

Supporting Information for:

**Electrochemical Transformation of Trace Organic
Contaminants in Latrine Wastewater**

Justin T. Jasper, Oliver S. Shafaat, Michael R. Hoffmann

42 pages
13 figures
2 tables

Reaction Rate Constants for FAC and NH_2Cl . Test compound (100 nM) removal was followed in buffered solutions (pH 8.7; 20 mM borate) with excess FAC (0.03-5.0 mM) or NH_2Cl (0.3-3.0 mM). NH_2Cl solutions were prepared by adding stock solutions of FAC dropwise to excess NH_4Cl (1.2:1 $\text{NH}_4\text{Cl}:\text{HOCl}$). Samples were quenched (45 mM $\text{Na}_2\text{S}_2\text{O}_3$) prior to analysis. Solution pH values and total chlorine concentrations did not change significantly over the course of the experiments. Rate constants were calculated based on first-order removal kinetics of the test compounds using measured steady-state total chlorine concentrations. For ciprofloxacin, rate constants were measured at multiple NH_2Cl and FAC concentrations to verify that observed ciprofloxacin transformation rates were not dependent on oxidant concentrations, but rather represented decay of the rapidly formed N-chlorinated ciprofloxacin intermediate as previously reported.¹ First-order ciprofloxacin decay rates were therefore reported.

Reaction Rate Constants for the Chlorine Radical Anion, $\cdot\text{Cl}_2^-$. Second-order rate

constants for the reaction of test compounds with the dichlorine radical anion ($\cdot\text{Cl}_2^-$) were measured by nanosecond transient absorption laser flash photolysis of solutions containing $\text{Na}_2\text{S}_2\text{O}_8$ (25 mM), NaCl (100 mM), and test compounds (0-100 μM). Excitation at 266 nm (8 ns FWHM, 10 Hz repetition rate) produced $\cdot\text{SO}_4^-$ that rapidly reacted with Cl^- to produce $\cdot\text{Cl}_2^-$ ($\sim 1 \mu\text{M}$). Solution pH values ranged from 5.5 to 6.0. The decay of $\cdot\text{Cl}_2^-$ was monitored at 340 nm over 10 ms, and data was averaged over 10 shots. $\cdot\text{Cl}_2^-$ decay was log-normalized and plotted versus test compound concentrations to determine second-order rate constants. $\cdot\text{Cl}_2^-$ decay was log-normalized and fit according to:

$$A(340) = fe^{-gt} + h \quad (\text{S1})$$

where $A(340)$ is the absorbance at 340 nm, f is the initial absorbance, g is the decay rate, and h is an offset due to a longer lived species formed after $\cdot\text{Cl}_2^-$ decay.² $\cdot\text{Cl}_2^-$ decay rates were plotted versus test compound concentrations to determine second-order rate constants:

$$g = k_{\text{cmpd}, \cdot\text{Cl}_2^-} [\text{cmpd}] + b \quad (\text{S2})$$

where $k_{\text{cmpd}, \cdot\text{Cl}_2^-}$ is the second-order rate constant for the reaction of a test compound with $\cdot\text{Cl}_2^-$, $[\text{cmpd}]$ is the test compound concentration, and b is the y-intercept that represents loss of $\cdot\text{Cl}_2^-$ due to other reactions (e.g., quenching by H_2O). Under these conditions $\cdot\text{Cl}$ is predicted to contribute less than 5% to measured second-order rate constants greater than $10^7 \text{ M}^{-1} \text{ s}^{-1}$.

Chronoamperometric Experiments. Chronoamperometric experiments were

performed to verify direct electron transfer for compounds that underwent transformation in the absence of Cl^- and were soluble enough dissolve rapidly (i.e., ranitidine and cimetidine). About 5 mM of a solid test compound was added to a solution of Na_2SO_4 (20 mM) undergoing potentiostatic electrolysis (3.5 V cathode vs. anode), and the current response was observed. A spike in the current upon test compound addition implied that the compound could be oxidized directly at the anode.^{3,4} The possibility that current increased due to a change in electrolyte strength was eliminated based on the current's return to near baseline values after test compound oxidation was complete.

Transformation Product Identification. Test compound transformation product

formulae were calculated based on accurate masses and typically had a mass error of less than 2 mDa (Table SI 2). Transformation product halogen content was confirmed by characteristic isotopic patterns using the $m/z+2$, $m/z+4$, etc. peaks. Isotopic peak ratios were typically within 10% of theoretical values for the proposed formulae. Transformation product structures were proposed based on the literature when possible (see bold references in Table SI 2), by comparing exact mass values and fragment ions. In other cases, structures were proposed simply based on determined formulae. In these cases, the exact position of certain functional groups (e.g., hydroxyl and chlorine) could not be determined. When possible, proposed structures were verified with authentic standards, as specified in Table SI 2.

Analytical Methods. Dissolved organic carbon and dissolved inorganic carbon were measured by persulfate digestion using an Aurora TOC analyzer.⁵ NO_3^- , Cl^- , PO_4^{3-} , SO_4^{2-} , and NH_4^+ were analyzed by ion chromatography (Dionex ICS 2000; AS19G anions, CS12A cations).⁵

Test compounds were separated by an Acquity BEH C18 column (2.1x50 mm; 1.7 μm particles), eluted at 0.5 mL min^{-1} using acetonitrile and water with 0.1% formic acid and 1% acetonitrile. The following gradient was used: 0 min, 5% acetonitrile; 0.2 min, 5% acetonitrile; 3.2 min, 95% acetonitrile; 3.5 min, 95% acetonitrile; 3.6 min, 5% acetonitrile; 5 min, 5% acetonitrile. Column temperature was held at 35 °C. pCBA were detected by UV absorbance (270 nm). Trace organic compounds were detected by positive electrospray ionization (ESI+) in resolution mode with a capillary voltage of 0.2 kV, a cone voltage of 50 V, and a source offset of 80 V. Source temperature was 120 °C and desolvation temperature was 400 °C. Cone gas was 40 L/h and desolvation gas was 800 L/h. Scan time was 0.3 s in continuum mode. A collision energy of 1.0 eV was used. A second acquisition channel scanned collision energy from 0-30 eV for fragment identification. Accurate mass values were corrected by the software using a leucine lock-mass.

101 **Calculation of $\cdot\text{Cl}_2^-$ Reaction with Propranolol.** The fraction of $\cdot\text{Cl}_2^-$ reacting with
 102 propranolol ($F_{\cdot\text{Cl}_2^-, \text{propranolol}}$) in latrine wastewater (100 mg L⁻¹ TOC) was calculated using $\cdot\text{Cl}_2^-$
 103 measured second-order reaction rate constants with propranolol ($k_{\cdot\text{Cl}_2^-, \text{propranolol}}$) and TOC
 104 ($k_{\cdot\text{Cl}_2^-, \text{TOC}}$) to be:

$$\begin{aligned}
 F_{\cdot\text{Cl}_2^-, \text{propranolol}} &= \frac{k_{\cdot\text{Cl}_2^-, \text{propranolol}}[\text{propranolol}]}{k_{\cdot\text{Cl}_2^-, \text{propranolol}}[\text{propranolol}] + k_{\cdot\text{Cl}_2^-, \text{TOC}}[\text{TOC}]} \\
 &= \frac{1.9 \times 10^9 \text{ M}^{-1} \text{ s}^{-1} * 1 \times 10^{-6} \text{ M}}{1.9 \times 10^9 \text{ M}^{-1} \text{ s}^{-1} * 1 \times 10^{-6} \text{ M} + 1.9 \times 10^3 (\text{mg L}^{-1})^{-1} \text{ s}^{-1} * 100 \text{ mg L}^{-1}} \\
 &= 0.0099
 \end{aligned}$$

105

Test Compound Transformation Products. Carbamazepine's predominant

transformation products were confirmed to be acridine, hydroxyacridine, and acridine-9-carboxaldehyde (Table SI 2; Figures SI 12a and 13), based on authentic standards. No chlorinated transformation products were identified during latrine wastewater or buffered Cl⁻ solution electrolysis. Transformation products were similar to those previously observed during oxidation and chlorination of carbamazepine.⁶⁻¹⁰

Metoprolol and propranolol were predominantly transformed to chlorinated products that underwent further transformation to form products not detected by LCMS (Figures SI 12b,c and 9). Hydroxylated and multiply halogenated transformation products were also identified (Table SI 2). Most metoprolol transformation products were identified previously during reverse osmosis retentate electrolysis¹¹ or other treatment processes.^{12,13} Some propranolol transformation products observed in this study have been reported previously,¹⁴⁻¹⁷ but others have not, including the predominant chlorinated product (m/z 294.1276 amu; m/z+2 amu observed at ~33%).

Identified ciprofloxacin transformation products were typically transformed at the piperazine group rather than chlorinated (Figures SI 12d, 13), as previously observed during ciprofloxacin reaction with FAC and ClO₂.^{1,18}

Trimethoprim was transformed into a variety of hydroxylated and chlorinated transformation products, some of which have been identified during treatment using ferrate,¹⁹ activated sludge,²⁰ and FAC.²¹

Only a few significant acetaminophen and cimetidine transformation products were observed (Figures SI 12f and 13), all of which are produced by reaction with FAC.²²⁻²⁴ As with

128 trimethoprim, different acetaminophen transformation products were observed in the absence of
129 NH_4^+ , including formation of dichlorinated acetaminophen.

130 Ranitidine transformation products were chlorinated, demethylated, and oxidized, with
131 the latter two transformations likely occurring at the furan group.²⁵ Although a few of the
132 identified products have previously been observed during chloramination of ranitidine,²⁵ further
133 work is needed to confirm transformation product structures (Table SI 2). Many transformation
134 products observed during latrine wastewater electrolysis were not observed during electrolysis of
135 buffered Cl^- solutions (Figures SI 12g and 13; see main text).

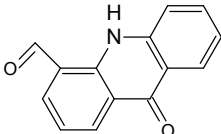
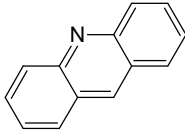
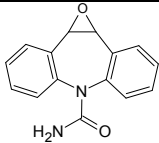
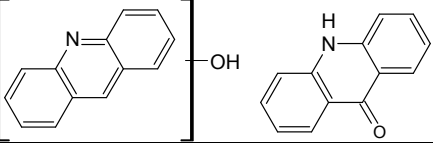
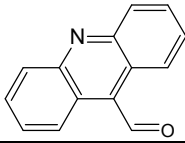
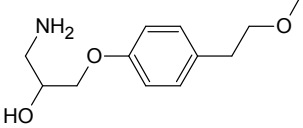
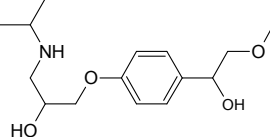
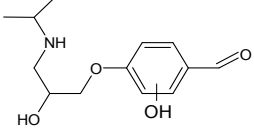
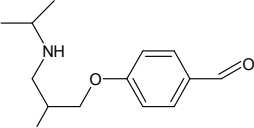
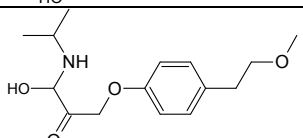
Table SI 1. Comparison of Measured Test Compound Reaction rates with HOCl and NH₂Cl to Literature Values.

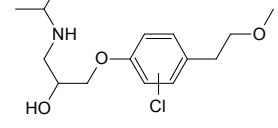
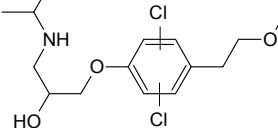
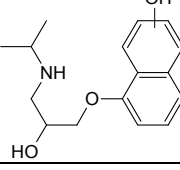
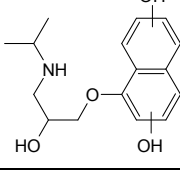
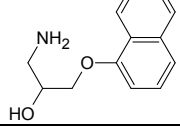
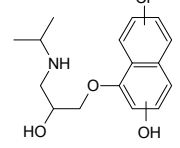
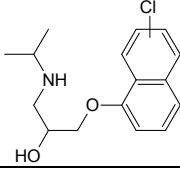
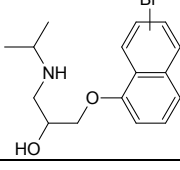
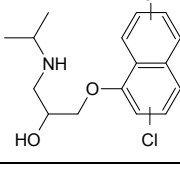
compound	property					
	$k_{\text{FAC}} (\text{M}^{-1}\text{s}^{-1})^{a,b}$		$k_{\text{NH}_2\text{Cl}} (\text{M}^{-1}\text{s}^{-1})^a$		$k_{\text{Cl}_2} (\text{M}^{-1}\text{s}^{-1})$	
	measured	literature	measured	literature	measured	literature
carbamazepine	$8.9(\pm 1.0) \times 10^{-2}$	$2.7 \times 10^{-2} \text{ }^{(10)}$	$< 3 \times 10^{-3}$	n.a. ^c	$\sim 1 \times 10^5$	$6.7 \times 10^4 \text{ }^{(14)}$
metoprolol	$1.9(\pm 1.0) \times 10^{-2}$	$2 \times 10^{-2} \text{ }^{(16)}$	$< 4 \times 10^{-3}$	$< 0.1 \text{ }^{(16)}$	$\sim 4 \times 10^3$	$0.6\text{-}1.4 \times 10^3 \text{ }^{(142)}$
propranolol	$0.27(\pm 0.02)$	$0.3 \text{ }^{(16)}$	$< 6 \times 10^{-3}$	$< 0.1 \text{ }^{(16)}$	$\sim 2 \times 10^5$	$3.3 \times 10^5 \text{ }^{(16)}$
ciprofloxacin	$5.6(\pm 0.4) \times 10^{-4} \text{ s}^{-1} \text{ }^e$	$7 \times 10^{-4} \text{ s}^{-1} \text{ }^{(1) e}$	$9 \times 10^{-4} \text{ s}^{-1} \text{ }^e$	$7 \times 10^{-4} \text{ s}^{-1} \text{ }^{(1) e}$	$\sim 6 \times 10^3$	$4.3 \times 10^3 \text{ }^{(1)}$
trimethoprim	$6.7(\pm 0.2)$	$8 \text{ }^{(21)}$	$< 1 \times 10^{-3}$	negligible ⁽²¹⁾	$\sim 3 \times 10^6$	$2 \times 10^5\text{-}2 \times 10^6 \text{ }^{(21)}$
acetaminophen	$7.0(\pm 0.4) \times 10^2$	$0.7\text{-}1.4 \times 10^2 \text{ }^{(16,23,27)}$	$0.9(\pm 0.1)$	$1.1 \times 10^{-2} \text{ }^{(16)}$	$\sim 3 \times 10^4$	n.a. ^c
ranitidine	$> 8 \times 10^3$	n.a. ^c	$11(\pm 1)$	$12 \text{ }^{(25)}$	$\sim 8 \times 10^4$	n.a. ^c
cimetidine	$> 6.5 \times 10^3$	$> 1 \times 10^3 \text{ }^{(24) g}$	$47(\pm 1)$	$48 \text{ }^{(24)}$	$\sim 4 \times 10^6$	n.a. ^c

^a Measured or calculated at pH 8.7. ^b FAC=[HOCl]+[OCl⁻]. ^c n.a.: not available. ^d Calculated at pH 1.3 based on reported acid catalyzed reactions with HOCl.

