

**Widespread Occurrence of an Intranuclear Bacterial Parasite in Vent and Seep Bathymodiolin Mussels [Supplementary Material]**

**SUPPLEMENTARY VIDEO LEGENDS**

**Videos.** Animations show stacks of 2D images created by moving the focal plane through the z-axis of 10  $\mu\text{m}$  thick sections at 0.2  $\mu\text{m}$  intervals. S1: developmental stage 2; S2: developmental stage 3; S3: developmental stage 4; S4: developmental stage 6.

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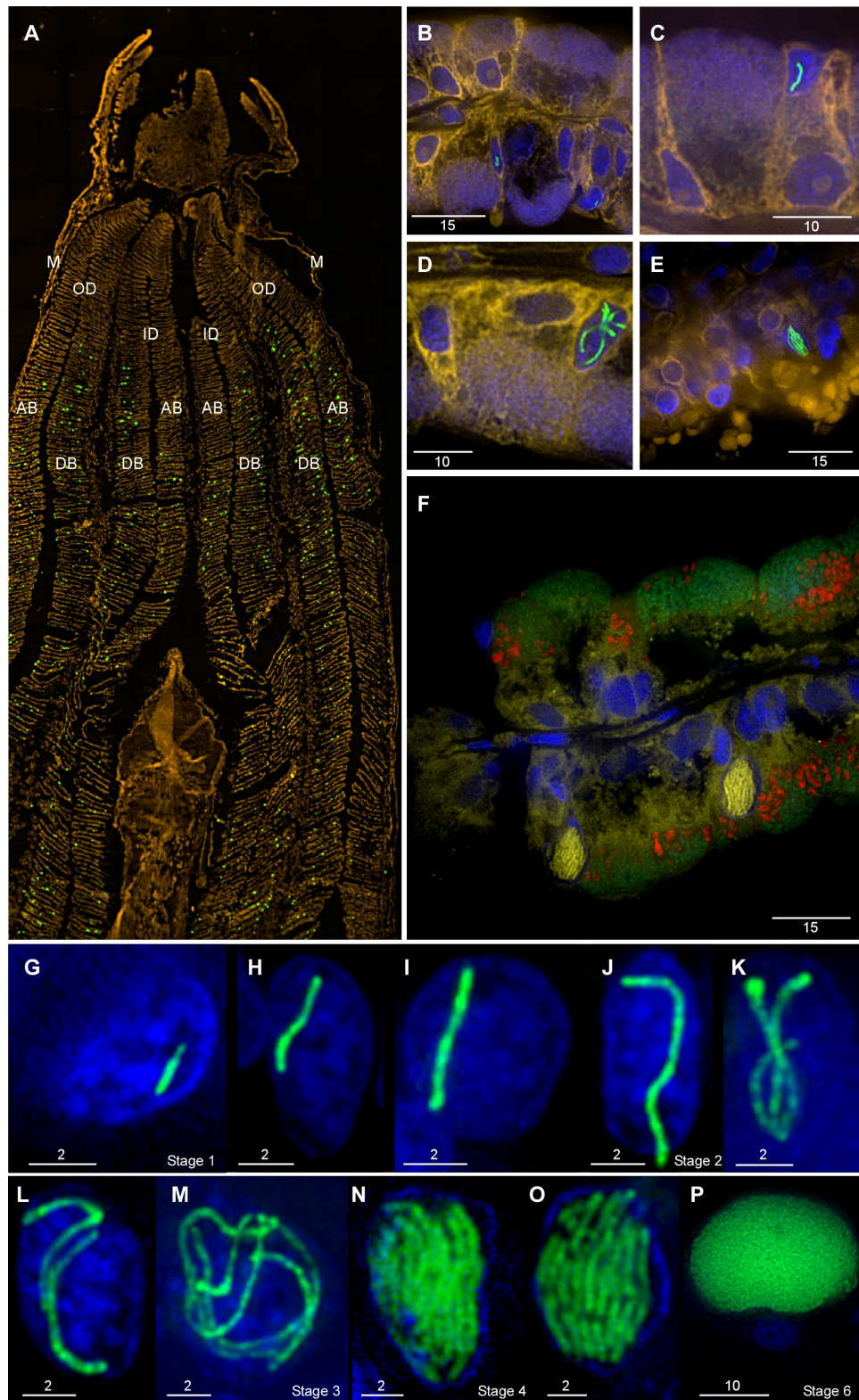
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## SUPPLEMENTARY FIGURES



**Figure S1.** “*Ca. E. bathymodioli*” in various mussel tissues and developmental stages (original RGB image of Fig. 2 in main document). For figure caption see main document.

