



Biogeochemistry of the Stable Isotopes of Hydrogen and Carbon in Salt Marsh Biota

Author(s): Bruce N. Smith and Samuel Epstein

Source: *Plant Physiology*, Vol. 46, No. 5 (Nov., 1970), pp. 738-742

Published by: [American Society of Plant Biologists \(ASPB\)](#)

Stable URL: <http://www.jstor.org/stable/4262256>

Accessed: 16/12/2014 00:25

Your use of the JSTOR archive indicates your acceptance of the Terms & Conditions of Use, available at <http://www.jstor.org/page/info/about/policies/terms.jsp>

JSTOR is a not-for-profit service that helps scholars, researchers, and students discover, use, and build upon a wide range of content in a trusted digital archive. We use information technology and tools to increase productivity and facilitate new forms of scholarship. For more information about JSTOR, please contact support@jstor.org.



American Society of Plant Biologists (ASPB) is collaborating with JSTOR to digitize, preserve and extend access to *Plant Physiology*.

<http://www.jstor.org>

Biogeochemistry of the Stable Isotopes of Hydrogen and Carbon in Salt Marsh Biota¹

Received for publication June 8, 1970

BRUCE N. SMITH² AND SAMUEL EPSTEIN

Division of Geological Sciences, California Institute of Technology, Pasadena, California 91109

ABSTRACT

Deuterium to hydrogen ratios of 14 plant species from a salt marsh and lagoon were 55‰ depleted in deuterium relative to the environmental water. Carbon tetrachloride-extractable material from these plants was another 92‰ depleted in deuterium. This gave a fractionation factor from water to CCl₄ extract of 1.147. This over-all fractionation was remarkably constant for all species analyzed. Plants also discriminate against ¹³C, particularly in the lipid fraction. Data suggest that different mechanisms for carbon fixation result in different fractionations of the carbon isotopes. Herbivore tissues reflected the isotopic ratios of plants ingested. Apparently different metabolic processes are responsible for the different degrees of fractionation observed for hydrogen and carbon isotopes.

MATERIALS AND METHODS

Sample Collection. Fourteen species of plants and six species of animals were collected in Southern California from three sites in a salt marsh and lagoon at Point Mugu Naval Air Base and from Leo Carillo State Beach on the open ocean. The organisms were collected from various habitats but utilized a common source of water. *Distichlis spicata* (L.) Greene, *Limonium commune* S. F. Gray, *Mesembryanthemum chilense* Mol., *Monthochloe littoralis* Engelm., and *Suaeda fruticosa* (L.) Forsk. were collected high in the marsh where the ground surface was dry. The low marsh site had a film of water on the ground. Collections included: *Enteromorpha marginata* J. Agardh, *Frankenia grandifolia* Cham. & Schl., *Salicornia bigelovii* Torr., and the dominant (about 300 snails/m²) *Cerithidia californica* (Haldeman). In the lagoon itself we collected: *Zostera marina* L., *Bulla gouldiana* (Pilsbry), *Dendroaster excentricus* (Escholtz), and *Navanax inermis* (Cooper). At Leo Carillo State Beach we obtained *Macrocystis pyrifera* (L.) C. A. Agardh, *Corallina chilensis* Descainne, *Grateloupia setchellii* Kylin, *Gigartina cristata* (Setchell) Setchell and Gardner, *Phyllospadix torreyi* Wats., *Mytilus californianus* (Conrad), and *Cypraea spadicea* (Swainson). Samples of water were taken from each collection site.

Plants remove hydrogen from water and transfer it to organic compounds. Although many organic hydrogen atoms are exchangeable with environmental water, once a carbon-hydrogen bond is formed in an organism, the hydrogen is no longer readily exchanged. Early studies on the natural abundance of deuterium and hydrogen in organic matter made no attempt to relate the D/H ratios from organic matter to the hydrogen source, *i.e.*, environmental water (3, 18). Zborowski *et al.* (22) determined D/H ratios from fatty acids of fish and rats and marine sediments. They found the fractionation to be constant between fatty acids and water removed from the tissues and sediments by lyophilization. Epstein and Weiss (unpublished) collected plants at successive elevations in the Sierra Nevada. The D/H ratio of total organic matter varied with altitude, as did ground water at each collection site. Schiegl and Vogel (20) recently attempted to correlate organic D/H ratios with waters precipitated from the atmosphere in Europe and South Africa. We studied several species of plants and animals growing in close proximity in salt marsh, lagoon, and intertidal habitats utilizing the same water source. Since the D/H ratio in ocean waters from different geographic areas is nearly constant (9, 10) our samples were utilizing a common isotopic pool. Carbon isotope abundance in organisms is somewhat better understood (4, 16, 21) than hydrogen isotope abundance, so we have included carbon data for the same samples.