^e First-order transformation rate is reported due to rapid quenching of the N-chlorinated ciprofloxacin intermediate by Na₂S₂O₃. ^f Reaction rate between HOCl and the predominant fully protonated form of ciprofloxacin at pH 1.3. ^g Estimated based on complete removal within 10 s.

153 Table SI 2. Test Compound Transformation Products (TPs).

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Carbamazepine	224.0715 (Cbz 224)	164.920, 167.073, 180.081, 196.076	0.58	C ₁₄ H ₁₀ NO ₂	0.3		6,7
	180.0812 ^d (Cbz 180)	152.0621	1.17	C ₁₃ H ₁₀ N	0.3		6,8
	275.0800 (Cbz 275)	180.0800, 210.0919, 236.0712, 253.0976	1.58	C ₁₄ H ₁₃ NO ₅	0.6		
	253.0974 ^d (Cbz 253)		1.59	C ₁₅ H ₁₃ N ₂ O ₂	0.3		6,10
	196.0759 ^e (Cbz 196)	167.073, 180.0810	1.65	C ₁₃ H ₁₀ NO	0.3		6-9
	208.0761 ^d (Cbz 208)	152.062, 180.082, 196.0760	2.07	C ₁₄ H ₁₀ NO	0.1		8,9
Metoprolol	226.1436 (Met 226)	116.1067, 121.0636, 149.0583	0.82	C ₁₂ H ₂₀ NO ₃	0.7		11,13
	284.1853 (Met 284)	116.1063, 226.1420, 268.141	0.85	C ₁₅ H ₂₆ NO ₄	0.9		11,13
	254.1393 (Met 254)	105.0692, 151.0381, 177.0540, 212.0911, 236.1275	0.93	C ₁₃ H ₂₀ NO ₄	<0.1		
	238.1439 (Met 238)	105.0696, 133.0642, 149.0590, 161.0589, 196.0962	1.04	C ₁₃ H ₂₀ NO ₃	0.4		12
	282.1706 (Met 282)	105.0696, 133.0642, 149.0590, 161.0589, 196.0962	1.05	C ₁₅ H ₂₄ NO ₄	<0.1		11

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Metoprolol (cont.)	302.1515 (Met 302)	98.0963, 116.1066, 167.0249, 193.0407, 260.1044, 284.1411	1.51	C ₁₅ H ₂₅ NO ₃ Cl	0.8		11
	336.1121 (Met 336)	72.0817, 98.0965, 116.1064, 276.0549, 294.0649, 318.102	1.62	C ₁₅ H ₂₄ NO ₃ Cl ₂	1.2		11
Propranolol	276.1599 (Pro 276)	116.1074	1.0-1.3 ^f	C ₁₆ H ₂₂ NO ₃	<0.1		17
	292.1545 (Pro 292)	116.107, 123.092, 150.128, 274.144, 272.128,	1.29	C ₁₆ H ₂₂ NO ₄	0.4		15
	218.1180 (Pro 218)	116.1071, 183.0804	1.45	C ₁₃ H ₁₆ NO ₂	0.1		16
	310.1210 (Pro 310)	72.0816, 116.1071, 135.1038, 260.1653	1.53	C ₁₆ H ₂₁ NO ₃ Cl	<0.1		
	294.1276 (Pro 294)	150.128, 260.165	1.85	C ₁₆ H ₂₁ NO ₂ Cl	1.5		
	338.0739 (Pro 338)	116.1070, 182.0725, 217.0417	1.94	C ₁₆ H ₂₁ NO ₂ Br	1.7		
	328.0868 (Pro 328)	72.0819, 116.1068, 219.1751	1.98	C ₁₆ H ₂₀ NO ₂ Cl ₂	0.3		

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Ciprofloxacin	306.1246 (Cfx 306)	217.0404, 268.1077, 288.1142	1.09	C ₁₅ H ₁₇ N ₃ O ₃ F	0.8		1,18
	366.1012 (Cfx 366)	225.0213, 245.1079, 279.0695, 322.1108	1.22	C ₁₇ H ₁₈ N ₃ O ₃ FCl	0.9		1
	362.1141 (Cfx 362)	344.1037	1.29	C ₁₇ H ₁₇ N ₃ O ₅ F	1.1		18
	291.0773 (Cfx 291)	245.0709, 273.0671	1.42	C ₁₄ H ₁₂ N ₂ O ₄ F	0.8		
	263.0828 (Cfx 263)	204.032, 245.0724	1.49	C ₁₃ H ₁₂ N ₂ O ₃ F	0.4		18
	298.0287 (Cfx 298)	280.0184	1.63	C ₁₃ H ₁₀ NO ₄ FCl	0.5		1
	289.0981 (Cfx 289)	271.0876	1.74	C ₁₅ H ₁₄ N ₂ O ₃ F	0.8		
Trimethoprim	342.1327 (Tmp 342)	273.0978, 307.1397	0.98	C ₁₄ H ₂₁ N ₅ O ₃ Cl	0.6		
	307.1391 (Tmp 307)	209.1644, 212.0686, 240.0634, 258.0742, 273.0975	0.98	C ₁₄ H ₁₉ N ₄ O ₄	1.5		19
	322.1508 (Tmp 322)		1.04	C ₁₄ H ₂₀ N ₅ O ₄	0.7		
	325.1499 (Tmp 325a)	173.0813, 219.0872, 233.1043, 293.1229	1.05	C ₁₄ H ₂₁ N ₄ O ₅	1.3		20
	343.1164 (Tmp 343)	181.0852	1.08	C ₁₄ H ₂₀ N ₄ O ₄ Cl	0.9		
Trimethoprim							

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
(cont.)	359.1125 (Tmp 359a)	215.0465, 240.0414, 236.0905	1.19	C ₁₄ H ₂₀ N ₄ O ₅ Cl	0.3		
	323.1345 (Tmp 323)		1.26	C ₁₄ H ₁₉ N ₄ O ₅	1.0		
	377.0773 (Tmp 377)	217.0435, 236.0913, 255.0416, 257.0388, 314.0898	1.26	C ₁₄ H ₁₉ N ₄ O ₄ Cl ₂	1.0		21
	393.0719 (Tmp 393)	100.0503, 126.0529, 249.0075, 270.0524	1.32	C ₁₄ H ₁₉ N ₄ O ₅ Cl ₂	1.4		
	325.1059 (Tmp 325b)	145.0267, 271.0918, 249.008	1.39	C ₁₄ H ₁₈ N ₄ O ₃ Cl	0.8		21
	411.0391 (Tmp 411)	249.0080, 366.0616	1.39	C ₁₄ H ₁₈ N ₄ O ₄ Cl ₃	0.3		21
	369.0558 (Tmp 369)	255.0886, 273.092, 325.1078, 339.0081	1.42	C ₁₄ H ₁₈ N ₄ O ₃ Br	0.4		
	359.0672 (Tmp 359b)	181.0854, 199.097, 289.0480, 298.0371, 329.0196	1.55	C ₁₄ H ₁₇ N ₄ O ₃ Cl ₂	0.6		21
	403.0174 (Tmp 403)	140.0686, 180.8981, 257.1478, 359.0669	1.58	C ₁₄ H ₁₇ N ₄ O ₃ BrCl	0.1		
Acetaminophen	186.0312 (Ace 186)	109.0515, 144.0201, 155.9838	1.17	C ₈ H ₉ NO ₂ Cl	1.0		22,23
	219.9919 (Ace 219)	143.0123, 177.9810	1.42	C ₈ H ₈ NO ₂ Cl ₂	1.3		22,23
Ranitidine	338.0561 (Rnt 338)	109.0278, 186.0565, 202.9997, 214.0240, 292.0612	0.63	C ₁₁ H ₁₇ N ₃ O ₅ SCl	1.6		25

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
	336.0419 (Rnt 336)	125.0600, 181.0201, 203.0023, 214.0259, 290.0497	0.66	C ₁₁ H ₁₅ N ₃ O ₅ SCl	0.2		
	364.1194 (Rnt 364)	109.0278, 163.0854, 189.1012, 258.0579	0.76	C ₁₃ H ₂₃ N ₅ O ₃ SCl	1.6		25
	335.0928 (Rnt 335)		0.85	C ₁₂ H ₂₀ N ₄ O ₃ SCl	1.6		
	349.1083 (Rnt 349)	124.075, 132.0438, 164.0159, 210.0092, 223.0893	0.88	C ₁₃ H ₂₂ N ₄ O ₃ SCl	1.8		25
	270.0903 (Rnt 270)	97.0759, 165.1013, 191.1167, 224.0971	1.02	C ₁₁ H ₁₆ N ₃ O ₃ S	0.9		
	322.061 (Rnt 322)	165.0236, 240.1152	1.08	C ₁₁ H ₁₇ N ₃ O ₄ SCl	1.8		
	320.0459 (Rnt 320)	109.028, 165.0238, 208.0392, 234.0191, 274.0529	1.12	C ₁₁ H ₁₅ N ₃ O ₄ SCl	1.3		
	294.12575 (Rnt 294)	109.0294, 150.1284, 177.0489, 188.0054, 275.053	1.85	C ₁₃ H ₂₅ NO ₂ SCl	3.7		
Cimetidine	117.0210 (Cmt 117)	90.0104	0.53	C ₄ H ₆ N ₂ Cl	1.0		24
	189.0435 (Cmt 189)	82.0403	1.06	C ₅ H ₉ N ₄ O ₂ S	1.1		24

^a Compounds with m/z ratios in bold exhibited isotopic patterns consistent with the halogen atoms specified in the calculated formula (i.e., m/z+2, m/z+4, etc. peaks). ^b Structure of unprotonated TP. Structure based on calculated formula, fragments, and literature. ^c References in bold proposed compound structure shown. References in italics observed similar major fragment ions. ^d TP identity confirmed based on authentic standard retention time and mass spectrum. ^e Authentic acridone standard (ketone product shown) retention time differed by ~0.03 min and had different fragmentation pattern, suggesting that hydroxyacridone was formed during electrolysis. ^f Multiple peaks observed.

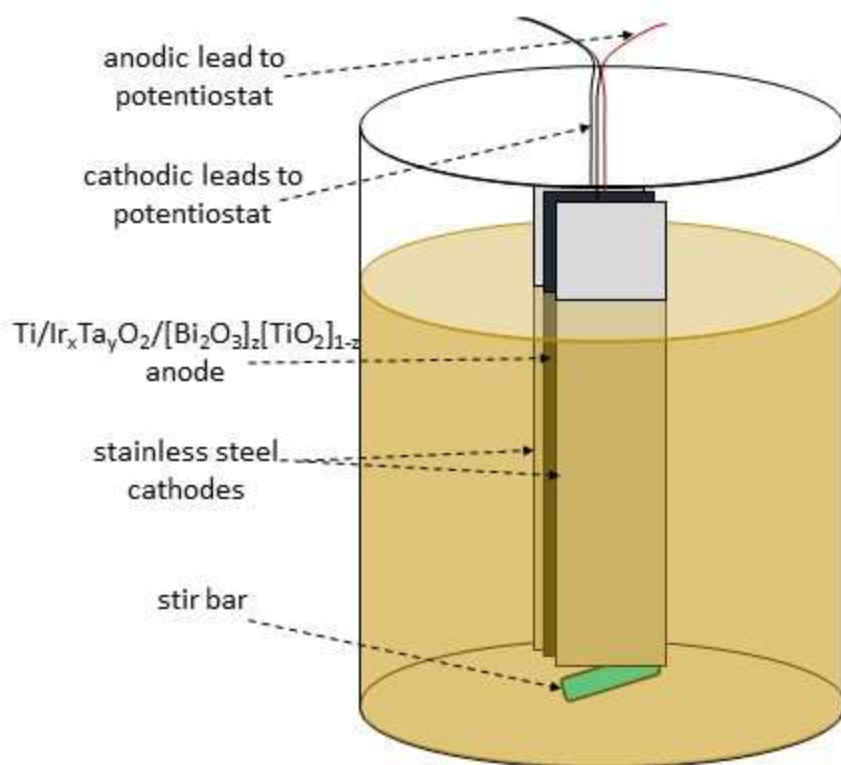
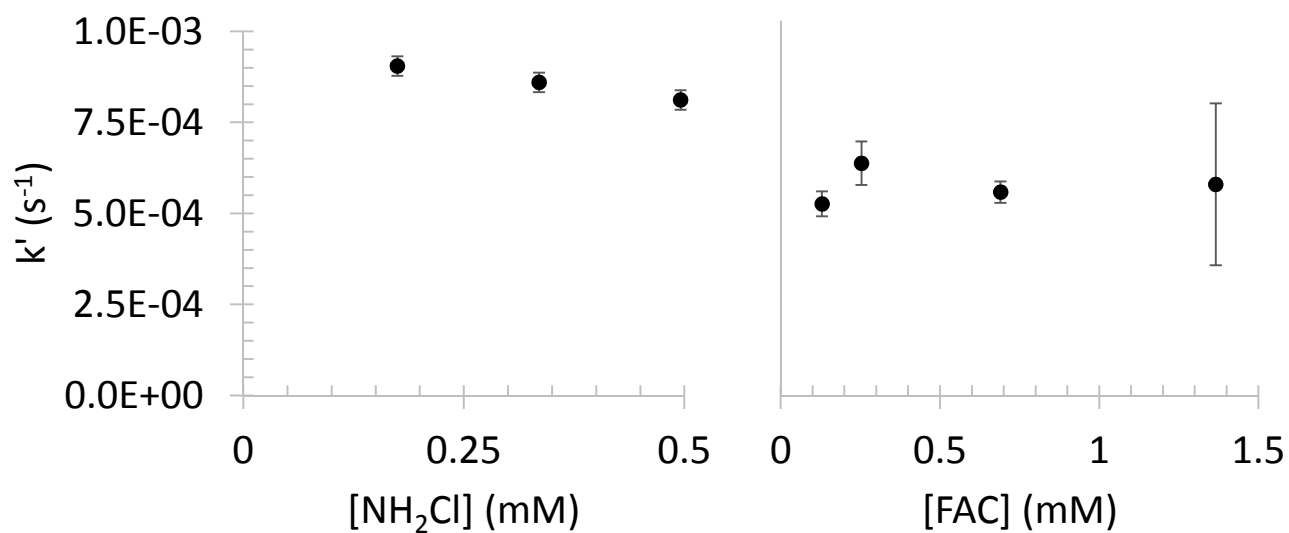


Figure SI 1. Schematic of electrochemical reactor used in this study.