## SUPPLEMENTARY TABLES

Table S1. Closest relatives of “*Candidatus Endonucleobacter bathymodioli*” (Logatchev)

Uncultured gamma proteobacterium or isolate	Identities [%]	Sequence length [bp]	Associated with	Taxon or species	Accession Number
clone B6F4	98.8	415	hydrothermal fluid	-	AM268617
clone Dd-spF-A85	97.2	389	marine sponge	<i>Discodermia dissoluta</i> <sup>a</sup>	AY897105
clone nerCTABwin06-4*	96.6	684	polychaete worm (gut)	<i>Nereis succinea</i> <sup>b</sup>	EU797581
clone sedCTABwin06-1*	96.0	705	marine sediment	-	EU797583
clone CD14B9 and 46 other clones	95-97	212-616	coral	<i>Diploria strigosa</i> <sup>a</sup>	AF442025
clone Fiji7-A3	95.4	479	seawater	-	DQ670587
clone C19; clone CRNA5	94.5; 95.1	1489; 413	colonial ascidian	<i>Cystodytes dellechiaiei</i> <sup>b</sup>	DQ884169; DQ884160
<i>Spongiobacter nickelotolerans</i> *	94.9	1492	marine sponge	-	AB205011
<i>Kistomonas amurensis</i> *	94.6	1343	seastar	<i>Asterias amurensis</i>	EU599216
<i>Endozoicomonas elysicola</i> *	94.4	1437	sea slug	<i>Elysia ornata</i> <sup>b</sup>	AB196667
clone H1	94.4	531	sea urchin	<i>Tripneustes gratilla</i>	AM495252
clone <i>Spongiobacter</i> sp. ME19 and 5 other clones	92-94	1502-1527	coral	<i>Muricea elongata</i> <sup>b</sup>	DQ917863
clone HOC27; clone HOC2	94.0	1473	marine sponge	<i>Halichondria okadai</i> <sup>b</sup>	AB054161; AB054136
clone PDA-OUT2; clone PDA-OUT3	93.6; 94.0	1500; 1503	coral	<i>Pocillopora damicornis</i> <sup>b</sup>	AY700600; AY700601
clone CN34	93.9	1448	marine sponge	<i>Chondrilla nucula</i>	AM259915
clone s1uc48	93.9	682	coral	<i>Oculina patagonica</i> <sup>b</sup>	DQ416306
clone H262 <sup>d</sup>	93.9	703	marine sponge; sea slug	Dictyoceratida, Thorectidae <sup>b</sup> ; Pleurobranchidae <sup>b</sup>	AY370006
clone H425*	93.9	740	marine sponge	Halichondrida, Axinellidae <sup>b</sup> ; Halichondrida, Halichondriidae <sup>b</sup>	AY370008
clone C23	93.4	1486	colonial ascidian	<i>Cystodytes dellechiaiei</i> <sup>b</sup>	DQ884170
clone HstpL43	93.4	1024	sea grass	<i>Halophila stipulacea</i> <sup>b</sup>	AF159674
clone MOLA 531*	93.3	1423	marine sponge	<i>Petrosia ficiformis</i>	AM990755
clone Gamma7	93.2	919	fish	<i>Salmo salar</i> <sup>c</sup>	AY494615
clone 1C227608	92.9	1420	seawater	-	EU799933
clone CD207E01 and 8 other clones	89.3-92.7	865-983	coral	<i>Montastraea annularis</i> <sup>b</sup>	DQ200605
clone NWCu007	92.2	895	marine sponge	<i>Rhopaloeides odorabile</i> <sup>b</sup>	AF313496
clone <i>Spongiobacter</i> sp. EC22; clone <i>Spongiobacter</i> sp. EC121	92.0; 91.8	1504; 1507	octocoral	<i>Erythropodium caribaeorum</i> <sup>b</sup>	DQ889931; DQ889891
nuclear inclusion X (NIX)	91.1	695	clam	<i>Siliqua patula</i> <sup>d</sup>	M94380-82
clone <i>Spongiobacter</i> sp. BME76	90.9	1512	coral	<i>Muricea elongata</i> <sup>b</sup>	DQ917830
clone OTU 6 and 7 other clones	88.1-90.8	1296-1298	coral	<i>Alcyonium antarcticum</i> <sup>b</sup>	DQ312244
<i>Zooshikella ganghwensis</i> *	90.6	1441	marine sediment	-	AY130994
<i>Oceanrickettsia ariakensis</i> ; R6; <i>O. ariakensis</i> ; R54	87.8	1300; 1304	oyster	<i>Crassostrea ariakensis</i>	DQ118733; DQ123914