Sample Preparation. In order to minimize possible exchange with atmospheric water vapor, all samples were prepared, combusted, and analyzed as soon as possible after collection. Since all samples were treated much the same, any exchange of hydrogen clearly does not affect the range of values obtained. *Phyllospadix* leaves and *Mytilus* flesh were lyophilized immediately upon return to the laboratory (2 hr after collection and stored in seawater until arrival in the laboratory). Water removed from the tissue was retained for analysis. Tissue samples were air-dried at room temperature and ground in a mortar. Portions of dried samples were refluxed 4 hr with CCl₄. After filtration, solvent was removed at room temperature in a rotary evaporator. The residue, a crude lipid fraction, was scraped from the flask. Plant material after extraction with CCl₄ gave essentially the same isotopic ratio in every case as whole, dry, unextracted plant material. Carbon tetrachloride (Baker, reagent grade) left a residue of 1.9 mg/liter of solvent ($\delta D = -126\text{‰}$; $\delta^{13}C = -27.7\text{‰}$). The ratio of solvent residue to total CCl₄-extractable material varied from 0.03 to 1.20% with an average for all samples of 0.36%. While this may have altered some values slightly, no correction was made for it.

Sample Combustion. Organic materials were combusted completely to CO₂ and H₂O over copper oxide in an oxygen atmosphere at 800 to 900 C as described by Craig (4). Water was reduced to hydrogen (10) by passing it slowly over uranium turnings at 600 to 700 C. Hydrogen gas evolving from the furnace was pushed into a sample tube with an automatic Toepler pump.

Mass Spectrometer Analysis. Hydrogen samples were analyzed in a Nier 60° sector type mass spectrometer as modified by McKinney *et al.* (13) but with an additional collector tube joined

¹ This work was generously supported by the National Science Foundation, Grant BG 7517.

² Present address: Department of Botany, The University of Texas, Austin, Texas 78712.

Table I. Hydrogen and Carbon Isotope Abundance Ratios of Plants and Their Respective Lipid Fractions

	Place Collected	$\delta\text{D}\%$ SMOW		$\delta^{13}\text{C}\%$ PDBI	
		Whole plant	Lipid	Whole plant	Lipid
1. Higher plants, high $\delta^{13}\text{C}$					
<i>Zostera marina</i> ¹	Lagoon	-76.0	-179.0	-10.0	-14.4
<i>Monanthochloe littoralis</i>	High marsh	-50.0	-149.0	-15.3	-22.2
<i>Distichlis spicata</i>	High marsh	-63.0	-164.0	-14.7	-23.2
<i>Phyllospadix torreyi</i>	Beach	-17.0	-158.0	-14.0	-25.5
2. Algae					
<i>Enteromorpha marginata</i>	Low marsh	-72.0	-161.0	-16.6	-22.9
<i>Macrocystis pyrifera</i>	Beach	-70.0	-159.0	-17.5	-24.0
<i>Corallina chilense</i>	Beach	-47.0	-156.0	-18.6	-24.9
<i>Grateloupia setchellii</i>	Beach	-94.0	-167.0	-22.7	-25.2
<i>Gigartina cristata</i>	Beach	-88.0	-148.0	-20.2	-27.2
3. Higher plants, low $\delta^{13}\text{C}$					
<i>Limonium commune</i>	High marsh	-67.0	-149.0	-23.2	-28.3
<i>Salicornia bigelovii</i>	Low marsh	-82.0	-169.0	-25.2	-29.8
<i>Mesembryanthemum chilense</i>	High marsh	-55.0	-146.0	-23.6	-30.1
<i>Frankenia grandifolia</i>	Low marsh	-61.0	-150.0	-26.4	-31.1
<i>Suaeda fruticosa</i>	High marsh	-64.0	-148.0	-26.5	-32.8
Average		-65.0	-157.0	-19.6	-25.8
Water from the lagoon			-10.0		
Water from low marsh			-12.0		
Ocean water, Leo Carillo Beach			-09.0		

