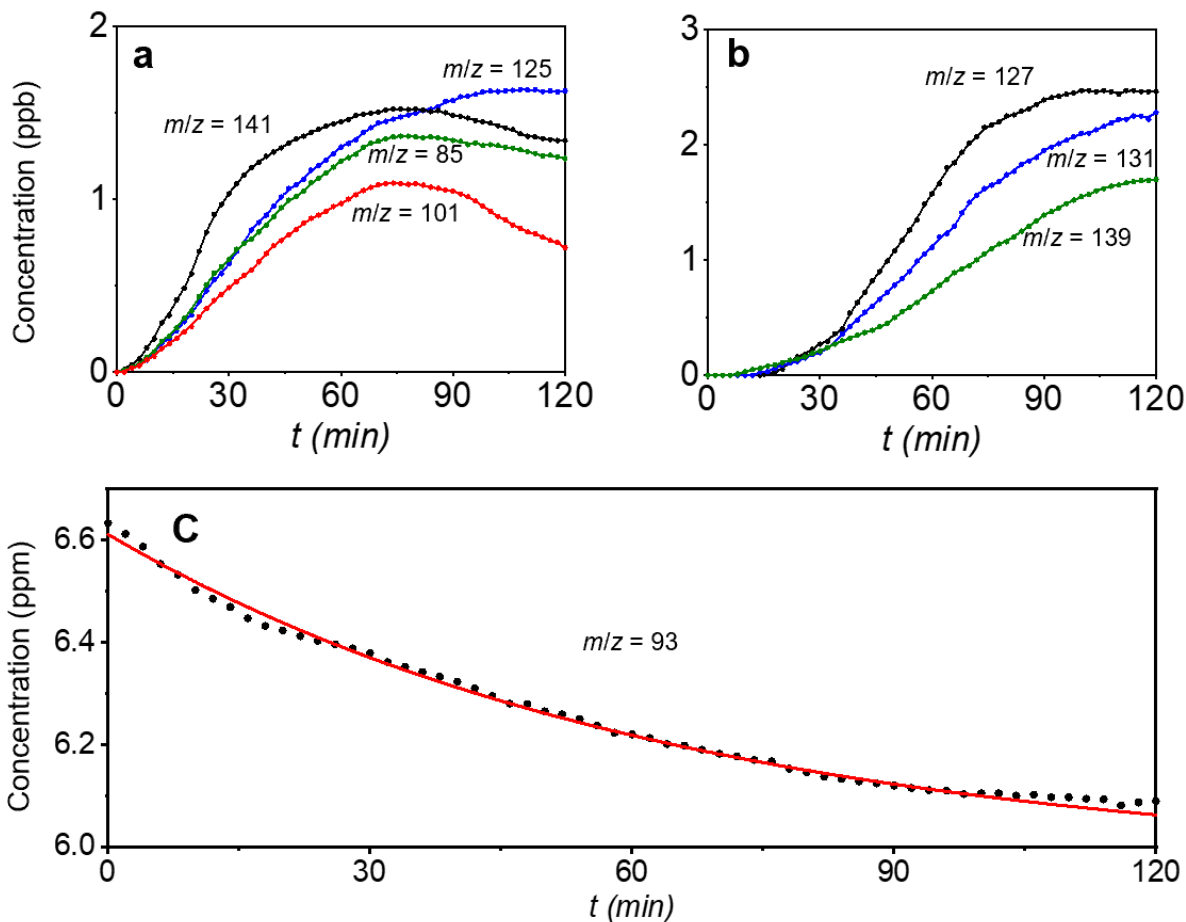




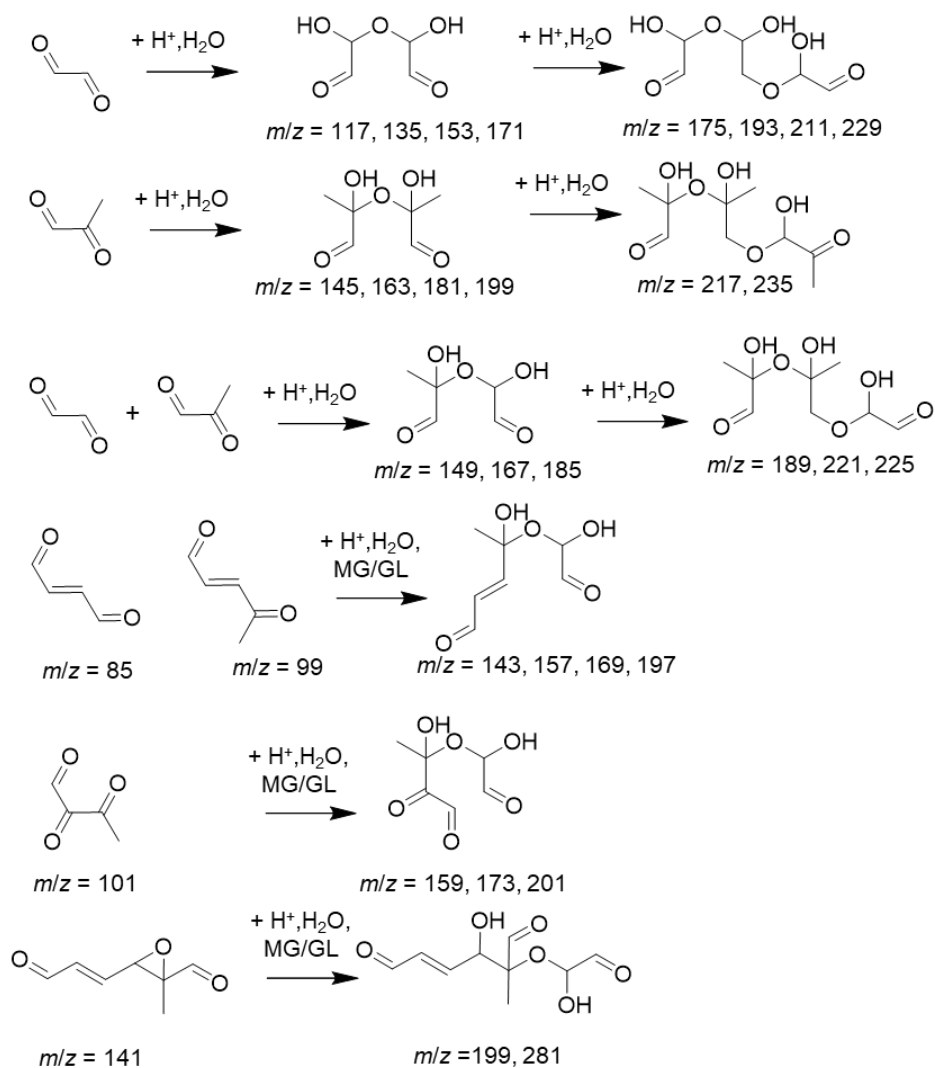
**Figure S1. Schematic representation of the experimental setup.** DMA for differential mobility analyzer; ID-CIMS for ion drift – chemical ionization mass spectrometry; APM for aerosol particle mass analyzer, TD-ID-CIMS for thermal desorption - ion drift – chemical ionization mass spectrometry; SMPS for scanning mobility particle sizer; and CRD for cavity ring-down spectroscopy. Monodispersed seed particles consisting of ammonium bisulfate, ammonium sulfate, and sodium chloride were exposed to oxidation products between toluene by OH and in the absence/presence of NH<sub>3</sub> vapors inside a 1 m<sup>3</sup> reaction chamber. RH was regulated between 10% and 90%, and *T* was maintained at 298 K. The gaseous concentrations of the oxidation products and particle properties, such as density, chemical composition, growth, and single scattering albedo, were simultaneously quantified by ID-CIMS, APM, TD-ID-CIMS, SMPS, and CRD-Nephelometer, respectively.



