Carbon Dioxide Reduction at Low-Valent Molybdenum Terphenyl Diphosphine Complexes with Dihydrogen and Silanes: Reductant Identity Dictates Mechanism

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General Considerations

Unless otherwise specified, all operations were carried out in an MBraun drybox under a nitrogen atmosphere or using standard Schlenk and vacuum line techniques. Solvents for air- and moisture-sensitive reactions were dried over sodium benzophenone ketyl, calcium hydride, or by the method of Grubbs. Deuterated solvents were purchased from Cambridge Isotope Laboratories and vacuum transferred from sodium benzophenone ketyl (C₆D₆) or CaH₂ (CD₃CN). Solvents, once dried and degassed, were vacuum transferred directly prior to use or stored under inert atmosphere over 4 Å molecular sieves. Molybdenum complexes 1, 2, and 3 were prepared and purified according to reported procedures. Unless indicated otherwise, all chemicals were obtained from standard commercial vendors and utilized as received. ¹H, ¹³C{¹H}, and ³¹P{¹H} NMR spectra were recorded on Varian 400 MHz or Varian INOVA-500 spectrometers with shifts reported in parts per million (ppm). ¹H and ¹³C{¹H} NMR spectra are referenced to residual solvent peaks. ³¹P{¹H} chemical shifts are referenced to external 85% H₃PO₄ (0 ppm).

Synthesis of 4

A brown/red solution of 1 (49.9 mg, 0.085 mmol) in C₆H₆ (3 mL) was transferred to a Schlenk tube charged with a stir bar. The solution was degassed via three freeze-pump-thaw cycles. The headspace was backfilled with 1 atm of CO₂ gas, enacting slight reddening of the solution. The Schlenk tube was sealed and then submerged in LN₂, freezing the contents and condensing the CO₂. It was then opened to 1 atm of H₂ gas, maintaining the -196 °C temperature. The flask was resealed and the contents were allowed to warm to ambient temperature with stirring for 2 h. The now red solution was lyophilized in vacuo, giving 4 as a reddish brown powder. Dissolution of this complex for spectroscopic analysis showed speciation consistent with an equilibrium between complexes 1 – 4, dependent on the reaction vessel atmosphere. The cleanest samples were prepared by adapting the procedure described above for in situ generation of 4 in a J. Young style NMR tube. Attempts to grow single crystals from samples of 4 (prepared as described above) under an N₂ atmosphere reproducibly returned crystalline samples of 1. ¹H NMR (400 MHz, C₆D₆, 23 °C) δ: 8.90 (s, 1H, OCHO), 7.24 (d, J = 7.07 Hz, 2H, aryl-H), 7.05-7.08 (br m, 2H, aryl-H), 6.99 (t, J = 7.46 Hz, 2H, aryl-H), 6.94 (t, J = 7.17 Hz, 2H, aryl-H), 6.21 (s, 2H, central arene-H), 3.59-3.62 (m, 2H, central arene-H), 2.46-2.57 (m, 2H, CH(CH₃)₂), 2.35-2.46 (m, 2H, CH(CH₃)₂), 1.81 (t, 1H, J = 86.41 Hz, Mo-H), 1.21-1.27 (m, 6H, CH(CH₃)₂), 0.94-1.05 (m, 12H, CH(CH₃)₂), 0.80-0.85 (m, 6H, CH(CH₃)₂). ³¹P{¹H} NMR (162 MHz, C₆D₆, 23 °C) δ: 89.82.
Synthesis of 5