¹ Plants listed in order of decreasing $\delta^{13}\text{C}$ of CCl_4 -extractable material.

in such a way that it would collect the mass 3 ion beam simultaneously with the mass 2 ion beam (10). All samples were corrected for the appropriate machine background and for mixing of sample and standard gas. In addition, CO_2 samples were corrected for ^{17}O contribution to the mass 45 peak (5). Results are expressed in terms of a δ value:

$$\delta\text{D}\% = \left[\frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right] \times 1000$$

where R_{standard} is the ratio of mass 3 to mass 2 of standard mean ocean water (6). With the SMOW³ value taken as 0‰, the lagoon water is -9‰. In the case of CO_2 analysis,

$$\delta^{13}\text{C}\% = \left[\frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right] \times 1000$$

R_{standard} is the ratio of mass 45 to mass 44 in CO_2 prepared from the fossil skeleton of a Cretaceous belemnite, *Belemnitella americana*, from the Peedee formation of South Carolina; this is known as the PDB standard (5). Thus, a sample with a $\delta^{13}\text{C}$ per mil of -10 has a $^{13}\text{C}/^{12}\text{C}$ ratio less than the standard by 10 per mil, or 1.0‰. For example, the $^{13}\text{C}/^{12}\text{C}$ ratio of atmospheric CO_2 is smaller than that of the PDB standard by about 7 per mil. Therefore, the $\delta^{13}\text{C}$ per mil of this CO_2 is -7. Carbonate values (Table IV) were very close to the PDB standard. The precision of measurement is $\pm 1.0\%$ for δD and $\pm 0.1\%$ for $\delta^{13}\text{C}$.

The enrichment factor or fractionation factor between two compounds A and B, is defined (14) as:

$$\alpha = \frac{D/H_A}{D/H_B}$$

also

$$\alpha \approx \frac{1 + \delta D_A/1000}{1 + \delta D_B/1000}$$

³ Abbreviation: SMOW: standard mean ocean water.

or

$$\alpha \approx 1 + \frac{\delta D_A - \delta D_B}{1000}$$

Thus if $\delta D_A - \delta D_B = 100\%$ the fractionation factor between A and B ≈ 1.100 .

RESULTS

Heavy isotopes of both hydrogen and carbon tend to be excluded from plant matter (Table I). Fractionation factors (α) from water hydrogen to whole plant hydrogen ranged from 1.008 to 1.085 with the average 1.055. That is, organic hydrogen from whole, dried plants had 55% less deuterium than the source water. No trend in fractionation factors from ocean and lagoon plants to high marsh plants was observed. A further discrimination of about 90% was observed between the whole plant and the CCl_4 extract of plant material. Fractionation factors from whole plant to lipid ranged from 1.060 to 1.141 with an average value for α of 1.092. The fractionation from water to lipid showed less variation (1.134 to 1.169, average 1.147) than either water to organic matter or organic matter to lipid. This consistency in δD values for lipids from land plants, intertidal forms, or plants always immersed is striking when you consider that these diverse organisms may differ substantially from one another in metabolic processes. The lightest δD value obtained was -179‰ for *Zostera* lipid.

Carbon isotopes appeared to experience a smaller difference than did hydrogen isotopes between the whole plant and the lipid fraction. The fractionation factor between whole plant carbon and lipid carbon was 1.0062 (range = 1.0025-1.0115). No trends were apparent when high marsh and lagoon plants were compared. However, even taking into account differences in measurement precision, the $^{13}\text{C}/^{12}\text{C}$ ratios of total plant tissue were quite variable in contrast with hydrogen values. Not only was the well documented difference between marine and land plants apparent (4, 15), but *Suaeda* and *Distichlis*, even though growing side by side in the upper marsh, differed in $\delta^{13}\text{C}$ by 12‰.