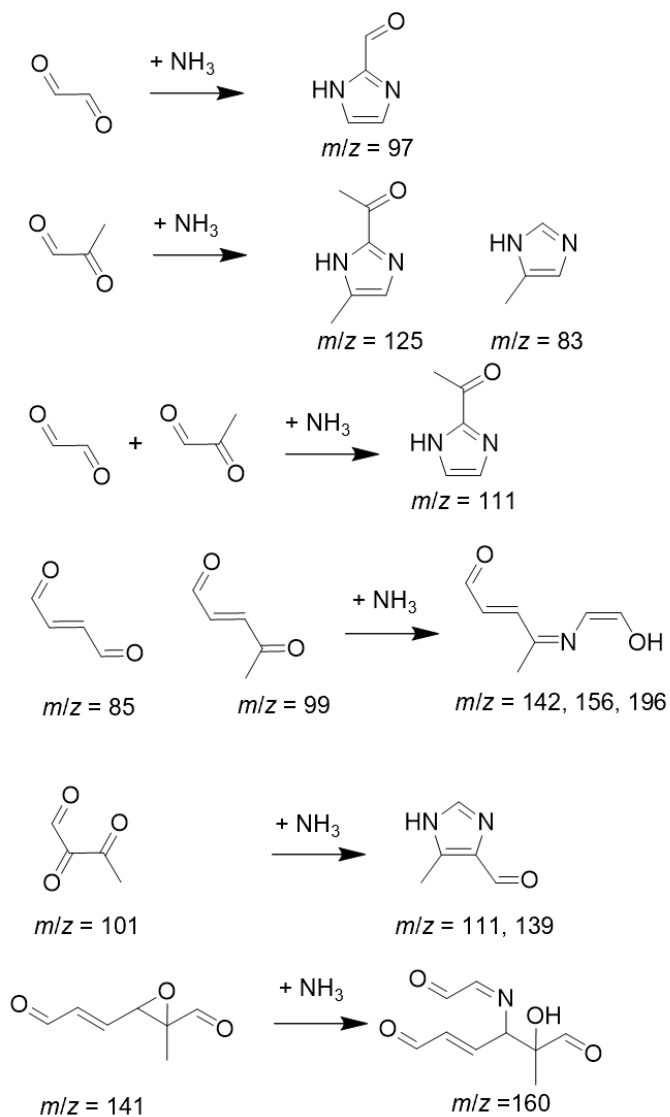
**Figure S2. Multi-generation products from toluene-OH photooxidation.** The letters of G1, G2, and G3 denote the sequence of reactions with OH, and the numbers denote the mass to charge ratio ( $m/z$ ).



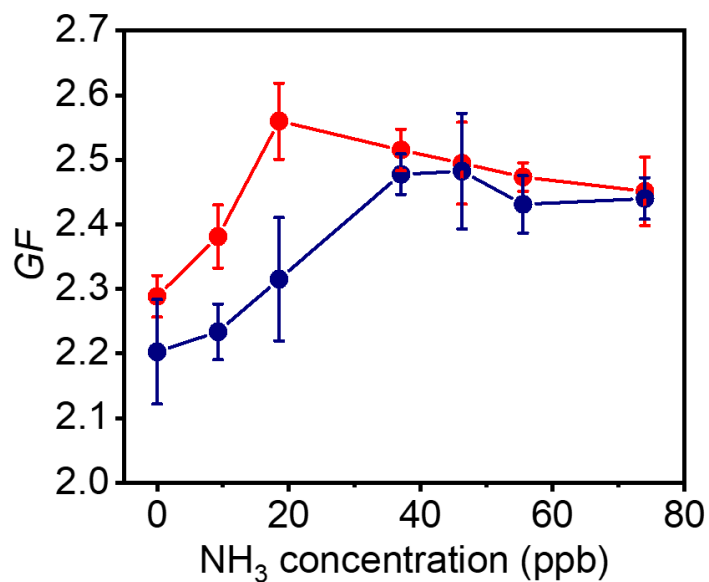
**Figure S3. Temporal evolutions in products and reactants from toluene oxidation.** The gaseous concentrations of G2 products (a), G3 products (b), and toluene (c). The red solid line in (c) corresponds to kinetic simulation of the toluene concentration. The initial concentration for toluene and the steady-state concentration for OH were  $6.6 \text{ ppm}$  and  $2.7 \times 10^6 \text{ molecules cm}^{-3}$ , respectively, with a bimolecular rate constant of  $5.5 \times 10^{-12} \text{ cm}^3 \text{ molecule}^{-1} \text{ s}^{-1}$ . The experiments were carried out on  $(\text{NH}_4)_2\text{SO}_4$  seed particles with  $19 \text{ ppb NH}_3$  and at  $298 \text{ K}$  and  $\text{RH} = 90\%$ . Initiation of photooxidation by ultraviolet light occurred at  $t = 0$ .