A 20 mL scintillation vial was charged with 1 (200 mg, 0.341 mmol), C₆H₁₀ (5 mL), and tBuC₅H₄C(O)OH (73 mg, 0.408 mmol). The red solution was allowed to stir for 8 h, at which time the vial contents were frozen solid in a -35 °C freezer and the C₆H₁₀ lyophilized under reduced pressure, giving 5 as a red powder. X-ray quality single crystals were grown via diffusion of pentane into a concentrated C₆H₁₀ solution of 5. ¹H NMR (400 MHz, C₆D₁₀, 23 °C) δ: 8.31 (d, J = 8.2 Hz, 2H, aryl-H), 7.28 (t, J = 7.0 Hz, 2H, aryl-H), 7.03-7.05 (m, 2H, aryl-H), 6.97 (t, J = 7.3 Hz, 2H, aryl-H), 6.89 (t, J = 7.4 Hz, 2H, aryl-H), 6.46 (v br, 2H, central arene-H), 3.61-3.64 (m, 2H, central arene-H), 2.60-2.64 (m, 2H, CH(CH₃)₂), 2.44-2.53 (m, 2H, CH(CH₃)₂), 2.10 (t, J = 8.4 Hz, Mo-H), 1.34-1.39 (m, 6H, CH(CH₃)₂), 0.96-1.01 (m, 6H, CH(CH₃)₂), 0.75-0.80 (m, 12H, CH(CH₃)₂). ³¹P{¹H} NMR (162 MHz, C₆D₁₀, 23 °C) δ: 89.61 (s).

Synthesis of 6

A brown/red solution of 1 (61 mg, 0.104 mmol) in benzene (4 mL) was transferred to a 20 mL scintillation vial, charged with a stir bar. With stirring, PhSiH₃ (12.8 µL, 0.104 mmol) was added via microsyringe, causing a lightening of the solution. Stirring continued for 30 min. At that time, the solution was lyophilized under reduced pressure, providing 6 as a yellow/orange powder (58.4 mg, 0.088 mmol, 85 %). X-ray quality crystals were grown by chilling a saturated hexanes solution of 6 (-35 °C). ¹H NMR (400 MHz, C₆D₁₀, 23 °C) δ: 7.67-7.69 (m, 2H, aryl-H), 7.33 (d, J = 7.3 Hz, 2H, aryl-H), 7.06-7.07 (m, 3H, aryl-H), 6.97-7.00 (m, 2H, aryl-H), 6.88-6.94 (m, 4H, aryl-H), 5.51 (s, 2H, Si-H), 4.59-4.62 (m, 2H, central arene-H), 4.46 (s, 2H, central arene-H), 2.13-2.23 (m, 2H, CH(CH₃)₂), 2.04-2.13 (m, 2H, CH(CH₃)₂), 1.31-1.18 (m, 6H, CH(CH₃)₂), 0.92-1.08 (m, 18H, CH(CH₃)₂), -3.13 (t, J = 43.41 Hz, 1H, Mo-H). ¹³C NMR (101 MHz, C₆D₁₀, 23 °C) δ: 149.5-149.79 (m, 1H, arene-C), 149.09 (vt, J = 12.35 Hz, arene-C), 146.96 (s, arene-C), 136.60 (s, arene-C), 130.32 (s, arene-C), 128.55 (br s, arene-C), 127.60 (vt, J = 4.4 Hz, arene-C), 127.25 (s, arene-C), 126.99 (s, arene-C), 126.34 (vt, J = 2.0 Hz, arene-C), 97.93 (vt, J = 2.3 Hz, central arene-C), 81.19 (s, central arene-C), 76.40 (s, central arene-C), 31.32 (vt, J = 6.2 Hz, CH(CH₃)₂), 29.01 - 28.57 (m, CH(CH₃)₂), 20.97 (vt, J = 3.6 Hz, CH(CH₃)₂), 20.22 (s, CH(CH₃)₂), 19.96 (vt, J = 2.6 Hz, CH(CH₃)₂). ³¹P{¹H} NMR (162 MHz, C₆D₁₀, 23 °C) δ: 70.79 (d, J = 8.22 Hz).
Preparation of 9