165 Figure SI 2. Measured first-order rate constants for the reaction of ciprofloxacin with various
166 concentrations of NH_2Cl and FAC. Error bars represent \pm one standard deviation.

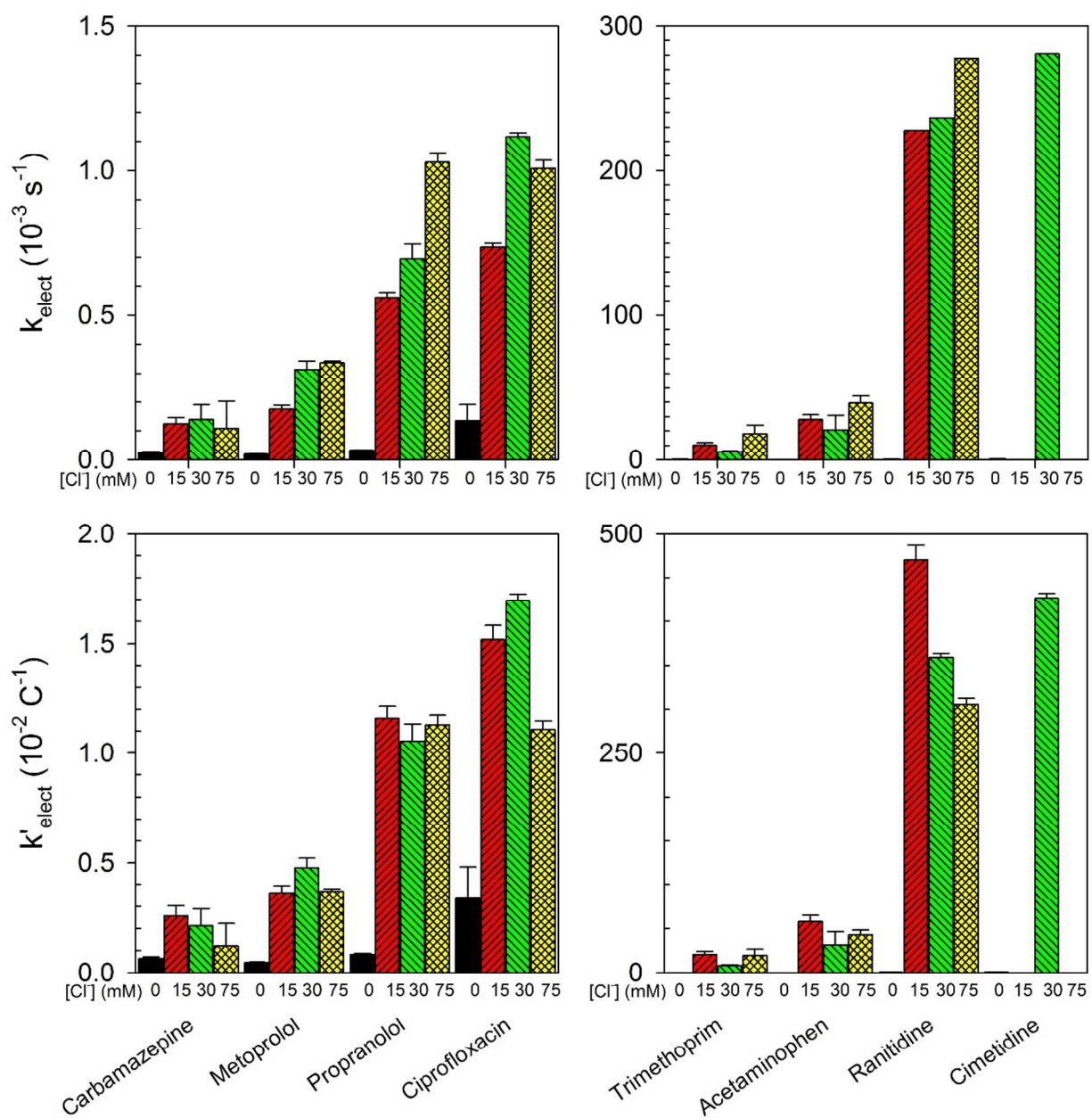


Figure SI 3. Pseudo-first order test compound electrolysis rates (k ; top) and rates normalized for charge (k' ; bottom). Solutions were electrolyzed at an applied potential of 3.5 V in 0-75 mM NaCl solution buffered at pH 8.7 (20 mM borate). Average current densities: 0 mM, 0.6 A L⁻¹; 15 mM, 0.7 A L⁻¹; 30 mM, 0.9 A L⁻¹; 75 mM, 1.3 A L⁻¹. FAC production rates: 15 mM, 0.9 $\mu\text{M s}^{-1}$; 30 mM, 1.6 $\mu\text{M s}^{-1}$; 75 mM, 3 $\mu\text{M s}^{-1}$. Error bars represent \pm one standard deviation.

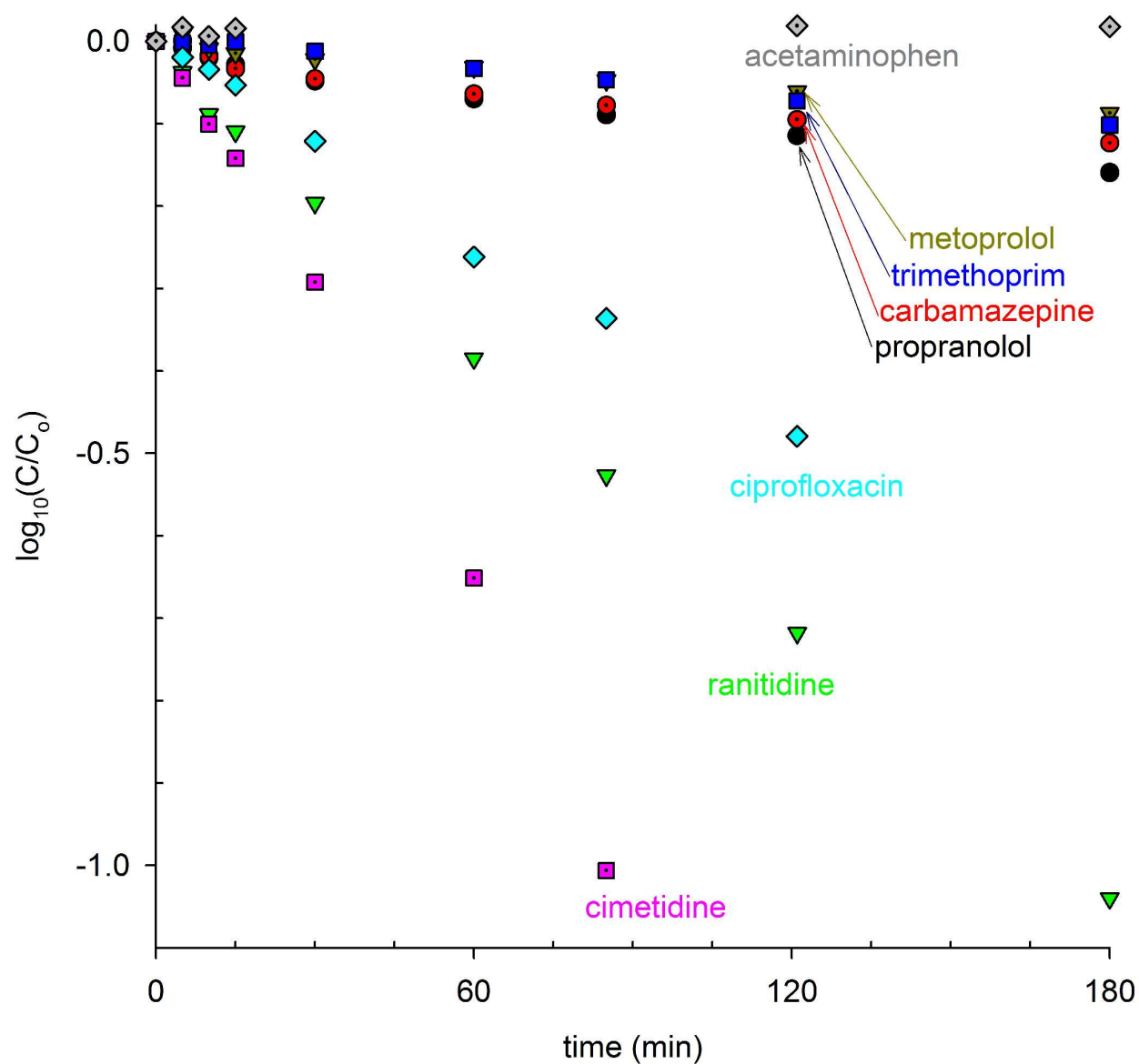
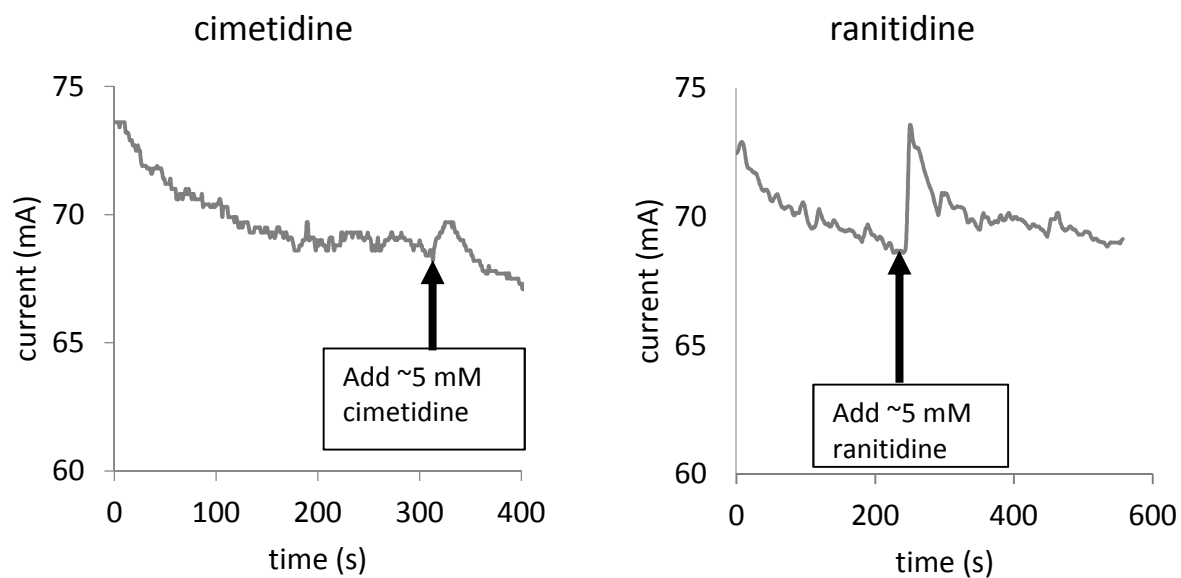


Figure SI 4. Test compound removal during electrolysis (3.5 V; 0.6 A L⁻¹) in water buffered at pH 8.7 (20 mM borate).



178
179 Figure SI 5. Current during addition of solid test compounds to solutions of Na_2SO_4 (20 mM)
180 undergoing electrolysis (3.5 V). Spikes in current upon test compound addition imply direct
181 electron transfer from the compound.
182

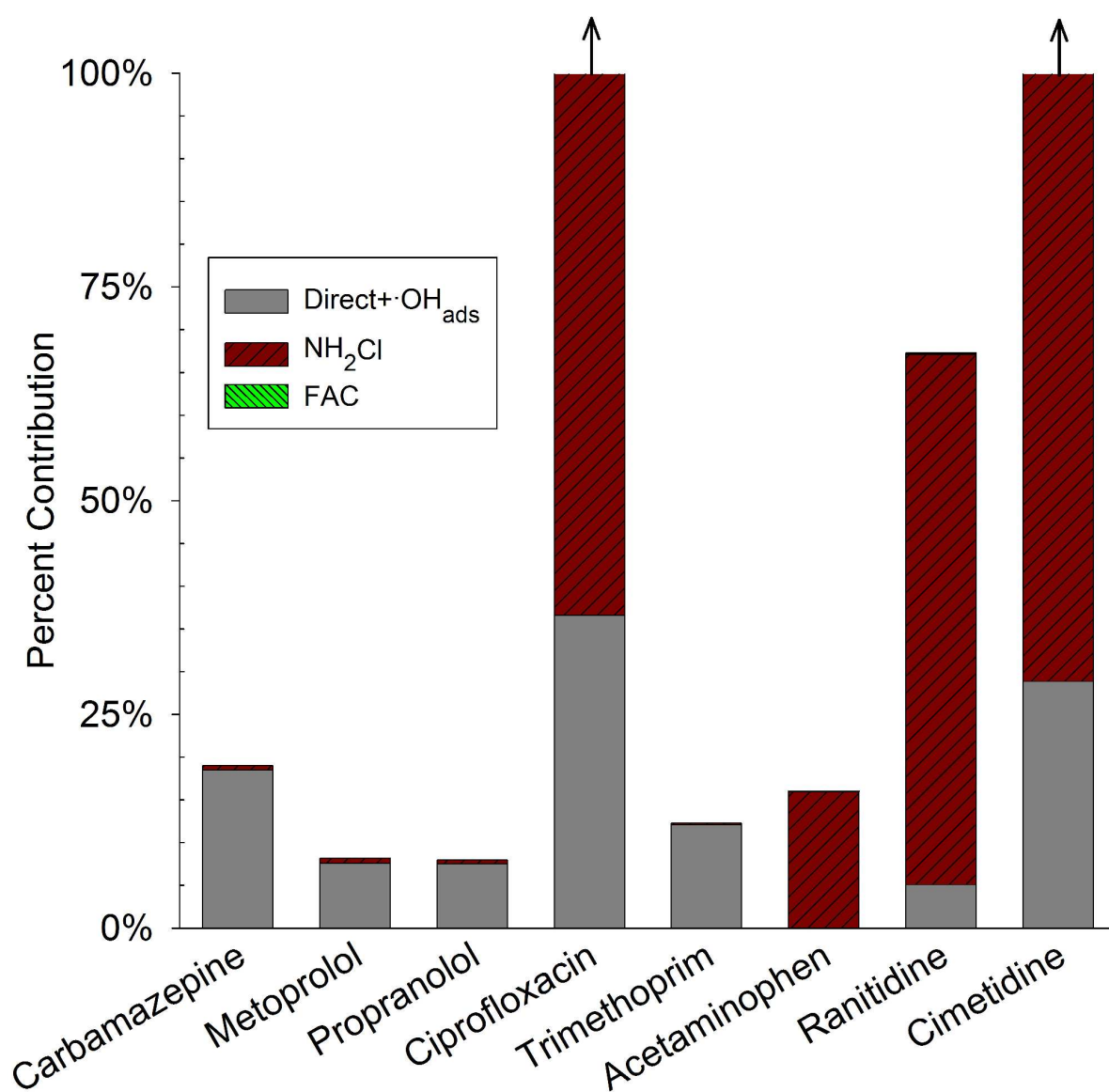


Figure SI 6. Calculated contributions of transformation mechanisms to test compound removal in latrine wastewater electrolyzed at 3.5 V (0.7 A L⁻¹). Ciprofloxacin and cimetidine removal rates were over-predicted by reaction with NH₂Cl.