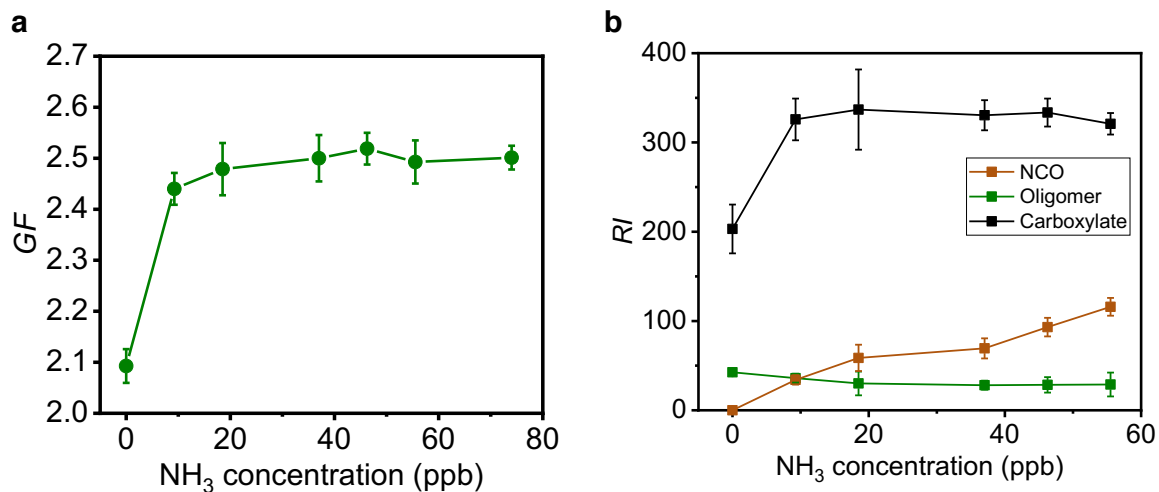
**Figure S4. Pathways for oligomerization.** The mechanisms leading to formation of particle-phase oligomers from  $\alpha$ -dicarbonyls,  $\gamma$ -dicarbonyls, tricarbonyls, and epoxide detected by TD-ID-CIMS.



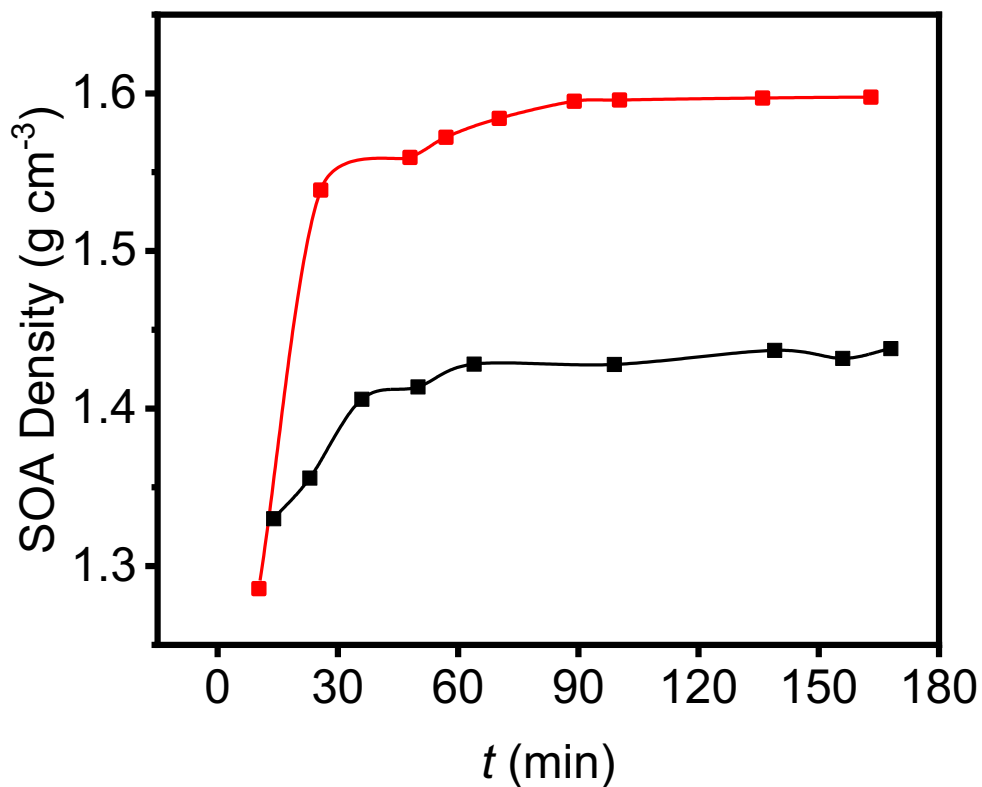
**Figure S5. Pathway for BrC formation.** The mechanisms leading to formation of particle-phase nitrogen-containing organics (NCO) from  $\alpha$ -dicarbonyls,  $\gamma$ -dicarbonyls, tricarbonyls, and epoxide.



**Figure S6. Size growth with NH<sub>3</sub>.** Comparison of measured *GF* at *t* = 90 mins between NH<sub>4</sub>HSO<sub>4</sub> (blue) and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (red) seed particles exposed to toluene-OH oxidation products at varying NH<sub>3</sub> concentrations between 0 and 74 ppb. The error bar denotes 1σ of 3 repeated measurements. All experiments were carried out at 298 K and RH = 90%.