A J. Young style NMR tube was charged with a deep red solution of 1 (25 mg, 0.043 mmol) in C₆D₆ (0.5 mL). The reaction was degassed via three freeze-pump-thaw cycles and then backfilled with OCS (1 atm.). The contents of the tube were mixed by inversion for 15 min, forming 9. Complex 9 was never isolated, but X-ray quality single crystals were obtained by allowing the J. Young tube to stand, unperturbed, for 12 h. ¹H NMR (400 MHz, C₆D₆, 23 °C) δ: 7.52-7.56 (m, 1H, aryl-H), 7.46-7.49 (m, 1H, aryl-H), 7.19-7.23 (m, 1H, aryl-H), 6.98-7.04 (m, 3H, aryl-H), 6.86 (br t, J = 6.6 Hz, 1H, aryl-H), 6.75 (t, J = 8.5 Hz, 1H, aryl-H), 4.79 (d, J = 4.6 Hz, 1H, central arene-H), 4.71 (d, J = 6.5 Hz, 1H, central arene-H), 4.03 (d, J = 6.6 Hz, 1H, central arene-H), 3.83 (d, J = 4.7 Hz, 1H, central arene-H), 3.45-2.60 (m, 2H, CH(CH₃)₂), 2.23-2.35 (m, 1H, CH(CH₃)₂), 1.80-1.91 (m, 1H, CH(CH₃)₂), 1.37-1.42 (m, 3H, CH(CH₃)₂), 1.18-1.27 (m, 6H, CH(CH₃)₂), 1.00-1.12 (m, 6H, CH(CH₃)₂), 0.77-0.95 (m, 9H, CH(CH₃)₂). ³¹P{¹H} NMR (162 MHz, C₆D₆, 23 °C) δ: 88.64 (d, J = 17.7 Hz, Mo-P), 59.57 (d, J = 17.8 Hz, P=S).
**NMR Spectra of Complexes**

**Figure S1.** $^1$H NMR Spectrum (400 MHz, C$_6$D$_6$, 23 °C) of 4. When complex 4 is prepared under an excess of H$_2$ gas, dihydride 2 is observed as a minor reaction component. The resonances associated with this complex are designated with asterisks above.

**Figure S2.** $^{31}$P{$^1$H} NMR Spectrum (162 MHz, C$_6$D$_6$, 23 °C) of 4. When complex 4 is prepared under an excess of H$_2$ gas, dihydride 2 is observed as a minor reaction component and is appropriately labeled in the spectrum above.

**Figure S3.** Partial $^1$H/$^1$H COSY NMR spectrum (600 MHz, C$_6$D$_6$, 23 °C) of complex 4. The correlation between the hydride resonance at 1.85 ppm and that of the central arene proton at 6.20 ppm is highlighted.
Figure S4. $^1$H NMR Spectrum (400 MHz, C$_6$D$_6$, 23 °C) of 5. The inset shows an enlargement of the hydride resonance at 2.10 ppm.

Figure S5. $^{31}$P{$^1$H} NMR Spectrum (162 MHz, C$_6$D$_6$, 23 °C) of 5.
Figure S6. $^1$H NMR Spectrum (400 MHz, C$_6$D$_6$, 23 °C) of 6.

Figure S7. $^{13}$C{$^1$H} NMR Spectrum (101 MHz, C$_6$D$_6$, 23 °C) of 6.

Figure S8. $^{31}$P{$^1$H} NMR Spectrum (162 MHz, C$_6$D$_6$, 23 °C) of 6.
Figure S9. $^1$H NMR Spectrum (300 MHz, C$_6$D$_6$, 23 °C) of in situ generated 9.

Figure S10. $^{31}$P{$^1$H} NMR Spectrum (121 MHz, C$_6$D$_6$, 23 °C) of in situ generated 9.
Silane Coordination Chemistry

A J. Young style NMR tube was charged with a solution of complex 1 (25 mg, 0.043 mmol, 1 equiv.) in C₆D₆ (0.4 mL). To this deep red solution, PhMeSiH₂ (5.9 µL, 0.043 mmol, 1 equiv.) was added via microsyringe, resulting in a slight lightening to red/orange. The tube was sealed, and the contents thoroughly mixed via shaking. ³¹P NMR spectroscopy evidenced ca. 50% conversion of the starting material to coupling doublets at 74.14 and 68.28 ppm (²J(P,P) = 24.7 Hz), assigned to asymmetric silyl hydride complex, 6' (Fig. S11, top). Degassing the reaction vessel via a single freeze-pump-thaw cycle increased the conversion, which was rendered quantitative after a second degassing (Fig. S11).

Figure S11. Partial ³¹P{¹H} NMR spectra (121 Hz, 23 °C, C₆D₆) demonstrating the conversion of dinitrogen adduct 1 to phenylmethylsilyl hydride complex 6'.