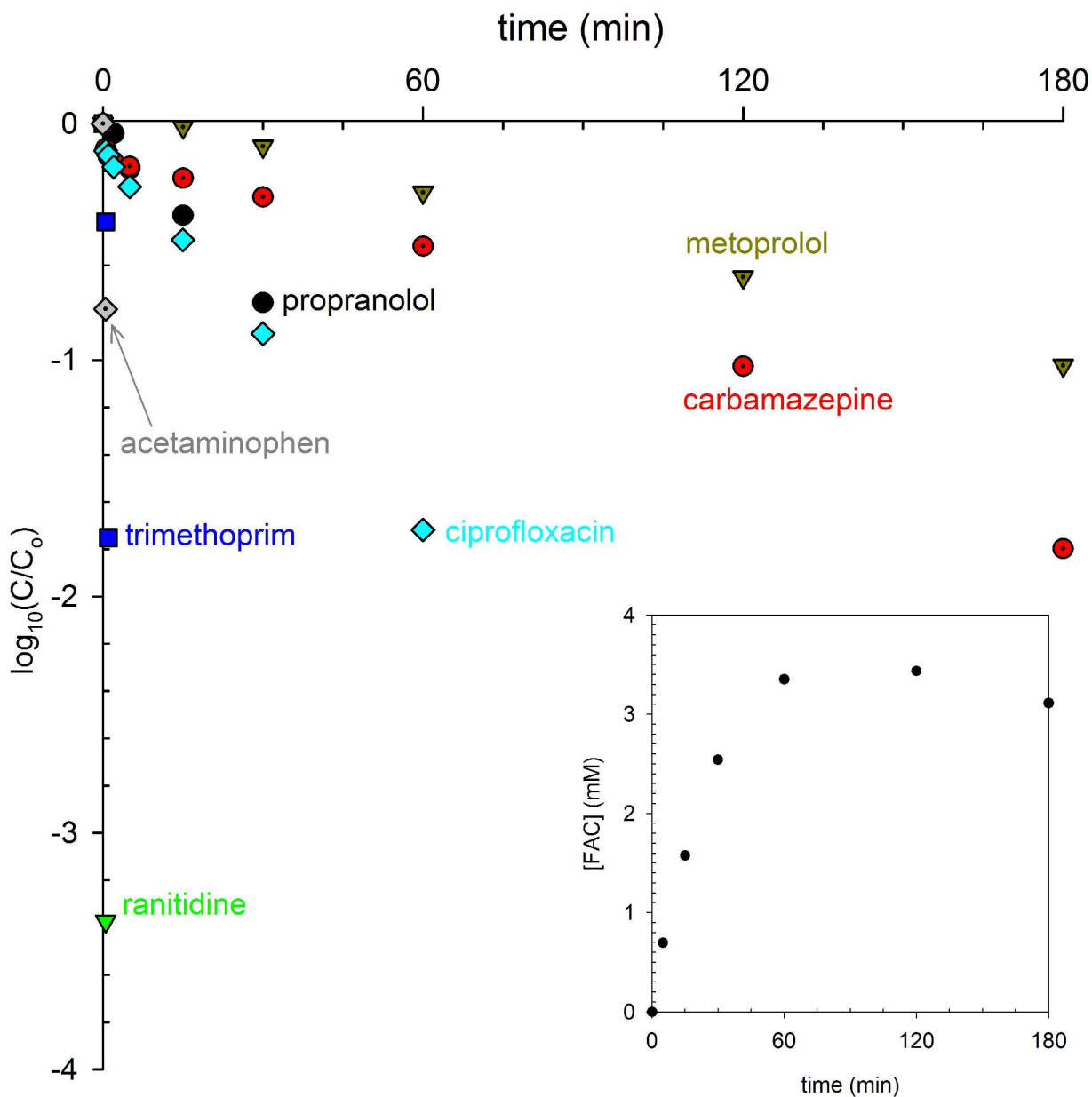
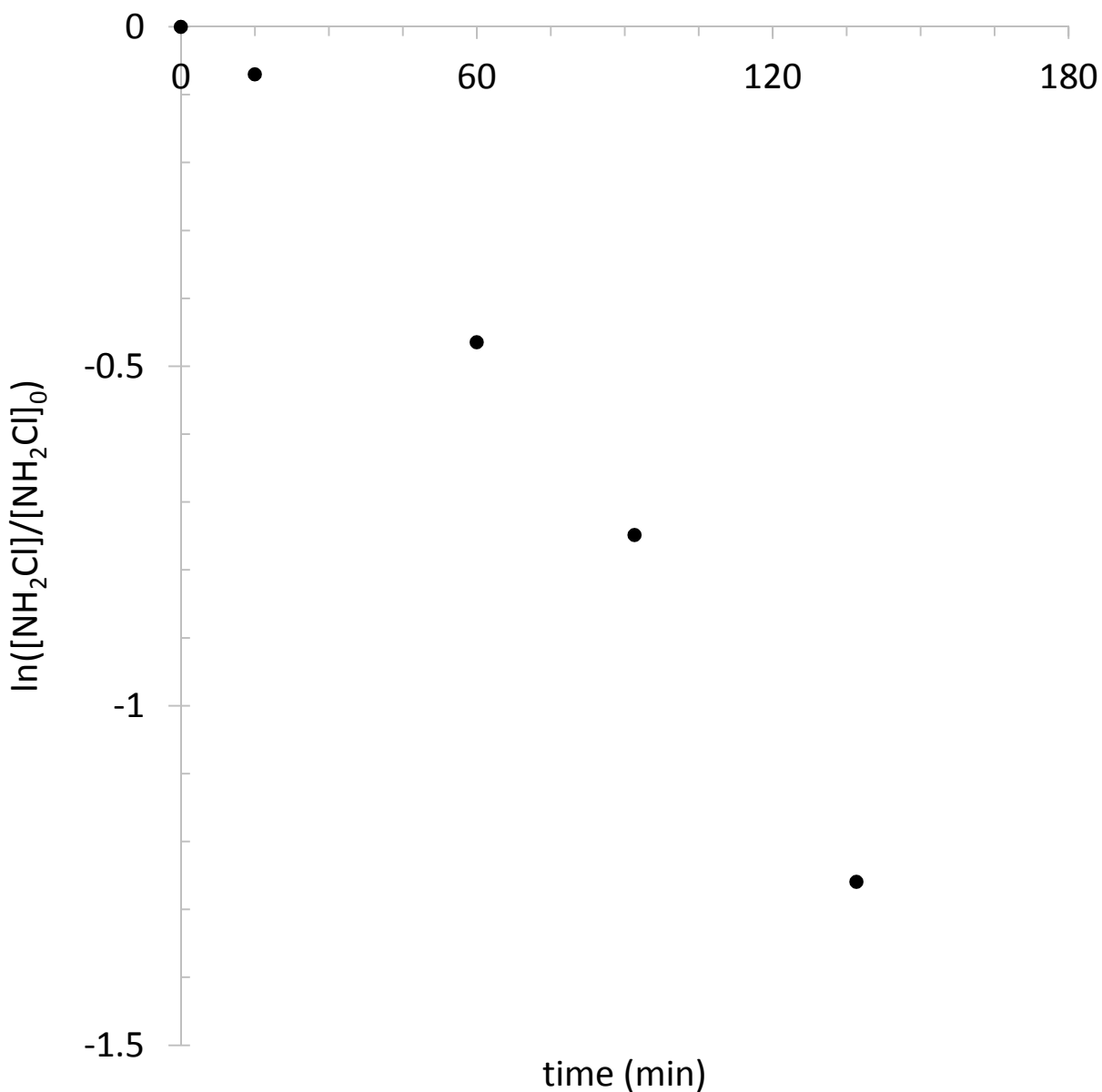


Figure SI 7. Test compound removal during electrolysis (3.5 V; 0.9 A L⁻¹) in 30 mM NaCl solution buffered at pH 8.7 (20 mM borate). Inset: [FAC] ([HOCl]+[OCl⁻]) during electrolysis. Note that FAC concentrations plateaued after ~60 min due to FAC reduction on the cathode.²⁸



192
 193 Figure SI 8. NH_2Cl reduction in buffered solution (pH 8.7) on a stainless steel cathode
 194 ($k=0.02\text{ C}^{-1}$). The experiment was performed in an H cell with anodic and cathodic chambers
 195 separated by a Nafion membrane to prevent formation of chloramines at the anode. The cathodic
 196 potential was controlled at a similar potential to that observed during wastewater electrolysis
 197 (i.e., about -1.5 V vs. an Ag/AgCl/Sat. NaCl reference electrode; 10 mA). $[NH_2Cl]_0=1\text{ mM}$.
 198 Migration of NH_2Cl through the Nafion membrane was insignificant over the 2 hr experiment.

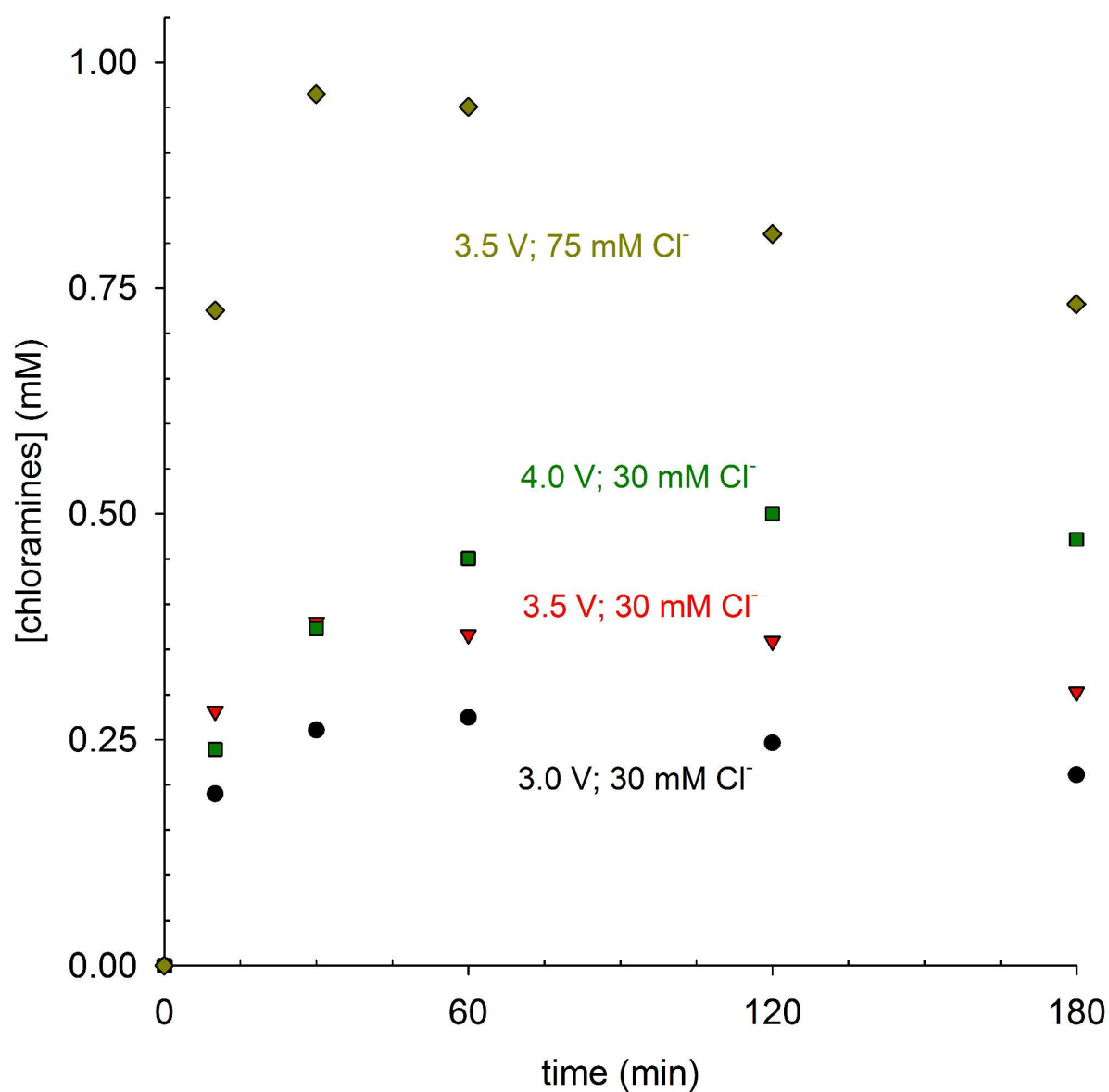


Figure SI 9. Chloramine concentration during electrolysis of latrine wastewater at various applied potentials (3.0-4.0 V) and chloride concentrations (30 mM; 75 mM). Average current densities: 3.0 V, 0.5 A L⁻¹; 3.5 V, 0.7 A L⁻¹; 4.0 V, 1.3 A L⁻¹; 3.5 V 75 mM Cl⁻, 1.1 A L⁻¹.