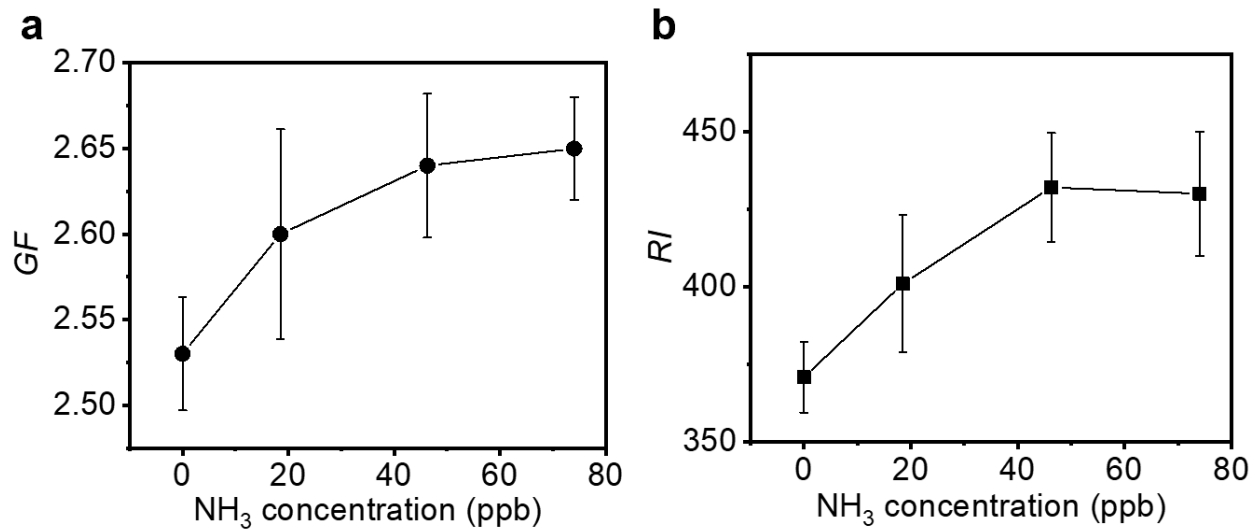


**Figure S7. Variations in size growth and composition with  $\text{NH}_3$ .** Measurements of  $GF$  and chemical compositions on NaCl seed particles exposed to toluene-OH oxidation products with varying  $\text{NH}_3$ . (a)  $GF$  at  $t = 90$  mi with varying  $\text{NH}_3$  concentrations between 0 and 74 ppb. (b) Particle-phase mass relative intensity ( $RI$ ) for carboxylates (black), oligomers (green), and NCO (brown). The error bar denotes  $1\sigma$  of 3 repeated measurements.

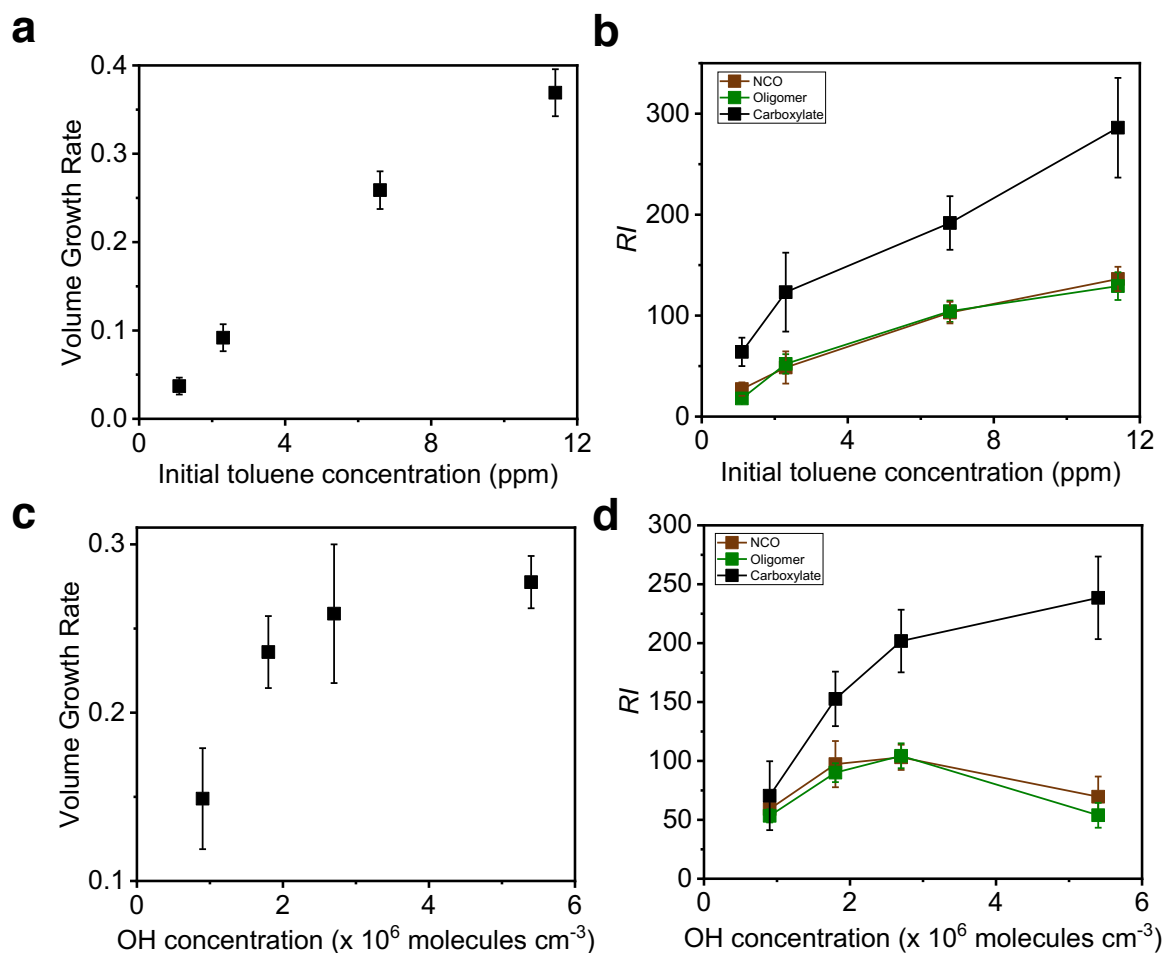


**Figure S8. Temporal evolution in SOA density.** SOA density at RH = 90% (red) and 10% (black). All experiments were carried out on  $(\text{NH}_4)_2\text{SO}_4$  seed particles with 19 ppb  $\text{NH}_3$ . Initiation of photooxidation by ultraviolet light occurred at  $t = 0$ .