It is, of course, possible that some plants accumulate ^{13}C -rich metabolites while others do not. The amount of lipid extracted with CCl_4 from whole, dry plants varied from 0.1% (weight lipid/dry weight whole plant) for *Grateloupia* to 3.2% for *Mesembryanthemum*, whereas the average lipid content for all plants was 1.1%. No correlation was noted between percentage of extractable lipid and variations of δD or $\delta^{13}\text{C}$ values. In contrast, Park and Epstein (15) did find a positive correlation between percentage of lipid and difference between $\delta^{13}\text{C}$ of the plant residue (after lipid extraction) and $\delta^{13}\text{C}$ of the lipid fraction. While it would be desirable to compare the same compounds in all plants, large differences (e.g., 12‰) in $\delta^{13}\text{C}$ probably do not result from an accumulation of a particular compound. Bender (2) suggests that the ^{13}C differences may indicate different carbon fixation mechanisms for the two groups of plants. She has reported several panicoid grasses to have relatively high $\delta^{13}\text{C}$ values, while festucoid grasses have low $\delta^{13}\text{C}$ values.

If we assume that source carbon for all plants reported is atmospheric carbon dioxide, -7‰ relative to the PDB standard, the fractionation factors (atmospheric carbon dioxide to whole plant carbon) fall into three groups as indicated in Table I: 1. average = 1.0065 (range 1.0030–1.0083); 2. average = 1.0121 (range 1.0096–1.0157); 3. average = 1.0180 (range 1.0162–1.0195). Each group differs from the next by nearly 6‰. The second group is composed of algae. The first group is composed of two aquatic monocots and two salt marsh grasses. Since sympatric plants were very different in $\delta^{13}\text{C}$ values, it is likely that either Bender's suggestion of a different metabolic pathway is valid or that significant differences in rates of carbon fixation exist between species. Smith and Epstein (unpublished) confirm these results for a wide variety of species. The lightest $\delta^{13}\text{C}$ value obtained was -32.8‰ for *Suaeda fruticosa* lipid.

Fleshy green leaves of *Limonium* were separated from dead, brown, basal leaves, and the tap root was removed from both. Separate analysis after drying of different plant parts (Table II) showed no large isotopic discrimination between plant organs.

Table II. Hydrogen and Carbon Isotope Abundance Ratios of Different Plant Parts

	$\delta\text{D}\text{‰}$ SMOW		$\delta^{13}\text{C}\text{‰}$ PDB _I	
	Whole organ	Lipid	Whole organ	Lipid
<i>L. commune</i> :				
Green leaves	-67.0	-149.0	-23.2	-28.3
Dead leaves	-62.0	-181.0	-23.1	-27.9
Roots	-67.0	-152.0	-24.1	-28.5
Average	-65.0	-161.0	-23.5	-28.2

Table III. Hydrogen and Carbon Isotope Abundance Ratios of Animals and Their Respective Lipid Fractions

	Place Collected	$\delta\text{D}\text{‰}$ SMOW		$\delta^{13}\text{C}\text{‰}$ PDB _I	
		Whole animal	Lipid	Whole animal	Lipid
		<i>Cypraea spadicea</i>	Beach	-71.0	-178.0
<i>Navanax inermis</i>	Lagoon	-94.0	-152.0	-14.0	-22.2
<i>Mytilus californianus</i>	Beach	-35.0	-178.0	-17.3	-22.9
<i>Dendroaster excentricus</i>	Lagoon	-75.0	-185.0	-8.6	-18.6
<i>Bulla gouldiana</i>	Lagoon	-95.0	-162.0	-9.1	-17.0
<i>Cerithidia californica</i>	Low marsh	-79.0	-171.0	-13.4	-21.6
Average		-79.0	-171.0	-13.4	-21.6