While not analytically pure, the salient ¹H NMR features for this new asymmetric species corroborate its assignment as a silyl hydride complex (Fig. S12). ¹H (300 MHz, C₆D₆, 23 °C) δ: 7.74 (br s, 2H, aryl-H), 7.32-7.40 (m, 2H, aryl-H), 7.09 (s, 2H, aryl-H), 6.85-7.03 (m, 7H, aryl-H), 5.72 (br s, 1H, Si-H), 4.67 (br s, 1H, central arene-H), 4.56 (br s, 1H, central arene-H), 4.44 (br s, 1H, central arene-H), 4.23 (br s, 1H, central arene-H), 2.03-2.20 (m, 4H, CH(CH₃)₂), 0.88-1.25 (m, 24H, CH(CH₃)₂), 0.67 (br s, 3H, Si-CH₃), -3.36 (t, J = 39.1 Hz, 1H, Mo-H).
Figure S12. $^1$H NMR Spectrum (300 MHz, C$_6$D$_6$, 23 °C) of *in situ* generated 6′.

Degassing samples prepared with a single equivalent of silane proved more effective at promoting Si–H oxidative addition than adding a gross excess of PhMeSiH$_2$ (Fig. S11, bottom).
**CO₂ Cleavage Kinetics**

In a representative reaction, a J. Young style NMR tube was charged with a deep red C₆D₆ solution (0.6 mL) of 1 (25 mg, 0.043 mmol, 1 equiv.). PhMeSiH₂ (5.9 µL, 0.043 mmol, 1 equiv.) was added via microsyringe, leading to a slight lightening to red/orange. The tube was sealed and the contents frozen solid in a -78 °C dry ice/acetone slurry. The headspace was evacuated thoroughly via three freeze-pump-thaw cycles. After the third evacuation cycle, the tube was carefully positioned such that only the solution was submerged in the cooling bath and CO₂ (1 atm.) was admitted to the head space. The tube was thawed directly prior to introduction to an NMR probe pre-heated to 70 °C. ¹H NMR spectra were collected at 2 minute intervals and the cleavage of CO₂ was monitored via growth of the ¹H resonance at 4.59 ppm, a diagnostic central arene resonance of monocarbonyl complex 8.

**Figure S13.** CO₂ cleavage kinetics at varying silane concentrations as a function of time. The rate of CO₂ reduction increases from 1 [●] to 5 equiv. [×] of PhMeSiH₂ and subsequently decreases at higher silane concentrations—10 [●], 20 [×], and 50 [●] equiv.

**Figure S14.** Substrate inhibition kinetics for CO₂ cleavage in the presence of PhMeSiH₂.
**GC/MS Data for Silanol Identification**

Phenylmethylsilanol was identified as the product of oxygen-atom transfer via the following NMR spectroscopy and gas chromatography/mass spectrometry experiments.

In a representative reaction, a J. Young style NMR tube was charged with a deep red C₆D₆ solution (0.6 mL) of 1 (25 mg, 0.043 mmol, 1 equiv.). PhMeSiH₂ (5.9 µL, 0.043 mmol, 1 equiv.) was added via microsyringe, leading to a slight lightening to red/orange. ¹H and ³¹P{¹H} NMR spectra at this juncture demonstrated ca. 31% conversion of 1 to 6′ (Fig. S15, top).

The tube was sealed and the contents frozen solid in a -78 °C dry ice/acetone slurry. The headspace was evacuated thoroughly via three freeze-pump-thaw cycles. After the third evacuation cycle, the tube was carefully positioned such that only the solution was submerged in the cooling bath and CO₂ (1 atm.) was admitted to the head space. The tube was thawed and placed in a 70 °C oil bath for 8 h. At this time, the reaction was analyzed again by multinuclear NMR spectroscopy, showing complete conversion to 8 (Fig. S15, bottom).

**Figure S15.** Partial ¹H (left; 300 MHz, 23 °C, C₆D₆) and ³¹P{¹H} (right; 121 MHz, 23 °C, C₆D₆) NMR spectra of a reaction mixture prepared as described above. The shaded regions and enlarged insets highlight the diagnostic central arene resonances of 1 (blue) and 8 (red), respectively.