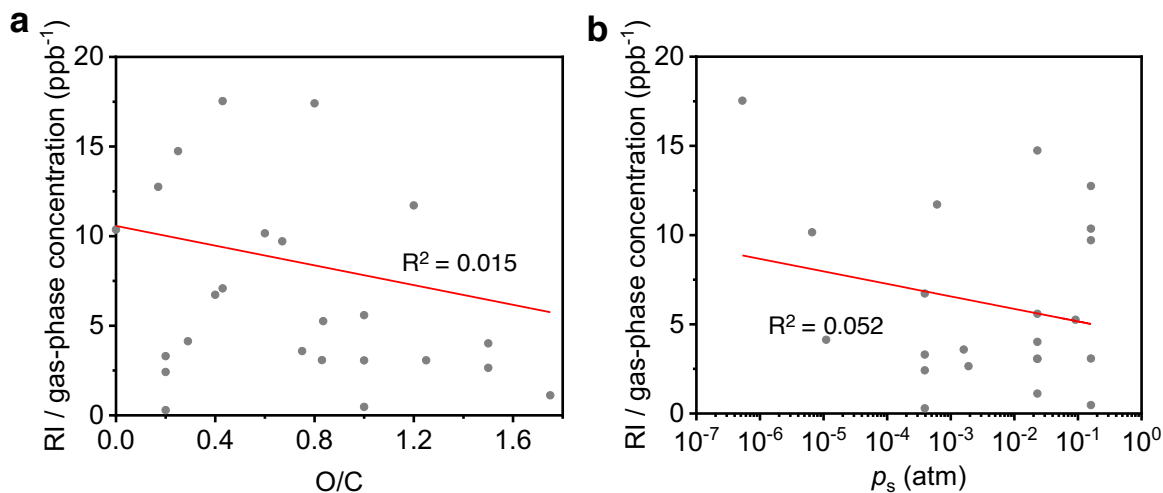




**Figure S9. Variations in size growth and composition with  $\text{NH}_3$ .**  $GF$  (a) and particle-phase mass relative intensity ( $RI$ ) for carboxylates (b) on  $(\text{NH}_4)_2\text{SO}_4$  seed particles at varying  $\text{NH}_3$  concentrations between 0 and 74 ppb and at  $\text{RH} = 10\%$ . The error bar denotes  $1\sigma$  of 3 repeated measurements.



**Figure S10. Variations in particle growth with reactants.** Particle growth under different concentrations of toluene and OH. (a) Particle volume growth rate (defined as  $GR_v = (D_2^3 - D_1^3) / (D_0^3 \Delta t)$ , where  $D_2$  and  $D_1$ , are the diameter of dry particles after 30 min and 10 min of exposure, respectively.  $\Delta t = 20$  min is the time duration) with initial toluene concentration from 1.1 to 11.4 ppm. (b) Particle-phase mass intensity ( $RI$ ) for carboxylates (black), oligomers (green), and NCO (brown) with initial toluene concentration from 1.1 to 11.4 ppm. The OH concentration in (a) and (b) is  $2.7 \times 10^6$  molecules  $\text{cm}^{-3}$ . (c) Measured  $GR_v$  with OH concentration from  $0.9 \times 10^6$  to  $5.4 \times 10^6$  molecules  $\text{cm}^{-3}$ . (d) Particle-phase mass intensity for carboxylates (black), oligomers (green), and NCO (brown) with varying OH concentrations from  $0.9 \times 10^6$  to  $5.4 \times 10^6$  molecules  $\text{cm}^{-3}$ . The initial toluene concentration in (c) and (d) is 6.6 ppm. The error bar denotes  $1\sigma$  of 3 repeated measurements.



**Figure S11. SOA formations versus O/C ratio and volatility.** Correlation of the particle-phase mass intensity ( $RI$ ) of identified SOA components normalized by the gaseous concentrations of the condensable oxidized organic to the oxygen-carbon (O/C) ratio (a) and the saturation vapor pressure at 298K (b). The experiments were performed on  $(\text{NH}_4)_2\text{SO}_4$  seed particles with 19 ppb  $\text{NH}_3$  at  $\text{RH} = 90\%$ , similar to Figure 1h. The red solid line represents linear-fitting through the data.




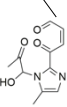

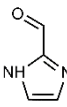

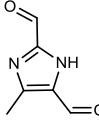
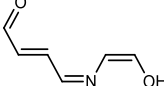
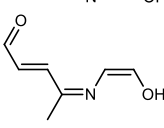
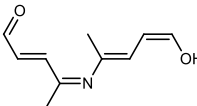
**Table S1. Density of identified major SOA components by TD-ID-CIMS.**

<b>Compounds</b>	<b><i>m/z</i></b>	<b>Density (g cm<sup>-3</sup>)</b>
Methylimidazole	83	1.03
Methylglyoxal	73	1.05
butenedial	85	1.06
4-Oxo-2-pentenoic acid	115	1.2
Imidazole	69	1.23
Pyruvic acid	89	1.25
Benzoic acid	123	1.27
Glyoxal	59	1.27
Dihydroxy toluene	125	1.29
Acetylpyruvic acid	131	1.3
Glyoxylic acid	75	1.38
Trihydroxy toluene	141	1.4
Glyoxal oligomers	175	1.71
2-hydroxy-5-methylquinone	139	1.4
Oxalic acid	91	1.9
Methylglyoxal oligomers	127	1.9