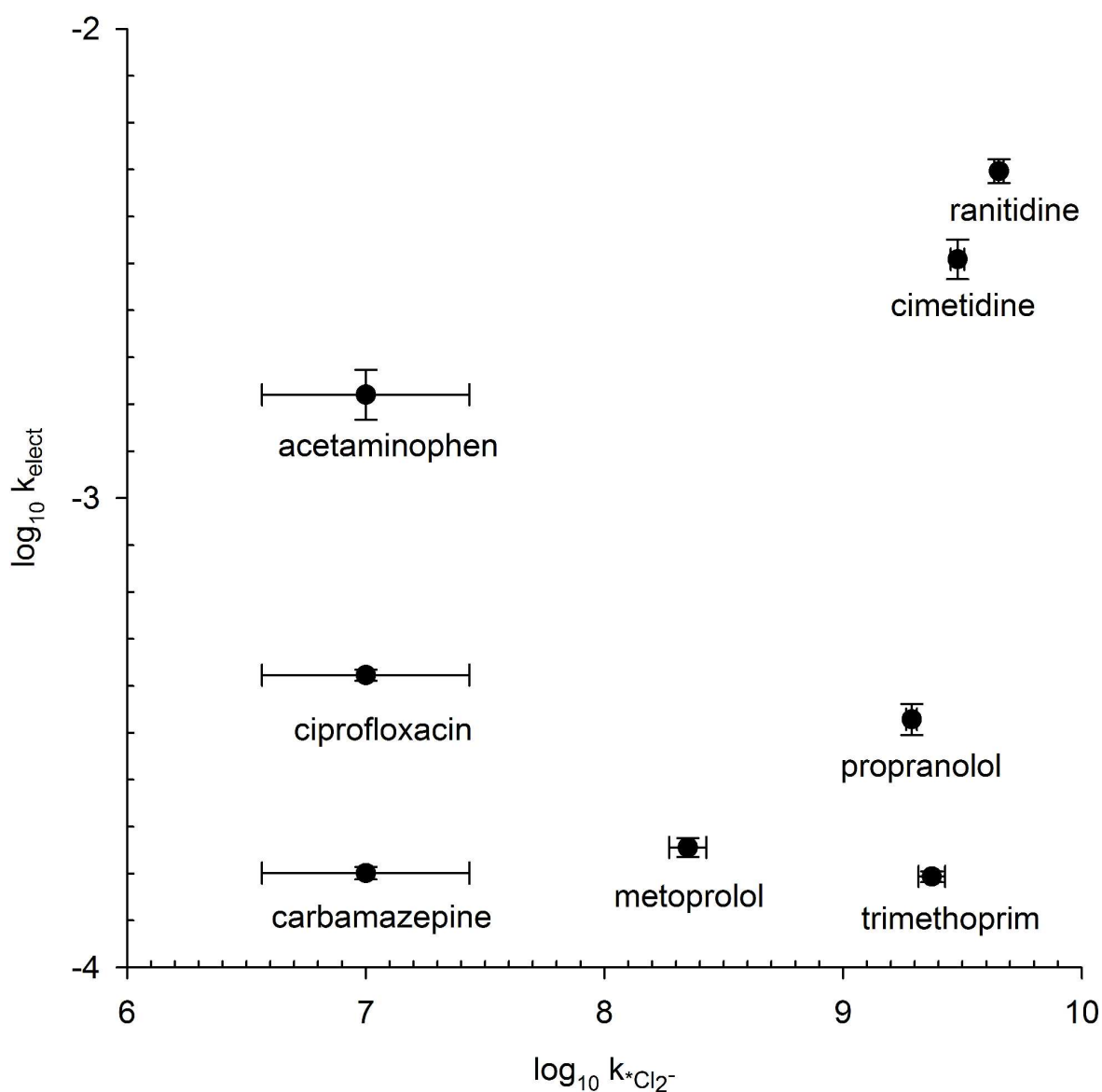


Figure SI 10. Correlation between test compound pseudo-first order electrolysis rates (k_{elect} ; s^{-1}) in latrine wastewater at an applied potential of 3.5 V and measured second-order reaction rates with $\cdot\text{Cl}_2^-$ ($k_{\text{Cl}_2^-}$; $\text{M}^{-1} \text{s}^{-1}$). $\cdot\text{Cl}_2^-$ reaction rates with compounds that were too slow to measure by flash photolysis (carbamazepine, ciprofloxacin, and acetaminophen) were assumed to be between $10^{6.5}$ and $10^{7.5} \text{ M}^{-1} \text{s}^{-1}$. Error bars represent \pm one standard deviation.

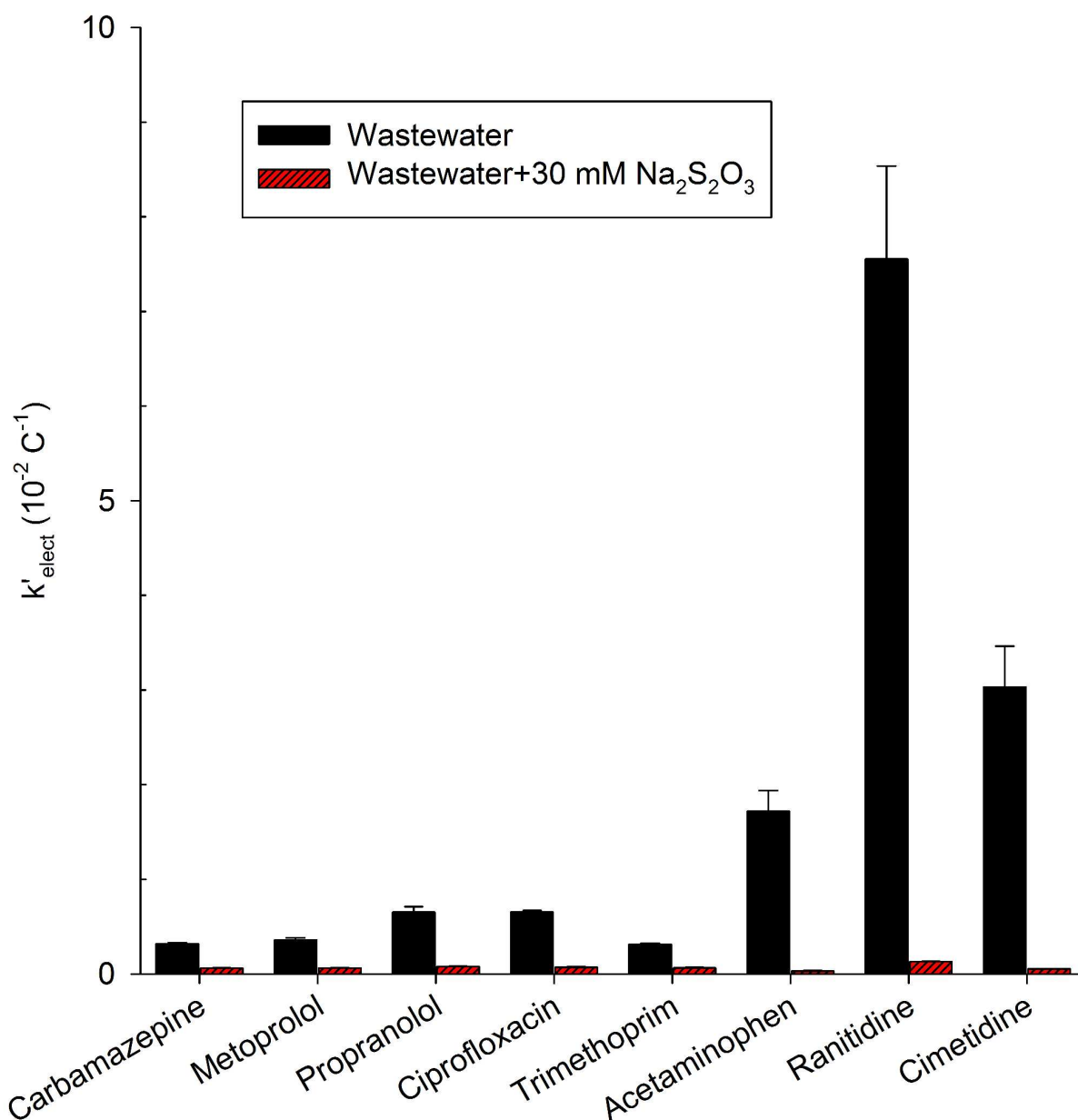
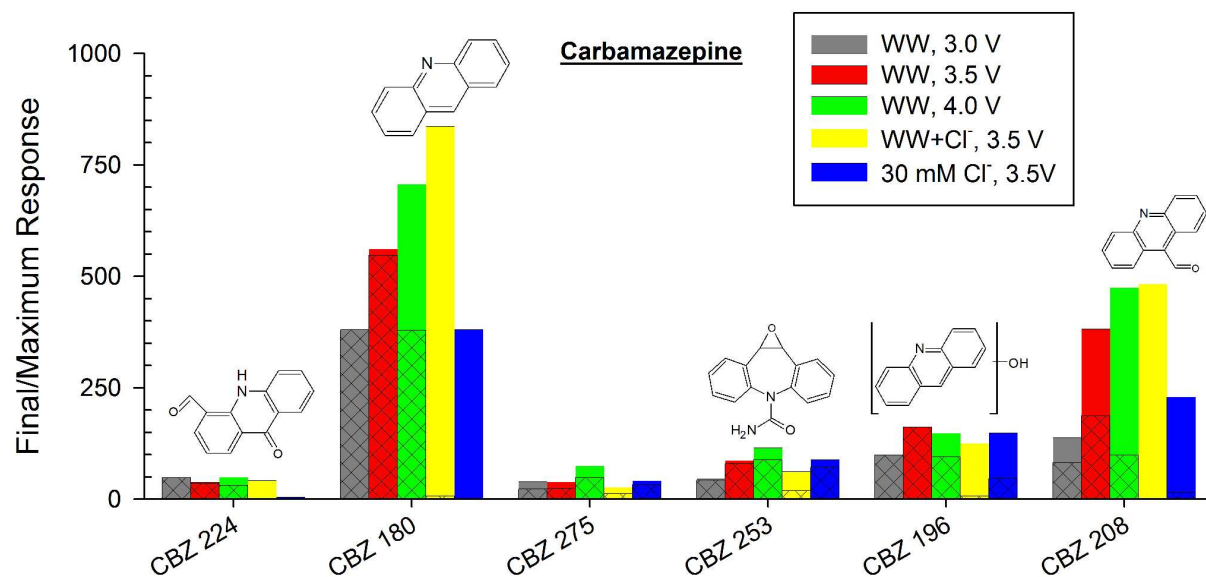


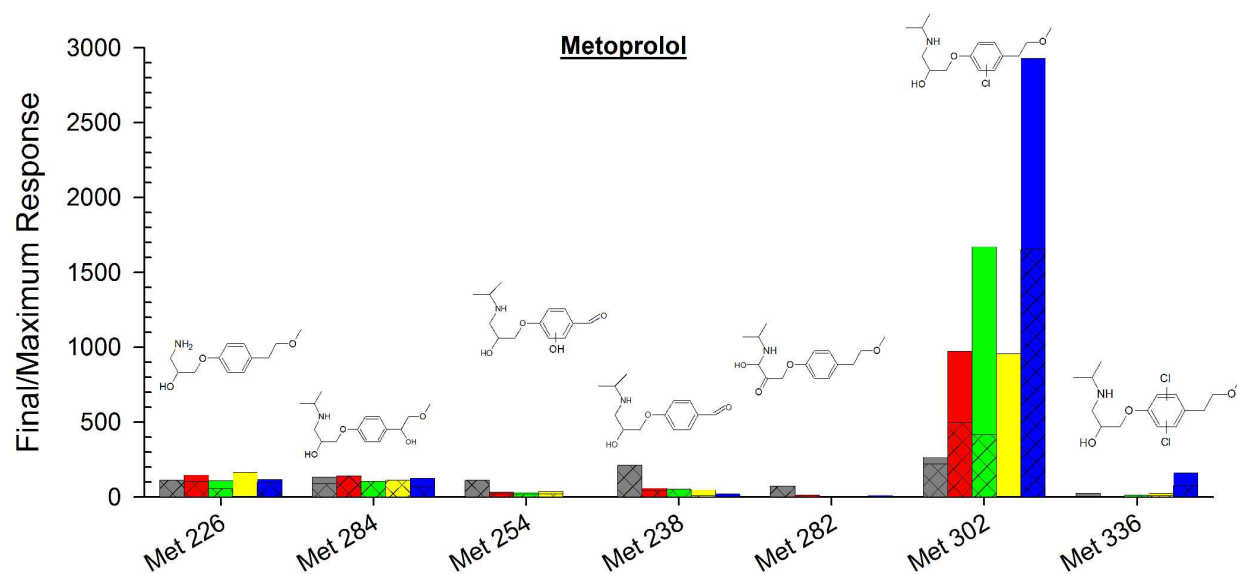
Figure SI 11. Pseudo-first order test compound electrolysis rates normalized for charge (k') in latrine wastewater with or without added $\text{Na}_2\text{S}_2\text{O}_3$. Solutions were electrolyzed at an applied potential of 3.5 V. Average current densities: Wastewater, 0.7 A L^{-1} ; Wastewater+30 mM $\text{Na}_2\text{S}_2\text{O}_3$, 0.9 A L^{-1} . Average chloramine concentration: Wastewater, 0.3 mM; Wastewater+30 mM $\text{Na}_2\text{S}_2\text{O}_3$, 0 mM. Error bars represent \pm one standard deviation.