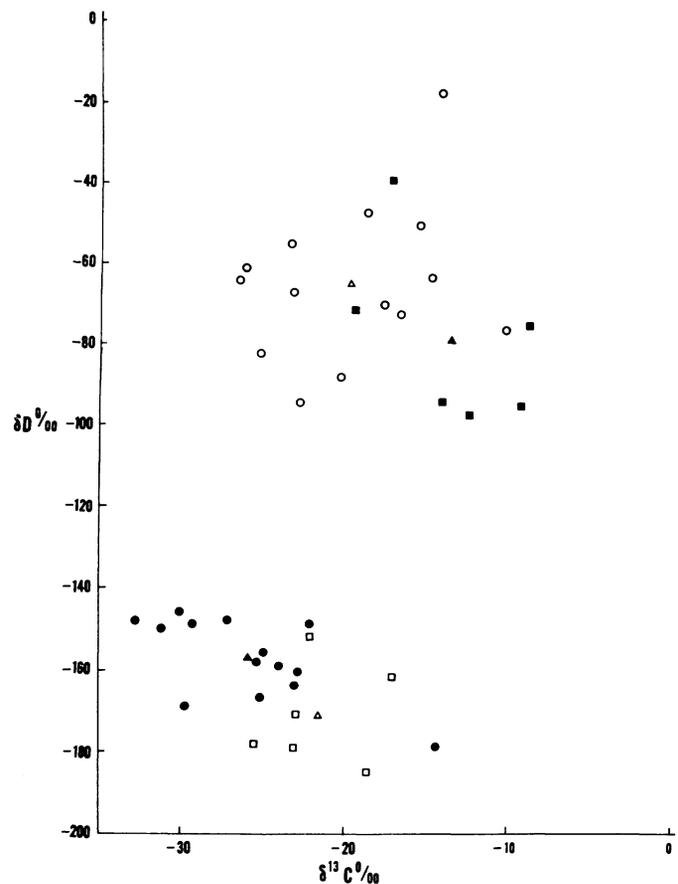


FIG. 1. Relationship between $\delta\text{D}\text{‰}$ and $\delta^{13}\text{C}\text{‰}$ for whole plant (○), whole animal (■), plant lipid (●), animal lipid (□), and average whole plant (△), average whole animal (▲), average plant lipid (◆).

Table IV. Carbon Isotope Abundance Ratios of Organic Carbonates

	$\delta^{13}\text{C}\text{‰}$ PDB _I
<i>Corallina chilense</i>	-1.4
<i>Bulla gouldiana</i>	+1.2
<i>Cerithidia californica</i>	-1.9
<i>Cypraea spadicea</i>	+1.3
<i>Dendroaster excentricus</i>	+0.3
<i>Mytilus californianus</i>	+0.6
Average	+0.02

This implies either good mixing and transport of carbon and hydrogen taken into the plant or a very constant source of isotopes.

Plants analyzed are the basis of an extensive food chain in marsh and marine environments. It would be interesting if each link in the chain were to perform a successive fractionation of hydrogen and carbon isotopes ingested. That such was not the case can be seen in Table III. Isotopically these animals nicely reflected their presumed diet. However, on the whole, animal hydrogen was lighter than plant hydrogen while animal carbon was heavier than plant carbon (Fig. 1). That carbon and hydrogen isotopic fractionation are dissimilar is not surprising, particularly in consideration of different probable modes of incorporation for the two elements.

Carbon was present not only in organic matter but also as calcium carbonate in shells of five animals and in concretions of

the calcareous red alga, *Corallina*. Carbon dioxide was liberated from carbonates with acid (8). Values in Table IV, from isotopic analysis of CO_2 , show carbonate-carbon to be compatible with published values (16).

DISCUSSION

Water is absorbed from the soil and moves through the xylem in the transpiration stream to several possible fates. Water may be extruded from hydathodes as water of guttation, but most commonly is transpired through the stomates into the atmosphere as vapor. A small amount of water taken up by plant cells is either passed through the tonoplast membrane into the vacuole or retained in the cytoplasm, itself about 70% water. Some water in the cytoplasm becomes associated with the chloroplast. Light energy absorbed by photosystem II then splits the water molecule releasing gaseous oxygen (Fig. 2). Hydrogen is released to the medium from water split in photosynthesis. NADP then takes up hydrogen from the medium (1). Hydrogen involved in reduction of carbon dioxide comes from the NADPH. Thus, there is no direct link between hydrogen released in photosynthesis and the hydrogen atoms which end up in carbohydrate. Sugars and other early products of photosynthesis are rapidly transformed by various metabolic processes to all the different compounds of which living things are composed. One of the classes of compounds is lipid.