In a fume hood, the contents of the tube were filtered through a short silica plug (ca. 1.5” in a 9” Pasteur pipette) into a 20 mL scintillation vial. The plug was further washed with 1 mL of DCM (benchtop) and two drops of the combined filtrate were transferred to a GC vial, diluted with MeOH, and analyzed by GC/MS.

A control reaction was run, following the procedure outlined above, but with the omission of complex 1.
Figure S16. GC trace from a CO$_2$ cleavage reaction mixture demonstrating the formation of silanol, silyl ethers, and polysiloxane (black). The GC trace of a control reaction run in the absence of 1 is included for reference (red).
**Standard Procedures for Catalytic Reactions**

**Standard procedure for CO$_2$ hydrogenation catalysis:**

In the glove box, 1 (14.7 mg, 25 mmol), base (added as a solid or via microsyringe), and additive (added as a solid, only in entry 1) were loaded into a Teflon stoppered Schlenk tube charged with a stir bar. THF (5 mL) was added, resulting in a red mixture of 1 (5 µM), base (0.5 mM), and additive (0.5 mM, where applicable). After sealing, the vessel was brought out of the glove box, connected to a Schlenk line, and the reaction solution was degassed via three freeze-pump-thaw cycles. The flask was backfilled with CO$_2$ (1 atm) and then re-frozen in LN$_2$, condensing the CO$_2$. The tube was positioned such that only the solution was submerged in LN$_2$ and H$_2$ (1 atm) was admitted. After the gas addition, the reaction solution was allowed to thaw and the Schlenk tube placed in an oil bath pre-heated to 70 °C. Following completion, the reaction was quenched by opening the vessel to the air. The solution was pumped down under reduced pressure, then deuterium oxide (ca. 3 mL) was added to solubilize any formate salts. The insoluble materials were removed by filtration through a glass fiber. DMF was added as an internal standard to the filtrate and the formation of formate salts was quantified by $^1$H NMR spectroscopy.

For catalytic conditions, refer to Table 1 of the manuscript.

**Standard procedure for FA dehydrogenation catalysis:**

In the glove box, 1 (1.55 mg, 0.00265 mmol, 0.1 mol%) in dioxane (5 mL)—prepared as a stock solution and kept frozen when not in use—and additive (0.0265 mmol, 10 mol%—either added as a solid or via microsyringe) were loaded into 25 mL Schlenk flask charged with a stir bar. After sealing, the Schlenk flask was brought out of the glove box and connected to Eudiometry setup filled with nitrogen atmosphere. Formic acid (0.1 mL, 2.65 mmol, 1 equiv.) was added quickly via syringe. Following regulation of the pressure, the Eudiometer was opened, and the gas evolution was monitored as a function of time.

For catalytic conditions, refer to Table 2 of the manuscript.

**Characteristic Gas Evolution Time-Course**

![Representative Gas Evolution Timecourse](image)

**Figure S17.** Gas evolution as a function of time for a typical high turnover formic acid dehydrogenation experiment (the data shown corresponds to Table 2, entry 8). In all cases, negligible gas evolution was observed after the reported time/TON.
Catalyst Decomposition Analysis

To explore the fate of the catalyst, a formic acid dehydrogenation trial was run under modified standard conditions with a 1 mol% loading of dinitrogen adduct 1. Removal of volatiles and analysis of the resulting products by multinuclear NMR spectroscopy corroborated formation of a mixture of 8, P2\(^{O}\), and free ligand (P2).