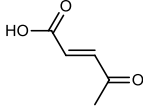
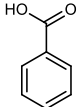
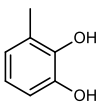
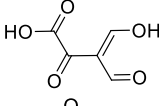
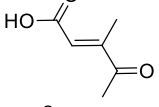
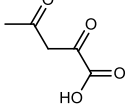
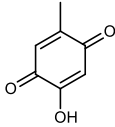
**Table S2. The percent contributions of condensable organic species from toluene-OH oxidation to the particle-phase mass intensity.** GL: glyoxal, MG: methylglyoxal, GL+MG: cross-reaction of glyoxal and methylglyoxal. Note that the cross-reaction products of  $\gamma$ -carbonyls/tricarbonyls with  $\alpha$ -dicarbonyls are summed into the contribution of  $\gamma$ -carbonyls/tricarbonyls. The experiments were carried on  $(\text{NH}_4)_2\text{SO}_4$  seed particle with 19 ppb of  $\text{NH}_3$  and with the initial concentrations of toluene and OH of 6.6 ppm and  $2.7 \times 10^6$  molecules  $\text{cm}^{-3}$ , respectively.

<b>Condensable oxidized organics</b>	<b>Contribution</b>		
	<b>Oligomerization</b>	<b>NCO formation</b>	<b>Total</b>
<b>GL</b>	6%	7%	13%
<b>MG</b>	4%	6%	10%
<b>GL+MG</b>	6%	3%	9%
<b><math>\gamma</math>-dicarbonyls</b>	5%	5%	10%
<b>tricarbonyls</b>	2%	4%	6%
<b>epoxide</b>	3%	1%	4%
<b>Sum</b>	26%	26%	52%
<b>Carboxylates</b>			40%
<b>LVO</b>			8%

**Table S3. Mass peak assignments for nitrogen-containing organics (NCO) from  $\alpha$ -dicarbonyls n-heterocycles and  $\gamma$ -carbonyls n-chains detected by TD-ID-CIMS.**

<b>m/z</b>	<b>MW</b>	<b>Formula</b>	<b>Compound</b>
<b>Methylglyoxal n-heterocycles</b>			
83	82	C <sub>4</sub> H <sub>6</sub> N <sub>2</sub>	
119	118	C <sub>4</sub> H <sub>10</sub> O <sub>2</sub> N <sub>2</sub>	
125	124	C <sub>6</sub> H <sub>8</sub> ON <sub>2</sub>	
251	250	C <sub>6</sub> H <sub>10</sub> O <sub>5</sub>	
<b>Glyoxal n-heterocycles</b>			
69	68	C <sub>3</sub> H <sub>4</sub> N <sub>2</sub>	
97	96	C <sub>4</sub> H <sub>4</sub> ON <sub>2</sub>	
<b>tricarboxyl n-heterocycles</b>			
111	110	C <sub>5</sub> H <sub>6</sub> ON <sub>2</sub>	
139	138	C <sub>6</sub> H <sub>6</sub> O <sub>2</sub> N <sub>2</sub>	
<b><math>\gamma</math>-carbonyls n-chains</b>			
142	141	C <sub>6</sub> H <sub>7</sub> O <sub>2</sub> N	
156	155	C <sub>7</sub> H <sub>9</sub> O <sub>2</sub> N	
196	195	C <sub>4</sub> H <sub>6</sub> O <sub>2</sub> N <sub>2</sub>	

**Table S4. Mass peak assignments for organic acids and low-volatile organics (LVO) detected by TD-ID-CIMS.**

m/z	MW	Formula	Compound
89	88	C <sub>3</sub> H <sub>4</sub> O <sub>3</sub>	
115	114	C <sub>5</sub> H <sub>6</sub> O <sub>3</sub>	
123	122	C <sub>7</sub> H <sub>6</sub> O <sub>2</sub>	
125	124	C <sub>7</sub> H <sub>8</sub> O <sub>2</sub>	
127	126	C <sub>5</sub> H <sub>4</sub> O <sub>5</sub>	
129	128	C <sub>6</sub> H <sub>8</sub> O <sub>3</sub>	
131	130	C <sub>5</sub> H <sub>6</sub> O <sub>4</sub>	
139	138	C <sub>7</sub> H <sub>6</sub> O <sub>3</sub>	