Hydrogen isotope fractionation factors are summarized in Table V. Our results indicate that membranes discriminate very little between the isotopes of hydrogen, since the fractionation measured between environmental water and water removed from tissue by lyophilization was about 1.011. This represents a small though significant fractionation and is the sum of all processes up to the splitting of water in photosynthesis. Aquatic plants do not seem to differ greatly from higher plants in the fractionation factor for this step. If water uptake is in a steady state and is not rate-limiting, then the fractionation factor should be the same regardless of the D/H ratio of the environmental water. Indeed, similar fractionation factors have been measured for intracellular water of aquatic animals kept in distilled water, which is isotopically different from sea water (Smith and Epstein, unpublished). That part of the hydrogen which reduces CO_2 is depleted in deuterium relative to the source (vacuolar and cytoplasmic water). Thus, the splitting of water in photosynthesis has a frac-

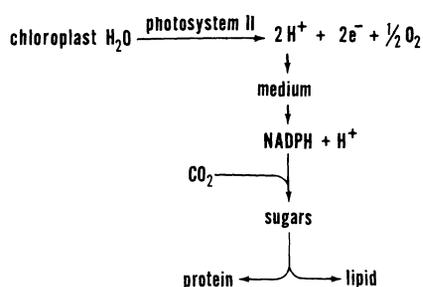


FIG. 2. Hydrogen metabolism in plants.

Table V. Summary of Hydrogen Isotope Fractionation Factors for Salt Marsh and Marine Plants

System	Fractionation Factor (α) \pm 0.005
Extracellular water \rightleftharpoons intracellular water	1.011
Intracellular water \rightleftharpoons photosynthate	1.044
Photosynthate \rightleftharpoons lipid	1.092

tionation factor of 1.044. This indicates a difference in δD between water in the chloroplast and carbohydrate produced via photosynthesis. Transport of hydrogen within the chloroplast is actively being studied (7, 12, 19) as a vital part of the photosynthetic process. Key kinetic steps probably account for observed fractionations of the natural isotopes. Another large fractionation occurs during lipid synthesis. A fractionation of 1.092 occurs between carbohydrate and lipid. It has been suggested (11, 17) that NADPH contributes the large part of hydrogen during lipid synthesis. If our suggestion for isotope fractionation is correct, there could exist two completely different pools of NADPH—one associated with photosynthesis, the other produced metabolically and used in lipid synthesis. Thus, either the cofactor or the dehydrogenases would have to be strictly compartmentalized. Another possibility during lipid biosynthesis is that precursor molecules themselves contribute hydrogen depleted in deuterium. The fact that lipids have low $\delta^{13}\text{C}$ values as well as low δD values would strengthen this idea. The isotopic fractionation between total organic matter and fatty acids is considerably larger than the isotopic fractionation between whole tissue and the CCl_4 extract (Smith and Epstein, unpublished).

The δD values reported here agree with values reported for natural organic substances (2, 10, 20, 21). The remarkable similarity in δD values indicate that both marine and salt marsh plants enjoyed a common water source. *Salicornia* collected in December were the dry, brown remains of annual plants dead since the previous fall. However, these plants were isotopically very similar to sympatric species alive at the time of collection. The isotopic record was bound into plant material with little apparent change by death and subsequent decay. This may mean that, as Wickman (21) suggested, isotopic analysis of plants long dead (*e.g.*, herbarium specimens) or even fossil organic matter may tell us something of the environment or metabolism of the organism.

Acknowledgments—We wish to thank Dr. Ian Kaplan and Dr. John Warme of the Geology Department, University of California, Los Angeles, for assistance in making collections at Point Mugu. Discussions of our data with Dr. Kaplan have also been most helpful. The technical assistance of Mr. Joop Goris and Mrs. Jane Young is gratefully acknowledged.