Figure S18. \(^1\text{H}\) (left—400 MHz, 23 °C, C\(_6\)D\(_6\)) and \(^{31}\text{P}\{\text{^1}\text{H}\}\) (right—162 MHz, 23 °C, C\(_6\)D\(_6\)) spectra following catalytic formic acid dehydrogenation (top) in comparison to an authentic sample (bottom) of Mo monocarbonyl complex 8. The insets highlight the central arene protons of 8 and the shaded boxes designate speciation in the \(^{31}\text{P}\{\text{^1}\text{H}\}\) NMR spectrum.
Stoichiometric Reactions Targeting Catalytically Relevant Reaction Steps

Stoichiometric Addition of Formic Acid
To investigate the stoichiometric reactivity between dinitrogen adduct 1 and formic acid, a J. Young style NMR tube was charged with a solution of 1 (25 mg, 0.043 mmol, 1 equiv.) in C₆D₆ (0.5 mL). Formic acid (8.1 µL, 0.215 mmol, 5 equiv.) was added via microsyringe. The J. Young tube was then sealed, mixed, and the contents analyzed by multinuclear NMR spectroscopy. The ¹H NMR spectrum showed free formic acid and the ³¹P{¹H} NMR spectrum evidenced two primary diamagnetic molybdenum complexes (Fig. S19, top). Upon heating the reaction solution to 70 °C for one hour, the formic acid was consumed, hydrogen was generated, and the predominant metal complex in solution was formate hydride complex 4 (Fig. S19, middle).

Figure S19. Stacked partial ¹H (left—300 MHz, 25 °C, C₆D₆) and ³¹P{¹H} (right—121 MHz, 25 °C, C₆D₆) spectra monitoring conversion of 1 to 4 in the presence of formic acid. The bottom spectra show independently prepared 4 for comparison.

Thermal Decomposition of 4
A sample of 4 in a J. Young style NMR tube was prepared as follows: 1 (25 mg, 0.043 mmol, 1 equiv.) was dissolved in 0.5 mL of C₆D₆. The tube was sealed, and the contents frozen solid in liquid nitrogen. The headspace was evacuated, the contents of the tube thawed, and CO₂ (1 atm.) was admitted at room temperature, leading to a lightening of the deep red solution to red orange. The tube was re-frozen in liquid nitrogen, simultaneously condensing the CO₂. While frozen, the tube was opened to H₂ (1 atm.). Following this second gas addition, the tube was carefully thawed and mixed thoroughly via inversion. ³¹P{¹H} NMR spectroscopy demonstrated ca. 91% conversion to formate hydride 4.
Mixing the reaction via inversion of the J. Young tube for 3 h led to little change in the reaction speciation; however, after heating to 70 °C for 1 h, 4 converted to dihydride 2 (40%), CO$_2$ adduct 3 (19%), and dinitrogen complex 1 (23%).

![Figure S20](image)

**Figure S20.** Tracking thermal dehydrogenation of formate hydride complex 4 via $^{31}$P{^1}H NMR spectroscopy (121 MHz, 25 °C, C$_6$D$_6$). An *in situ* prepared sample of 4 (91%) shows little conversion at room temperature (middle) but converts to 1 – 3 when heated to 70 °C for 1 h (bottom).

**Stoichiometric Reactions with Lewis Acids and Base**

Given that the following reactions were run at room temperature, they employed isolated 4 (prepared via the procedure described on page S3) under an N$_2$ atmosphere.

In a typical reaction, a J. Young style NMR tube was charged with a solution of 4 (20 mg, 0.033 mmol, 1 equiv.) in THF (0.6 mL). To this same tube, DBU (49.4 µL, 0.330 mmol, 10 equiv.), LiOTf (52 mg, 0.330 mmol, 10 equiv.) or both were added. The tube was sealed, at the content mixed via inversion for 8 h.

![Figure S21](image)

**Figure S21.** Stacked $^{31}$P{^1}H NMR spectra (121 MHz, 23 °C, THF) of reactions of 4 with DBU (top), LiOTf (middle), and both (bottom). The enlargement to the right highlights the closely spaced but resolved resonances of 4 and hydride cation 12.