217 **a**



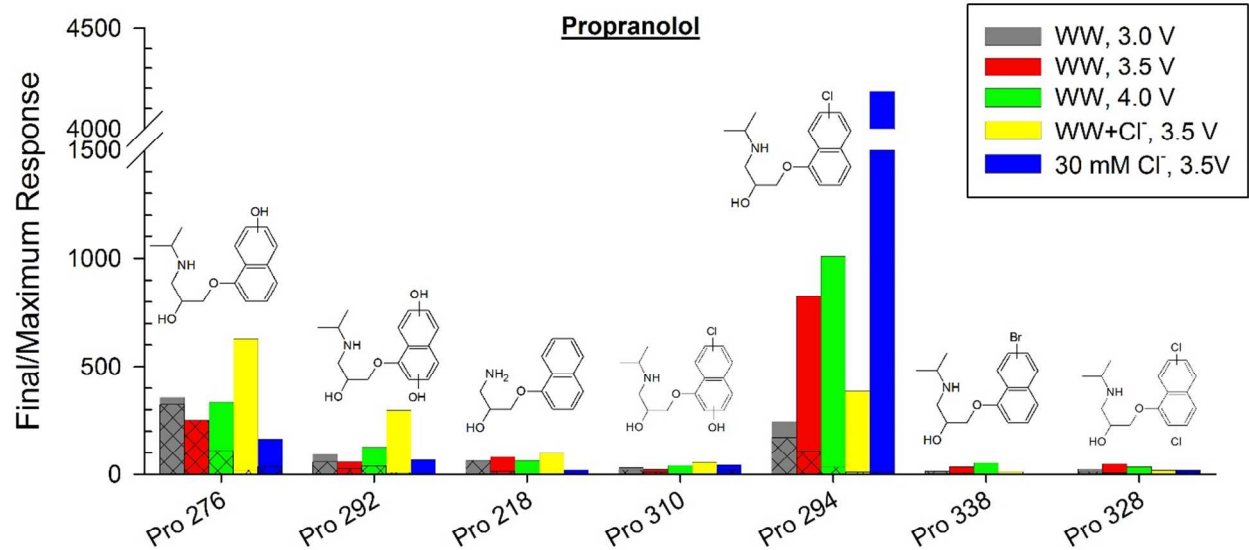
218

219 **b**



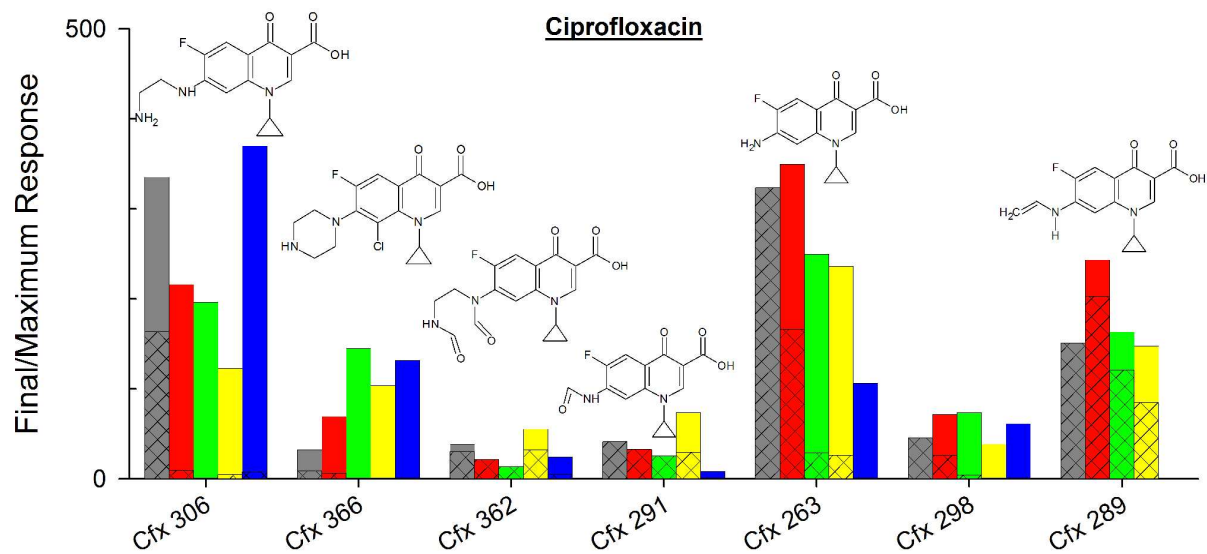
220

221 c



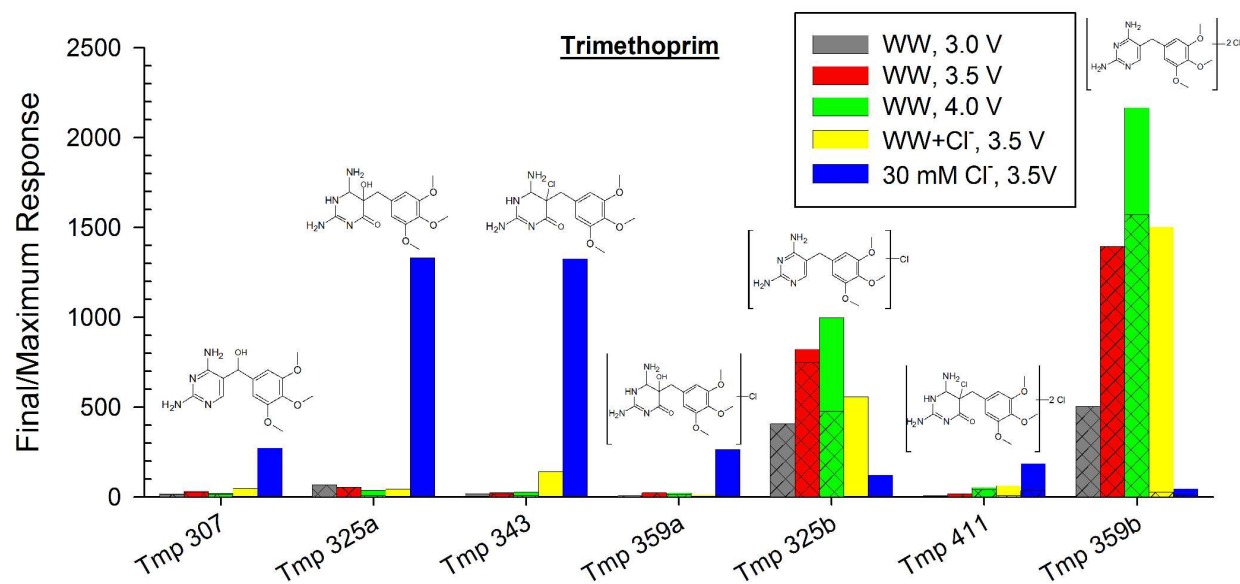
222

223 d



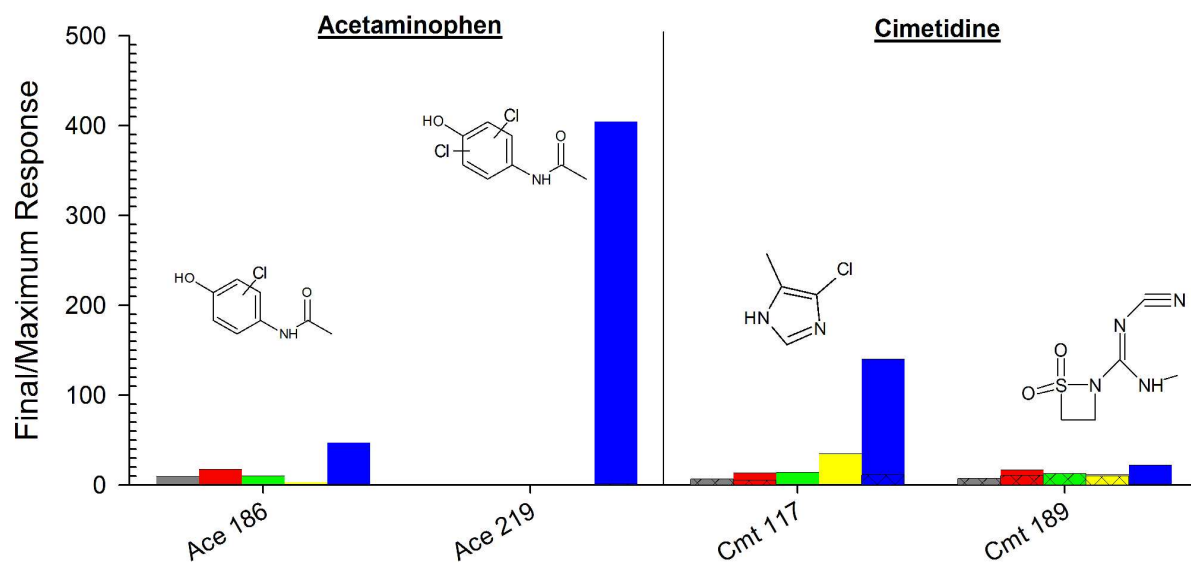
224

225 e



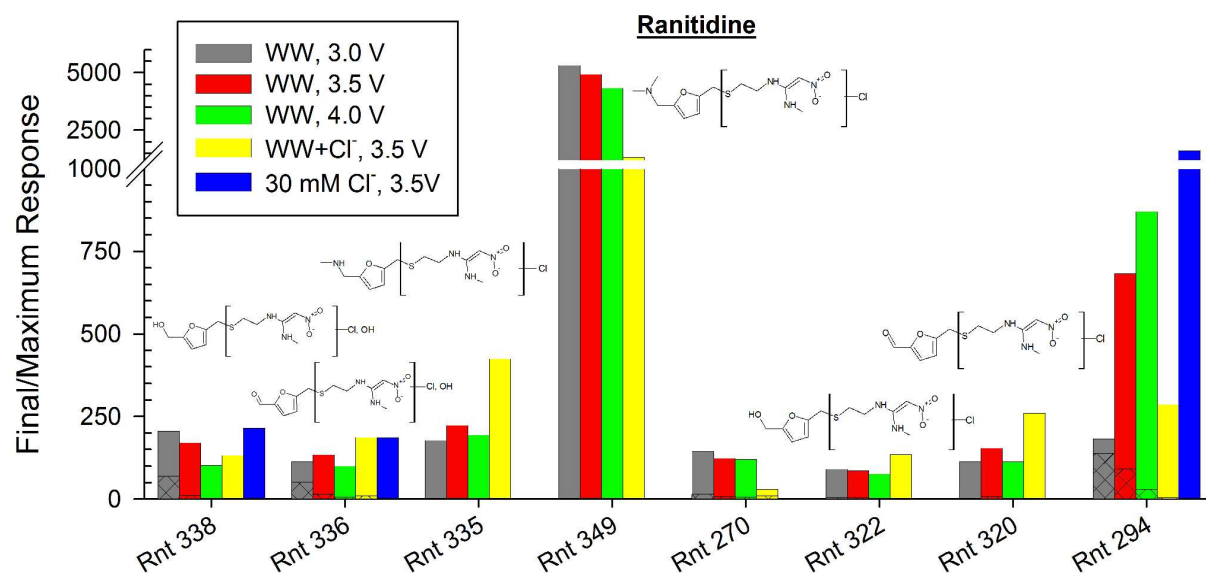
226

227 f



228

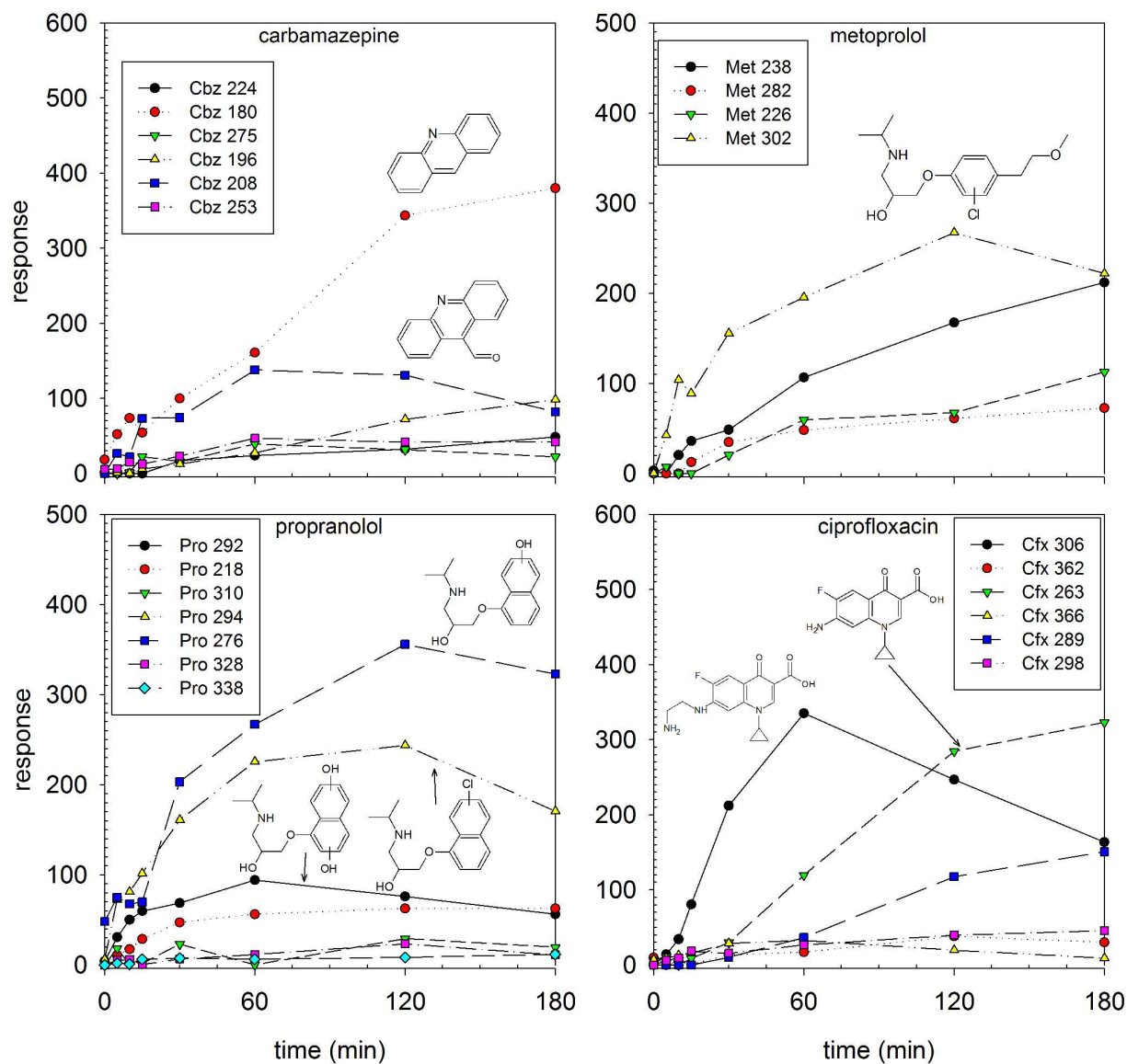
229 g



230
 231 Figure SI 12. Maximum responses of predominant transformation product formed during
 232 electrolysis (entire bar) and final amount remaining at end of electrolysis cycle (cross-hatched
 233 portion). Electrolysis cycle was 180 min. Transformation product IDs represent the nominal
 234 mass to charge ratio for each compound. Structures are proposed based on accurate mass to
 235 charge ratios, fragments, and transformation products reported in the literature. Further
 236 transformation product information is given in Table SI 2.
 237