LITERATURE CITED

- ALEEM, M. I. H. 1965. Path of carbon and assimilatory power in chemosynthetic bacteria. I. *Nitrobacter agilis*. Biochim. Biophys. Acta 107: 14–28.
- BENDER, M. M. 1968. Mass spectrometric studies of carbon 13 variations in corn and other grasses. Amer. J. Sci. Radiocarbon Suppl. 10: 468–472.
- BOKHOVEN, D. AND H. H. J. THEEUWEN. 1956. Deuterium content of some natural organic substances. Koninklijke Nederlandse Akademie van Wetenschappen te Amsterdam. Ser. B 59: 78–83.
- CRAIG, H. 1953. The geochemistry of the stable carbon isotopes. Geochim. Cosmochim. Acta 3: 53–92.
- CRAIG, H. 1957. Isotopic standards for carbon and oxygen and correction factors for mass-spectrometric analysis of carbon dioxide. Geochim. Cosmochim. Acta 12: 133–149.
- CRAIG, H. 1961. Standard for reporting concentrations of deuterium and oxygen-18 in natural waters. Science 133: 1833–1834.
- DILLEY, R. A. AND L. P. VERNON. 1967. Quantum requirement of the light-induced proton uptake by spinach chloroplasts. Proc. Nat. Acad. Sci. U.S.A. 57: 395–400.
- EPSTEIN, S., R. BUCHSBAUM, H. LOWENSTAM, AND H. C. UREY. 1951. Carbonate-water isotopic temperature scale. Bull. Geol. Soc. Amer. 62: 417–426.
- EPSTEIN, S. AND T. MAYEDA. 1953. Variation of ^{18}O content of waters and other substances. Geochim. Cosmochim. Acta 4: 89–103.
- FRIEDMAN, I. 1953. Deuterium content of natural waters and other substances. Geochim. Cosmochim. Acta 4: 89–103.
- KATZ, J. AND R. ROGNSTAD. 1966. The metabolism of tritiated glucose by rat adipose tissue. J. Biol. Chem. 241: 3600–3610.
- LYNN, W. S. 1968. Changes in internal hydrogen concentration associated with photophosphorylation in intact and sonically treated chloroplasts. J. Biol. Chem. 243: 1060–1064.
- McKINNEY, C. R., J. M. MCCREA, S. EPSTEIN, H. A. ALLEN, AND H. C. UREY. 1950. Improvements in mass spectrometers for the measurement of small differences in isotope abundance ratios. Rev. Sci. Instrum. 21: 724–730.
- PARK, R. AND S. EPSTEIN. 1960. Carbon isotope fractionation during photosynthesis. Geochim. Cosmochim. Acta 21: 110–126.

15. PARK, R. AND S. EPSTEIN. 1961. Metabolic fractionation of C^{13} and C^{12} in plants. *Plant Physiol.* 36: 133-138.
16. PARKER, P. L. 1964. The biogeochemistry of the stable isotopes of carbon in a marine bay. *Geochim. Cosmochim. Acta* 28: 1155-1164.
17. RAGLAND, T. E. AND D. P. HACKETT. 1965. Radioactive tracer studies of the metabolic fates of intracellularly generated NADH and NADPH in higher plant tissues. *Plant Physiol.* 40: 1191-1197.
18. RITTENBERG, D. 1963. The effect of isotopic substitution on the living cell. *J. Chem. Phys.* 60: 318-323.
19. RUMBERG, B. AND U. SIGGEL. 1969. pH changes in the inner phase of the thylakoids during photosynthesis. *Naturwissenschaften* 56: 130-132.
20. SCHIEGEL, W. E. AND J. C. VOGEL. 1970. Deuterium content of organic matter. *Earth and Planetary Science Letters* 7: 307-313.
21. WICKMAN, F. E. 1952. Variations in the relative abundance of the carbon isotopes in plants. *Geochim. Cosmochim. Acta* 2: 243-253.
22. ZBOROWSKI, G., L. PONTICORVO, AND D. RITTENBERG. 1967. On the constancy of deuterium fractionation in the biosynthesis of fatty acids since the Miocene period. *Proc. Nat. Acad. Sci. U.S.A.* 58: 1660-1663.