**Note:** The conversion to 1 observed in the reaction of 4 with 10 equiv. of LiOTf could be attributed to minor dehydrogenation under N$_2$ as trace amounts of 2 and 3 were likewise present in the reaction mixture.
Independent Synthesis of Hydride Cation 12

A J. Young style NMR tube was charged with 1 (25 mg, 0.043 mmol, 1 equiv.), LutHOTf (11 mg, 0.043 mg, 1 equiv.), and C₆D₆ (0.6 mL). The heterogeneous mixture was heated to 70 °C for 2 h. Analysis by multinuclear NMR spectroscopy identified a new species, 12, assigned as a hydride cation with the following diagnostic resonances: ¹H (300 MHz, 23 °C, C₆D₆) δ: 7.22 (d, J = 7.1 Hz, 2H, aryl-H), 7.12 (dd, J = 7.2 & 5.4 Hz, 2H, aryl-H), 7.02 (dd, J = 7.4 & 6.9 Hz, 4H, aryl-H), 2.39 - 2.50 (m, 4H, CH(CH₃)₂), 2.31 (t, J = 86.1 Hz, 1H, Mo-H), 1.15 (dd, J = 16.0 & 7.0 Hz, 12H, CH(C₆H₃)), 1.02 (dd, J = 14.5 & 7.2 Hz, 12H, CH(CH₃)₂). A very broad resonance centered at 4.6 ppm, ΔfFWHM ca. 600 Hz, is attributed to the central arene protons. ³¹P{¹H} (121 MHz, 23 °C, C₆D₆) δ: 89.33. A ³¹P spectrum corroborated the ²J(P,H) scalar coupling between the hydride ligand and phosphine donors.

Removal of volatiles from a sample prepared as described above followed by addition of CD₃CN results in formation of previously reported hydride acetonitrile cation 13 (Fig. S2, bottom), further corroborating the proposed assignment.

![Figure S22](image_url)  
**Figure S22.** Stacked ¹H (left–400 MHz, 23 °C, C₆D₆) and ³¹P{¹H} (right–162 MHz, 23 °C, C₆D₆) NMR spectra of *in situ* prepared hydride cation 12 and subsequent conversion to reported acetonitrile adduct, 13. The insets shows an enlargement of the hydride resonance at 2.31 ppm (²J(H,P) = 86.1 Hz) and the corresponding ³¹P NMR signal showing appropriate splitting and scalar coupling. Spectra of the same sample in CD₃CN (bottom) match previously reported 13. Asterisks designate a minor starting material (1) impurity that is insoluble in MeCN.

Protonation Attempts of Formate Hydride 4 and Carboxylate Hydride 5

A sample of 4 was prepared as described above and 20.0 mg (0.033 mmol) was charged in a J.Young style NMR tube. This tube was sealed, the contents degassed, and C₆D₆ (0.5 mL) admitted via vacuum transfer. ¹H and ³¹P{¹H} NMR spectroscopy showed a mixture comprised predominantly of 4, but with impurities of dinitrogen adduct 1 and carbon dioxide adduct 3 (Fig. S23, top).

A second J. Young style NMR tube was charged with complex 4 (24.0 mg, 0.040 mmol, 1 equiv.) from the same batch of starting material as the example above. Solid LutHOTf (10.1 mg, 0.040 mmol, 1 equiv.) was added to the same tube which was subsequently sealed and degassed. C₆D₆ (0.5 mL) was vacuum transferred into this tube which was sealed, mixed, and analyzed by multinuclear NMR spectroscopy, demonstrating a mixture comprised primarily of hydride cation 12. To confirm the formation of 12, the
volatiles were removed under reduced pressure and CD$_3$CN (0.5 mL) was added to the tube via vacuum transfer. $^1$H and $^{31}$P{$^1$H} NMR spectroscopy demonstrated 13 as the predominant product (Fig. SX, bottom).

These results suggest the successful protonation of complex 4 to afford 12. Whereas there are equilibrium processes (namely de-insertion from 4 to regerate 1, 2 and 3–all of which can react with acid to yield 12) that may be at play, these are slow on the timescale of the protonation. The experimental results are not inconsistent with the proposed reactivity via formate cation 14, but they likewise fail to provide direct support for this pathway. As such, protonation of complex 5 (a surrogate of 4) was explored.