238 a

3.0 V, 30 mM Cl⁻



239

240

241

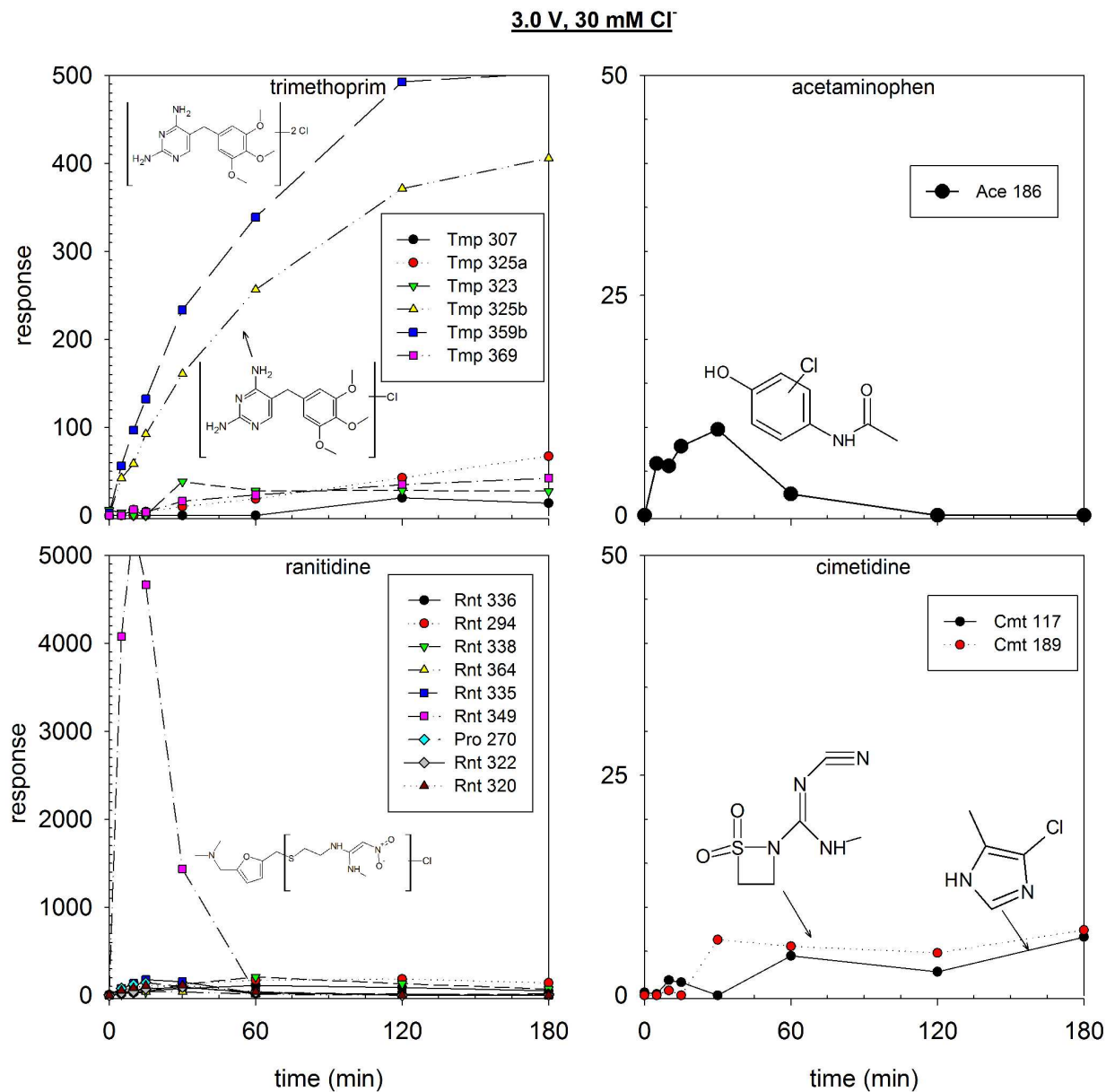
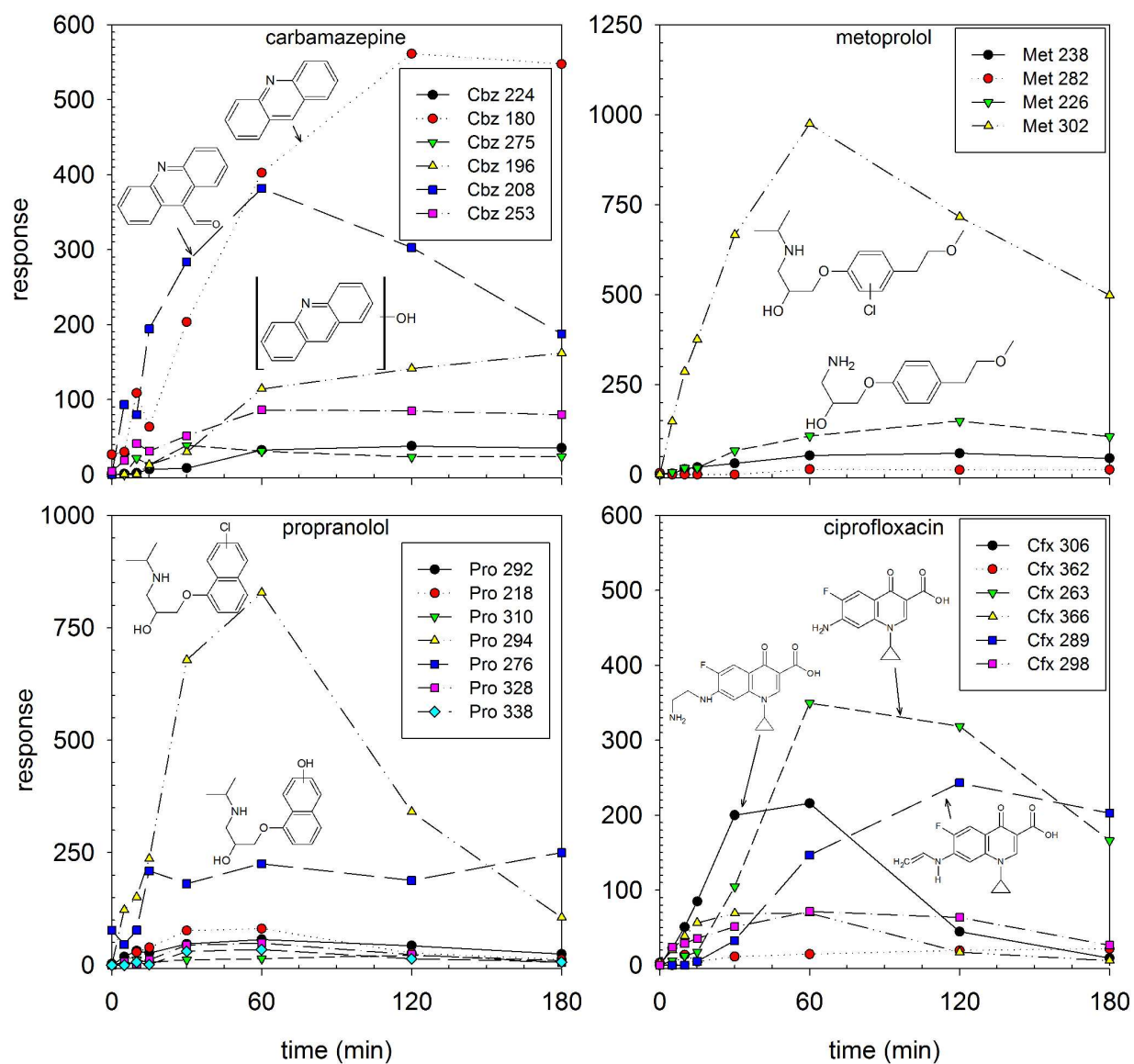


Figure SI 13a. Test compound transformation product formation during latrine wastewater electrolysis at 3.0 V. Transformation product IDs represent the nominal mass to charge ratio for each compound. Further transformation product information is given in Table SI 2.

3.5 V, 30 mM Cl⁻

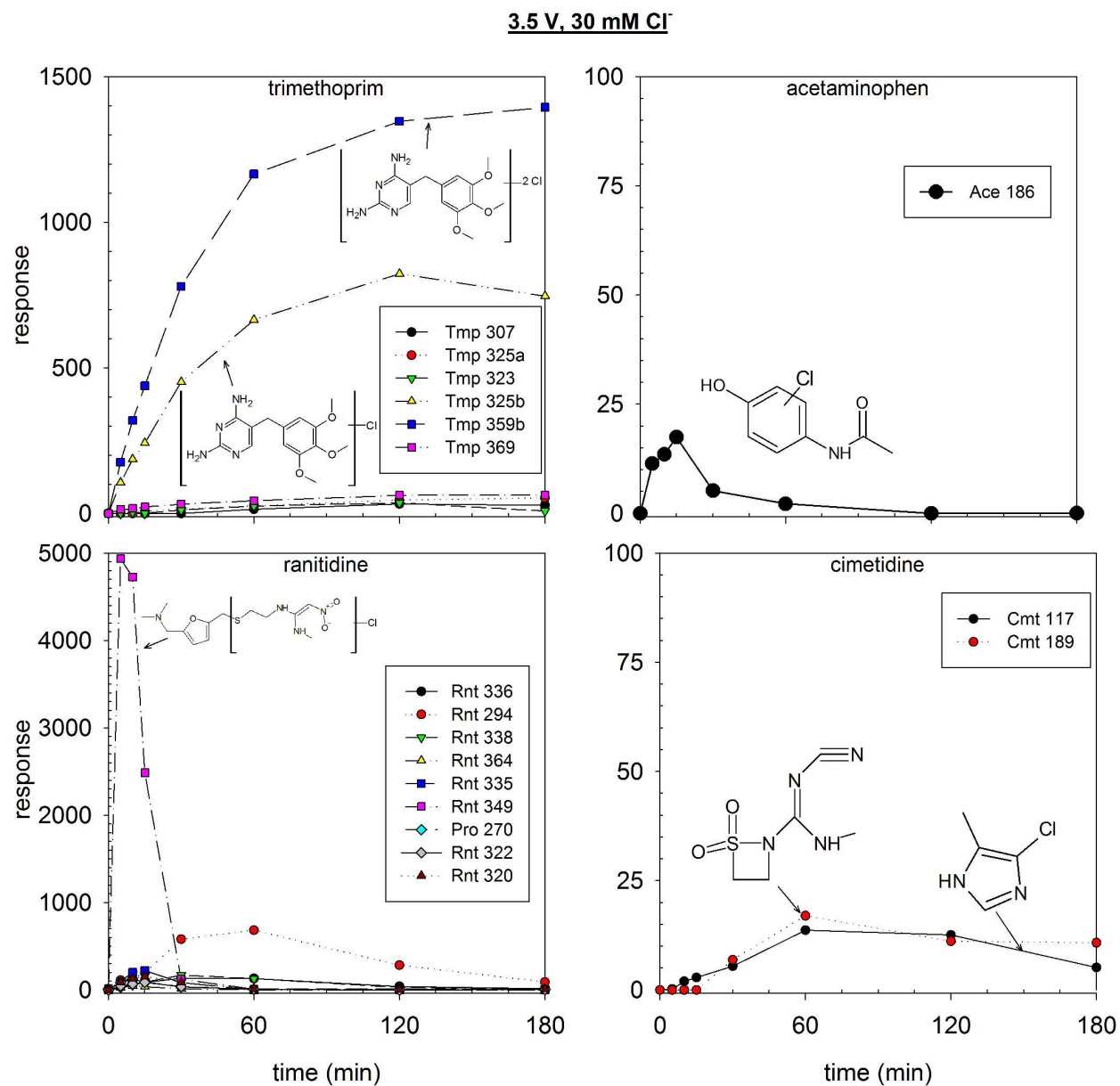
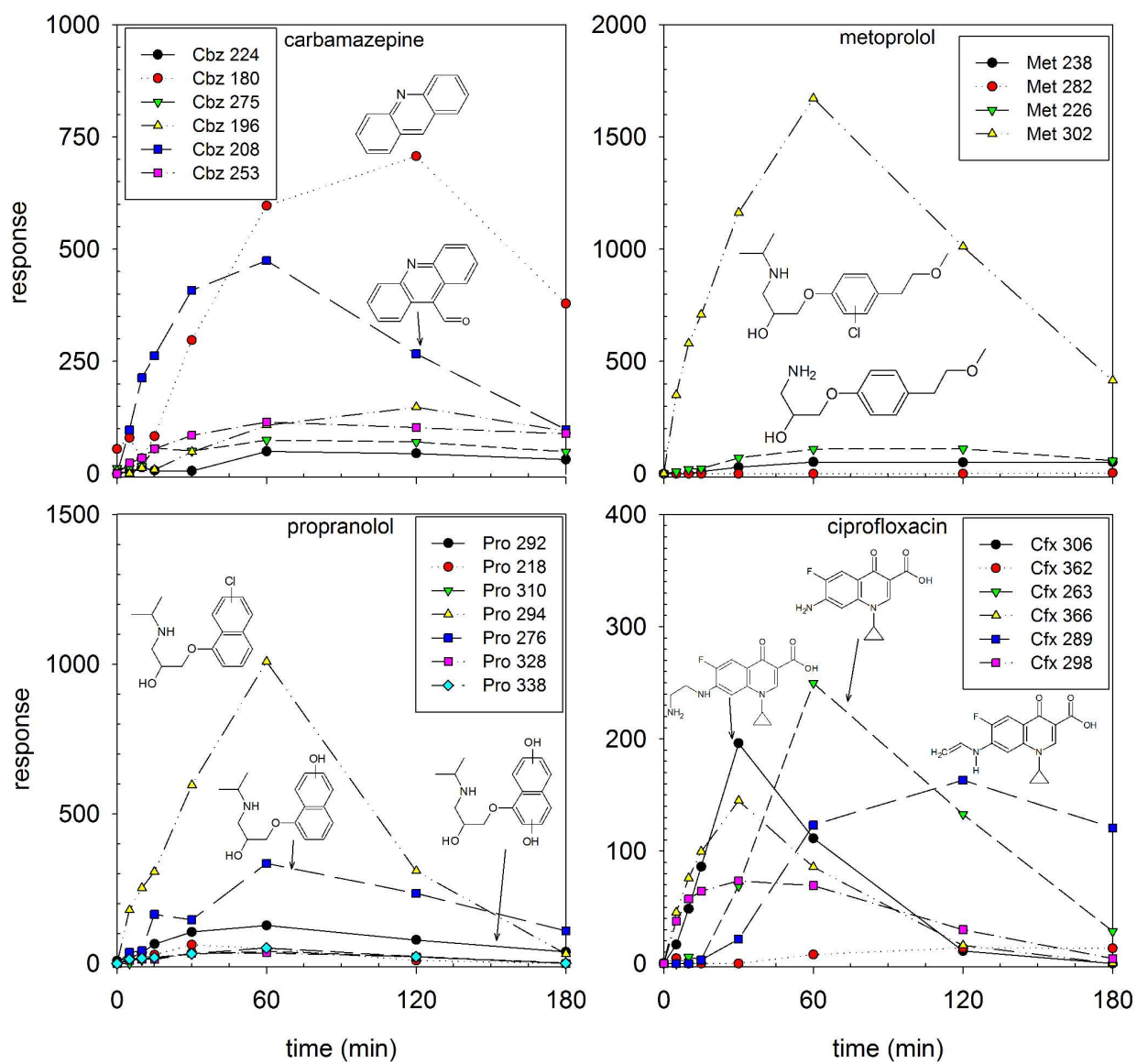