<sup>a</sup> Clones obtained from healthy specimens; <sup>b</sup> Condition of specimens not stated; <sup>c</sup> Clone obtained from specimen infected with amoebic gill disease;

<sup>d</sup> bacterium involved in massive mortalities of the Pacific razor clam *Siliqua patula*; \* isolates

**Table S2. Nucleotide differences of dominant and minor “*Ca. E. bathymodioli*” 16S rRNA sequences**

Specimen	Location		# examined <sup>a</sup>	# infected <sup>b</sup>	# full sequences <sup>c</sup>				nucleotide position <sup>d</sup>	Probe <sup>e</sup>	Acc. No	FISH <sup>f</sup>	
					I	II	III	IV					total
<i>B. brooksi</i>	AT, AL, CH	GoM	4	4	8	8	5	6	<b>27</b>		o	FM162182-FM162184	+
<i>B. heckerae</i>	WF, CH	GoM	3	1	5	-	-	na	<b>5</b>		o	FM162185	+
“ <i>B.</i> ” <i>childressi</i>	AL, MC	GoM	2	1	-	2	na	na	<b>2</b>		o	FM244838	+
<i>B. sp.</i>	Lilliput	MAR	1	1	4	na	na	na	<b>4</b>		y	FM162186	/
<i>B. azoricus</i>	Menez Gwen	MAR	3	1	1	-	-	na	<b>1</b>		o	FM162187	+
<i>B. puteoserpentis</i>	Logatchev	MAR	4	3	-	3	3	3	<b>9</b>	474			+
dominant phylotype					-	1	2	3	<b>6</b>		<b>G</b>	FM162188	
minor phylotype					-	2	1	-	<b>3</b>		<b>A</b>	FM162189	
<i>B. sp.</i>	Wideawake	MAR	4	4	8	6	8	8	<b>30</b>	474	1023		+
dominant phylotype					8	4	5	-	<b>17</b>		<b>A</b>	FM162190	
minor phylotype I					-		1	8	<b>9</b>		<b>G</b>	FM162191	
minor phylotype II					-	2	1	-	<b>3</b>		<b>G</b>	FM162192	
minor phylotype III					-	-	1	-	<b>1</b>			FM162193	
<i>B. aff. thermophilus</i>	German Flats	PAR	4	3	6	8	8	-	<b>22</b>	137	184		+
dominant phylotype					5	7	7	-	<b>19</b>		<b>C</b>	FM162194	
minor phylotype					1	1	1	-	<b>3</b>		<b>T</b>	FM162195	

<sup>a</sup> number of examined individuals; <sup>b</sup> number of infected individuals; <sup>c</sup> number of full sequences per individual (I-IV); <sup>d</sup> position based on

*Escherichia coli* numbering; <sup>e</sup> probe specificity: x,y,z – probes Bnix64 (x), Bnix643 (y), and Bnix1249 (z) match their target specifically (no

mismatches), probe Bnix64 is applicable only to “*Ca. E. bathymodioli*” from *B. puteoserpentis* (Logatchev) and “*B.*” *childressi* as this part of the

16S rRNA sequence is not known from the other phylotypes, o – either one of the probes y and z has one single mismatch; <sup>f</sup> results of gill tissue

analysis by fluorescence in situ hybridization (FISH); Abbreviations: AT – Atwater Valley, AL – Alaminos Canyon, CH – Chapopote, MC –