A J. Young style NMR tube was charged with solid 5 (26.1 mg, 0.035 mmol, 1 equiv.), LutHOTf (9.1 mg, 0.035 mmol, 1 equiv.), and C$_6$D$_6$ (0.5 mL). The tube was sealed and the contents mixed via rapid inversion. $^1$H and $^{31}$P{$^1$H} NMR spectra evidenced conversion to hydride cation 12 (Fig. S24, middle), which was further confirmed via removal of the reaction volatiles and addition of CD$_3$CN (Fig. S24, bottom). Free $^1$BuC$_6$H$_4$C(O)OH was likewise formed and the relative integration was consistent with exclusive pronolysis of the carboxylate.
Protonolysis of the formate ligand of 4, in an analogous fashion to the reactivity established above, would afford hydride cation 12 without release of H$_2$ or CO$_2$. Addition of excess carboxylic acid to complex 5 showed no reaction under catalytic conditions. These results are consistent with the reactivity of p-$t$-BuC$_6$H$_4$C(O)ONa differing from that of HC(O)OH, with the latter being both more acidic and being capable of additional decarboxylation chemistries. Taken together, these results are neither indicative of nor inconsistent with the mechanism proposed in the “fast” cycle of Scheme 5.

**Carboxylate Addition to Hydride Cation 12**

To support the proposed conversion of 12 to 4, addition of p-$t$-BuC$_6$H$_4$C(O)ONa to 12 was investigated in an effort to generate carboxylate hydride 5.

Hydride cation 12 was synthesized *in situ* in a J. Young style NMR tube according to the procedure outlined above. This tube was opened in a dinitrogen atmosphere glovebox and p-$t$-BuC$_6$H$_4$C(O)ONa (8.9 mg, 0.044 mmol, 1 equiv.) was added as a solid. The tube was resealed and the contents analyzed by both $^1$H and $^{31}$P-$^1$H NMR spectroscopy. The reaction was then heated to 70 °C and conversion of the starting material monitored over time (Fig. S25).
Figure S25. Stacked $^1$H (400 MHz, 23 °C, C$_6$D$_6$) NMR spectra evidencing the formation of 5 from the reaction of sodium carboxylate with in situ prepared 12.
**Refinement Details**

CCDC deposition numbers 2091815 - 2091817 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Refinement Details—In each case, crystals were mounted on a glass fiber or MiTeGen loop using Paratone oil, then placed on the diffractometer under a nitrogen stream. Low temperature (100 K) X-ray data were obtained on a Bruker KAPPA APEXII CCD based diffractometer (Mo fine-focus sealed X-ray tube, $K_α = 0.71073$ Å) or a Bruker D8 VENTURE Kappa Duo PHOTON 100 CMOS based diffractometer (Mo $I_μ$S HB micro-focus sealed X-ray tube, $K_α = 0.71073$ Å). All diffractometer manipulations, including data collection, integration, and scaling were carried out using the Bruker APEXII or APEXIII software. Absorption corrections were applied using SADABS. Space groups were determined on the basis of systematic absences and intensity statistics and the structures were solved in the Olex 2 software interface by intrinsic phasing using XT (incorporated into SHELXTL) and refined by full-matrix least squares on $F^2$. All non-hydrogen atoms were refined using anisotropic displacement parameters. Hydrogen atoms were placed in the idealized positions and refined using a riding model, unless noted otherwise. The structure was refined (weighed least squares refinement on $F^2$) to convergence. Graphical representation of structures with 50% probability thermal ellipsoids were generated using the Crystal Maker X visualization software.

### Table S1. Crystal and refinement data for complexes 5, 6, and 9.

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$a$ $R_1 = \sum||F_o| - |F_c||/\sum|F_o|$. $b$ wR$_2 = [\sum(w(F_o^2-F_c^2)^2)/\sum(w(F_o^2)^2)]^{1/2}$.
Figure S26. Solid-state structure of 5 with thermal anisotropic displacement ellipsoids shown at the 50% probability level. Hydrogen atoms have been omitted for clarity.
**Figure S27.** Solid-state structure of 6 with thermal anisotropic displacement ellipsoids shown at the 50% probability level.
Figure S28. Solid-state structure of 9 with thermal anisotropic displacement ellipsoids shown at the 50% probability level. Hydrogen atoms have been omitted for clarity.
References

10. Crystallographic data have been deposited at the CCDC, 12 Union Road, Cambridge CB2 1EZ, UK and copies can be obtained on request, free of charge, by quoting the publication citation and the respective deposition numbers.