**Table S5. Mass peak assignments for oligomeric products detected by TD-ID-CIMS.**

	<i>m/z</i>	Ion	MW	Formula
<b>Methylglyoxal</b>				
Dimers	145	[C <sub>6</sub> H <sub>10</sub> O <sub>5</sub> +H-H <sub>2</sub> O] <sup>+</sup>	162	C <sub>6</sub> H <sub>10</sub> O <sub>5</sub>
	163	[C <sub>6</sub> H <sub>10</sub> O <sub>5</sub> +H] <sup>+</sup>	162	C <sub>6</sub> H <sub>10</sub> O <sub>5</sub>
	163	[C <sub>6</sub> H <sub>12</sub> O <sub>6</sub> +H-H <sub>2</sub> O] <sup>+</sup>	180	C <sub>6</sub> H <sub>12</sub> O <sub>6</sub>
	181	[C <sub>6</sub> H <sub>12</sub> O <sub>6</sub> +H] <sup>+</sup>	180	C <sub>6</sub> H <sub>12</sub> O <sub>6</sub>
	181	[C <sub>6</sub> H <sub>14</sub> O <sub>7</sub> +H-H <sub>2</sub> O] <sup>+</sup>	198	C <sub>6</sub> H <sub>14</sub> O <sub>7</sub>
	199	[C <sub>6</sub> H <sub>14</sub> O <sub>7</sub> +H] <sup>+</sup>	198	C <sub>6</sub> H <sub>14</sub> O <sub>7</sub>
Trimers	217	[C <sub>9</sub> H <sub>12</sub> O <sub>6</sub> +H] <sup>+</sup>	216	C <sub>9</sub> H <sub>12</sub> O <sub>6</sub>
	217	[C <sub>9</sub> H <sub>14</sub> O <sub>7</sub> +H-H <sub>2</sub> O] <sup>+</sup>	234	C <sub>9</sub> H <sub>14</sub> O <sub>7</sub>
	235	[C <sub>9</sub> H <sub>14</sub> O <sub>7</sub> +H] <sup>+</sup>	234	C <sub>9</sub> H <sub>14</sub> O <sub>7</sub>
	235	[C <sub>9</sub> H <sub>16</sub> O <sub>8</sub> +H-H <sub>2</sub> O] <sup>+</sup>	252	C <sub>9</sub> H <sub>16</sub> O <sub>8</sub>
	253	[C <sub>9</sub> H <sub>16</sub> O <sub>8</sub> +H] <sup>+</sup>	252	C <sub>9</sub> H <sub>16</sub> O <sub>8</sub>
	253	[C <sub>9</sub> H <sub>18</sub> O <sub>9</sub> +H-H <sub>2</sub> O] <sup>+</sup>	270	C <sub>9</sub> H <sub>18</sub> O <sub>9</sub>
	271	[C <sub>9</sub> H <sub>18</sub> O <sub>9</sub> +H] <sup>+</sup>	270	C <sub>9</sub> H <sub>18</sub> O <sub>9</sub>
	271	[C <sub>9</sub> H <sub>20</sub> O <sub>10</sub> +H-H <sub>2</sub> O] <sup>+</sup>	288	C <sub>9</sub> H <sub>20</sub> O <sub>10</sub>
	289	[C <sub>9</sub> H <sub>20</sub> O <sub>10</sub> +H] <sup>+</sup>	288	C <sub>9</sub> H <sub>20</sub> O <sub>10</sub>
<b>Glyoxal</b>				
Dimers	117	[C <sub>4</sub> H <sub>6</sub> O <sub>5</sub> +H-H <sub>2</sub> O] <sup>+</sup>	134	C <sub>4</sub> H <sub>6</sub> O <sub>5</sub>
	135	[C <sub>4</sub> H <sub>6</sub> O <sub>5</sub> +H] <sup>+</sup>	134	C <sub>4</sub> H <sub>6</sub> O <sub>5</sub>
	153	[C <sub>4</sub> H <sub>8</sub> O <sub>6</sub> +H] <sup>+</sup>	152	C <sub>4</sub> H <sub>8</sub> O <sub>6</sub>
	171	[C <sub>4</sub> H <sub>10</sub> O <sub>7</sub> +H] <sup>+</sup>	170	C <sub>4</sub> H <sub>10</sub> O
Trimers	175	[C <sub>6</sub> H <sub>6</sub> O <sub>6</sub> +H] <sup>+</sup>	174	C <sub>6</sub> H <sub>6</sub> O <sub>6</sub>
	193	[C <sub>6</sub> H <sub>8</sub> O <sub>7</sub> +H] <sup>+</sup>	192	C <sub>6</sub> H <sub>8</sub> O <sub>7</sub>
	211	[C <sub>6</sub> H <sub>10</sub> O <sub>8</sub> +H] <sup>+</sup>	210	C <sub>6</sub> H <sub>10</sub> O <sub>8</sub>
	229	[C <sub>6</sub> H <sub>12</sub> O <sub>9</sub> +H] <sup>+</sup>	228	C <sub>6</sub> H <sub>12</sub> O <sub>9</sub>
	247	[C <sub>6</sub> H <sub>14</sub> O <sub>10</sub> +H] <sup>+</sup>	246	C <sub>6</sub> H <sub>14</sub> O <sub>10</sub>
<b>tricarbonyl</b>				
Monomers	101	[C <sub>4</sub> H <sub>4</sub> O <sub>3</sub> +H] <sup>+</sup>	100	C <sub>4</sub> H <sub>4</sub> O <sub>3</sub>
	159	[C <sub>6</sub> H <sub>6</sub> O <sub>5</sub> +H] <sup>+</sup>	158	C <sub>6</sub> H <sub>6</sub> O <sub>5</sub>
Dimers	173	[C <sub>7</sub> H <sub>8</sub> O <sub>5</sub> +H] <sup>+</sup>	172	C <sub>7</sub> H <sub>8</sub> O <sub>5</sub>
	201	[C <sub>8</sub> H <sub>8</sub> O <sub>6</sub> +H] <sup>+</sup>	200	C <sub>8</sub> H <sub>8</sub> O <sub>6</sub>
<b>γ-dicarbonyls</b>				
Monomers	85	[C <sub>4</sub> H <sub>4</sub> O <sub>2</sub> +H] <sup>+</sup>	84	C <sub>4</sub> H <sub>4</sub> O <sub>2</sub>
	99	[C <sub>5</sub> H <sub>6</sub> O <sub>2</sub> +H] <sup>+</sup>	98	C <sub>5</sub> H <sub>6</sub> O <sub>2</sub>
Dimers	143	[C <sub>6</sub> H <sub>6</sub> O <sub>4</sub> +H] <sup>+</sup>	142	C <sub>6</sub> H <sub>6</sub> O <sub>4</sub>
	157	[C <sub>7</sub> H <sub>8</sub> O <sub>4</sub> +H] <sup>+</sup>	156	C <sub>7</sub> H <sub>8</sub> O <sub>4</sub>
	169	[C <sub>8</sub> H <sub>8</sub> O <sub>4</sub> +H] <sup>+</sup>	168	C <sub>8</sub> H <sub>8</sub> O <sub>4</sub>
	197	[C <sub>10</sub> H <sub>12</sub> O <sub>4</sub> +H] <sup>+</sup>	196	C <sub>10</sub> H <sub>12</sub> O <sub>4</sub>
<b>epoxide</b>	141	[C <sub>7</sub> H <sub>8</sub> O <sub>3</sub> +H] <sup>+</sup>	140	C <sub>7</sub> H <sub>8</sub> O <sub>3</sub>



**Table S6.** Measured uptake coefficient ( $\gamma$ ) for condensable oxidized organics on ammonium sulfate seed particles in the presence of ammonia. The uncertainty in the calculated  $\gamma$  reflects the combined random errors in the measured five parameters, i.e.,  $D_o$  = initial diameter,  $D_p$  = final diameter after the exposure time of  $dt$ ,  $N$  = the particle number concentration,  $S$  = average particle surface area, and  $[A]$  = average concentration of species A. FR = percent contributions to the particle-phase mass intensity (Table S2). For each parameter, an uncertainty is estimated, corresponding to  $1\sigma$  of at least 3 measurements.