Mississippi Canyon, WF – West Florida Escarpment, GoM – Gulf of Mexico, MAR – Mid-Atlantic Ridge, PAR – Pacific-Antarctic Ridge; na – not

applicable, + positive FISH result, / negative FISH result

**Table S3. Intranuclear bacteria described from protists**

Protist taxon <i>Host species</i>	Protist super group and higher ranking taxa Designation of bacteria inside nuclei	Shape	Phylogeny	Reference
<b>Ciliophora</b> <i>Paramecium</i> spp. and numerous other ciliate genera	<b>Chromalveolata: Alveolata</b> endonuclear bacteria; ( <i>Holospora</i> spp. [9 species] <sup>a, c</sup> , <i>Caedibacter caryophilus</i> <sup>d</sup> , <i>C. macronucleorum</i> <sup>c</sup> , <i>Nonospora macronucleata</i> <sup>c</sup> , and several other but unnamed endonuclear bacteria)	rods	Alpha	(Görtz, 1983, 1986; Görtz and Brigge, 1998; Görtz, 2001; Fokin, 2004; Görtz, 2006)
<b>Dinoflagellata</b> <i>Gyrodinium instriatum</i>	<b>Chromalveolata: Alveolata: Dinozoa</b> endonucleoplasmic bacteria (unnamed) <sup>a, d</sup>	rods	Beta	(Silva and Franca, 1985; Alverca et al., 2002; Biegala et al., 2002)
<i>Gymnodinium splendens</i> ; <i>Glenodinium foliaceum</i>	endonuclear bacteria (unnamed) <sup>a, c</sup>	rods	nd	(Silva, 1978; Silva and Franca, 1985)
<b>Euglenida</b> <i>Euglena spirogyra</i> ; <i>Strombomonas conspersa</i> ; <i>Trachelomonas oblonga</i> ; <i>Lepocinclis ovum</i> ; <i>Hyalophacus ocellatus</i> <i>Euglena hemichromata</i> <i>Euglena deses</i>	<b>Excavata: Euglenozoa</b> endonuclear bacteria (unnamed) <sup>d</sup>  endonuclear bacteria (unnamed) <sup>a, b, c</sup> <i>Caryococcus hypertrophicus</i> <sup>b</sup>	rods  rods cocci	nd  nd nd	(Leedale, 1969)  (Shin et al., 2003) (Dangeard, 1902, 1933; Kirby, 1941, 1944)
<i>Trachelomonas</i> sp. <i>Peranema trichophorum</i>	electron dense bodies <sup>e</sup> rod-shaped bodies <sup>a, e</sup>	rods rods	nd nd	(Ueda, 1960) (Roth, 1959)
<b>Trichonymphida</b> <i>Trichonympha seapiculae</i> <i>Trichonympha corbula</i> <i>Trichonympha peplophora</i> <i>Trichonympha chattoni</i> <i>Staurojoenina assimilis</i>	<b>Excavata: Parabasalia</b> nuclear parasites <i>Caryococcus nucleophagus</i> <sup>c</sup> ; <i>Caryococcus cretus</i> <sup>c</sup> <i>Caryococcus invadens</i> <sup>c</sup> <i>Caryococcus dilatator</i> <sup>a, b, c</sup> intranuclear symbionts <sup>a, c</sup>	spherical bodies cocci cocci cocci cocci	nd nd nd nd nd	(Kirby, 1941) (Kirby, 1944) (Kirby, 1944) (Kirby, 1944) (Dolan et al., 2004)
<b>Cristamonadida</b> <i>Caduceia versatilis</i>	<b>Excavata: Parabasalia</b> intranuclear bacteria <sup>a, e</sup>	cocci	nd	(Tamm and Tamm, 1974; d'Ambrosio et al., 1999)
<b>Spirotrichonymphida</b> <i>Holomastigotoides hemygymnum</i>	<b>Excavata: Parabasalia</b> bactéries dans le noyau <sup>e</sup>	rods	nd	(Hollande and Carruette-Valentin, 1971)

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<sup>a</sup> bacteria cause hypertrophy of nuclei; <sup>b</sup> intranuclear bacteria are associated with the disappearance of chromatin; <sup>c</sup> obligate intranuclear; <sup>d</sup> facultative intranuclear; <sup>e</sup> unclear if obligate or facultative in the nucleus; nd – not determined