Species	$D_o$ (nm)	$D_p$ (nm)	$N$ ( $\text{cm}^{-3}$ )	$S$ ( $10^{-5}$ $\text{cm}^2 \text{cm}^{-3}$ )	$[A]$ (ppb)	FR	$\gamma$ ( $10^{-3}$ )
<b>RH = 90%</b>							
Glyoxal	100.0	256.0	15000	1.55	1.9	13%	$8.3 \pm 2.0$
Methylglyoxal	100.0	256.0	15000	1.55	0.9	10%	$12 \pm 2.8$
$\gamma$ -Dicarbonyls	100.0	256.0	15000	1.55	3.1	10%	$3.0 \pm 0.6$
Benzoic acid	100.0	256.0	15000	1.55	0.9	0.9%	$0.84 \pm 0.11$
4-Oxo-2-pentenoic acid	100.0	256.0	15000	1.55	1.9	4.8%	$2.2 \pm 0.4$
Pyruvic acid	100.0	256.0	15000	1.55	0.9	2.6%	$2.9 \pm 0.3$
LVO	100.0	256.0	15000	1.55	1.2	8.0%	$5.0 \pm 1.5$
Nitrophenol*	100.0	246.0	15000	1.46	1.4	3.1%	$1.6 \pm 0.3$
<b>RH = 10%</b>							
Glyoxal	100.0	260.0	15000	0.85	5.4	1.9%	$0.75 \pm 0.18$
Methylglyoxal	100.0	260.0	15000	0.85	3.2	1.5%	$0.90 \pm 0.20$
$\gamma$ -Dicarbonyls	100.0	260.0	15000	0.85	9.6	2.0%	$0.34 \pm 0.07$
Benzoic acid	100.0	260.0	15000	0.85	2.0	1.1%	$0.80 \pm 0.14$
4-Oxo-2-pentenoic acid	100.0	260.0	15000	0.85	6.8	9.1%	$2.0 \pm 0.5$
Pyruvic acid	100.0	260.0	15000	0.85	3.6	5.3%	$2.5 \pm 0.4$
LVO	100.0	260.0	15000	0.85	2.9	9.4%	$4.2 \pm 1.5$
Nitrophenol*	100.0	253.5	15000	0.82	2.3	3.0%	$1.6 \pm 0.3$

\* Measured in the presence of 100 ppb  $\text{NO}_x$

**Table S7. Relative contributions of condensable oxidized organics with each type of functionality to SOA formation on (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> seed particles with 19 ppb NH<sub>3</sub> and at 10% - 90% RH.**

Functionality	SOA Contribution (%)			
	10	50	70	90
Di-carbonyls	3	19	22	26
Di-carbonyls+NH <sub>3</sub>	2	14	20	26
Carboxylates	86	58	49	40
LVO	9	9	9	8

**Table S8. Oxygen to carbon (O/C) ratio for identified SOA components and saturation vapor pressures ( $P_s$ ) of condensable oxidized organics.**

<b>Compound</b>	<b>O/C</b>	<b><math>p_s</math> (atm, 298 K)</b>
<b>Oligomers</b>		
Glyoxal	1	$2.3 \times 10^{-2}$
Glyoxal dimers	1~1.75	
Glyoxal trimers	1~1.67	
Methylglyoxal	0.67	$1.6 \times 10^{-1}$
Methylglyoxal dimers	0.67~1.17	
Methylglyoxal trimers	0.67~1.11	
Butenedial	0.5	$7.9 \times 10^{-4}$
Butenedial dimers	0.5~0.87	
Methylbutenedial	0.4	$3.9 \times 10^{-4}$
Methylbutenedial dimers	0.4~0.7	
Tricarbonyl	0.75	$1.6 \times 10^{-3}$
Tricarbonyl dimers	0.75~0.9	
Epoxide	0.43	
<b>NCO</b>		
Glyoxal N-heterocycles	0~0.25	
Methylglyoxal N-heterocycles	0~0.17	
Butenedial N-heterochains	0~0.25	
Methylbutenedial N-heterochains	0~0.2	
Tricarbonyl N-heterocycles	0.2~0.4	
Epoxide N-heterochains	0~0.44	
<b>Organic acids</b>		
Benzoic acid	0.29	$1.1 \times 10^{-5}$
4-Oxo-2-pentenoic acid	0.6	$6.6 \times 10^{-6}$
Acetylpyruvic acid	0.8	
Pyruvic acid	1.2	$6.1 \times 10^{-4}$
Glyoxylic acid	1.5	$1.9 \times 10^{-3}$
<b>Others</b>		
Dihydroxy toluene	0.29	
Trihydroxy toluene	0.43	$1.1 \times 10^{-8}$
2-Hydroxy-5-methylquinone	0.43	$5.3 \times 10^{-7}$

**Table S9.** Comparison of measured gaseous concentrations of condensable oxidized organics from toluene-OH oxidation between 10% and 90% RH.

<i>m/z</i>	Average Concentration (ppb)		Ratio (10% to 90% RH)
	10% RH	90% RH	
59	5.4	1.9	2.8
73	3.2	0.9	3.4
85	2.6	0.7	3.7
89	3.6	0.9	4.0
91	5.2	2.9	1.8
99	9.6	3.1	3.1
101	3.0	0.7	4.3
107	6.5	4.2	1.5
109	22.6	13.3	1.7
111	5.9	3.1	1.9
115	6.8	1.9	3.6
123	2.0	0.9	2.2
125	1.3	1	1.3
127	2.5	1.3	1.9
131	2.3	1.1	2.0
139	2.3	0.9	2.5
141	2.4	1.1	2.1
155	0.7	0.2	3.2
157	0.5	0.13	3.8