Figure SI 13b. Test compound transformation product formation during latrine wastewater electrolysis at 3.5 V. Transformation product IDs represent the nominal mass to charge ratio for each compound. Further transformation product information is given in Table SI 2.

4.0 V, 30 mM Cl⁻

4.0 V, 30 mM Cl⁻

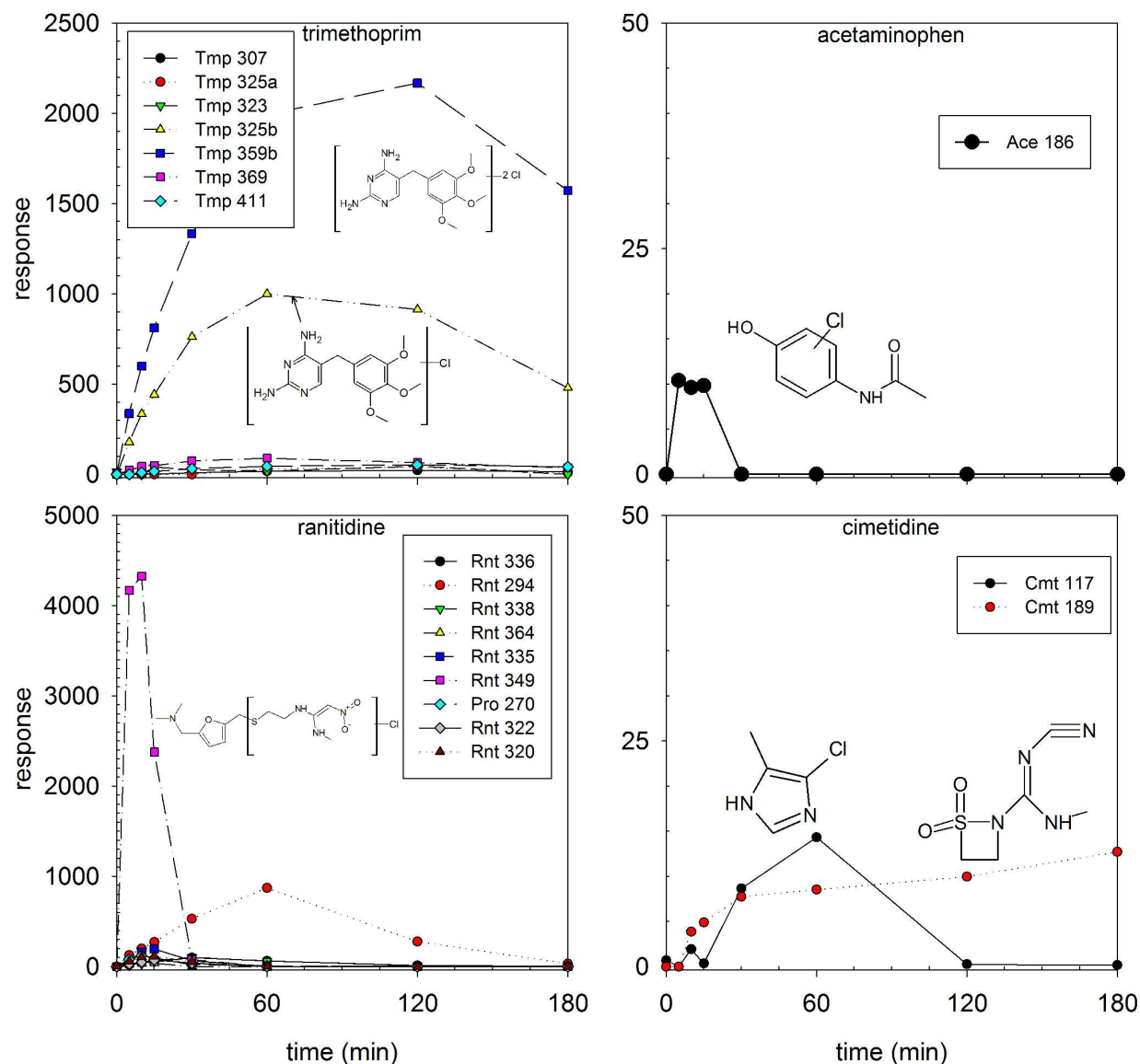
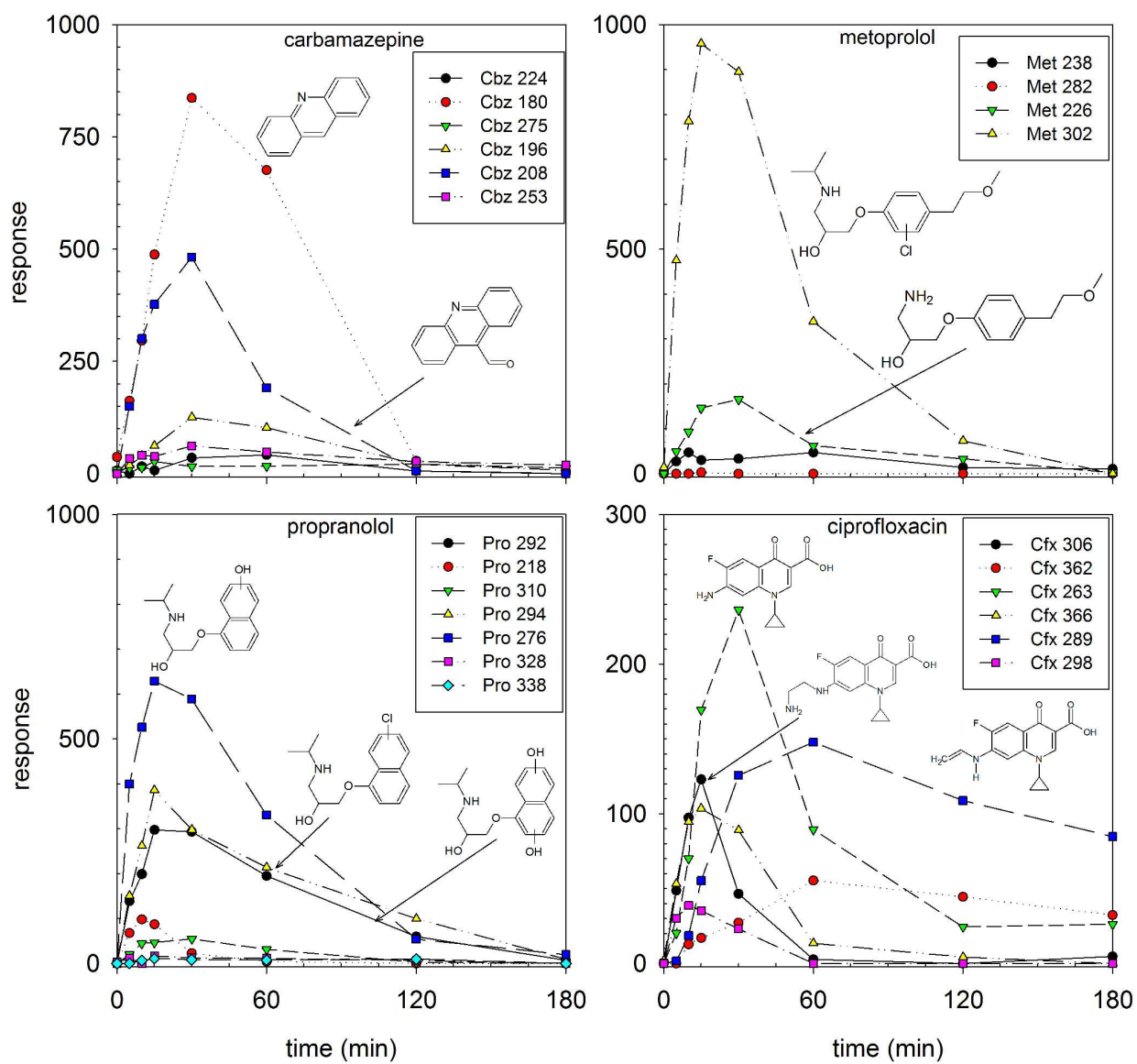


Figure SI 13c. Test compound transformation product formation during latrine wastewater electrolysis at 4.0 V. Transformation product IDs represent the nominal mass to charge ratio for each compound. Further transformation product information is given in Table SI 2.

3.5 V, 75 mM Cl⁻

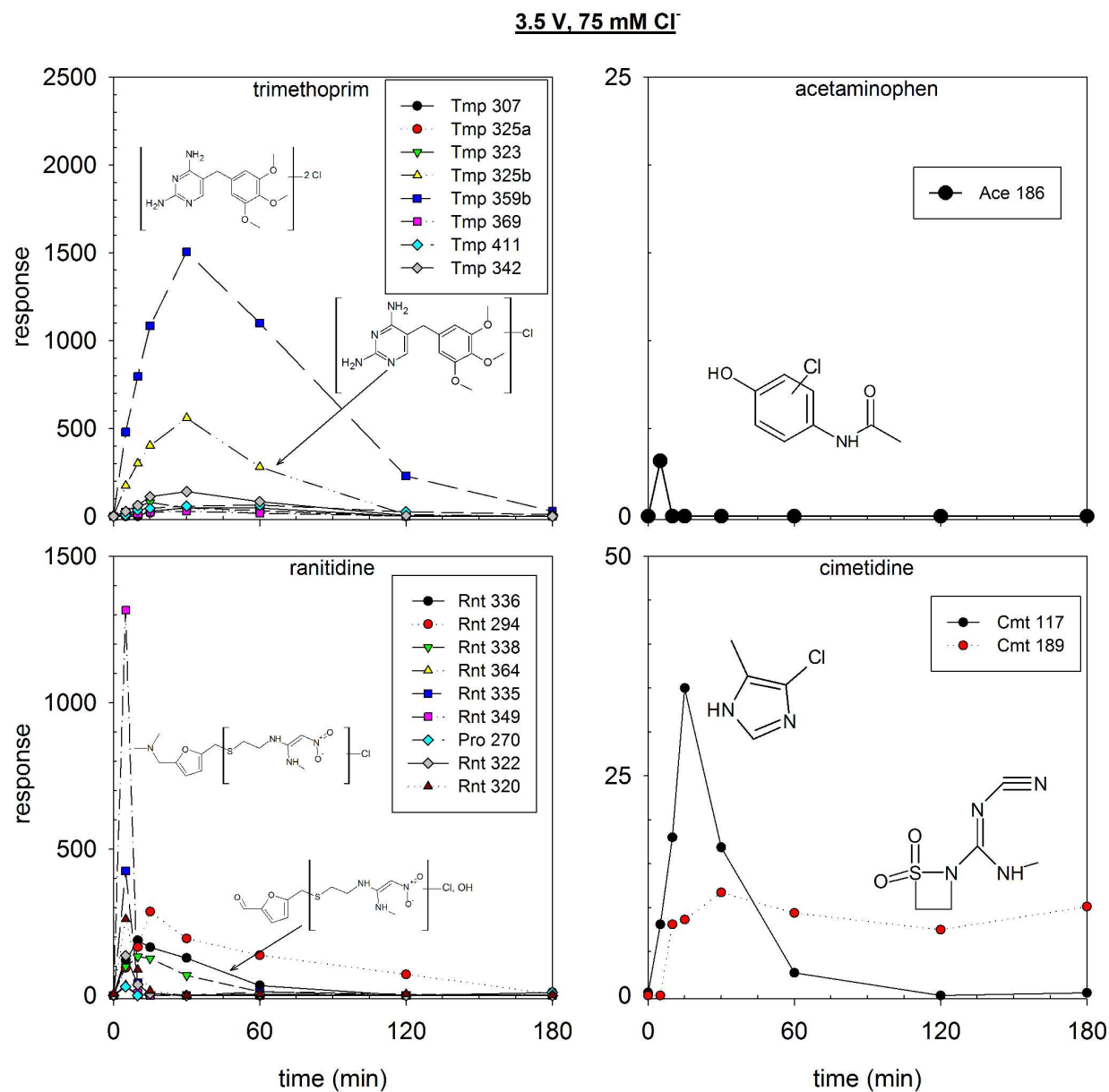
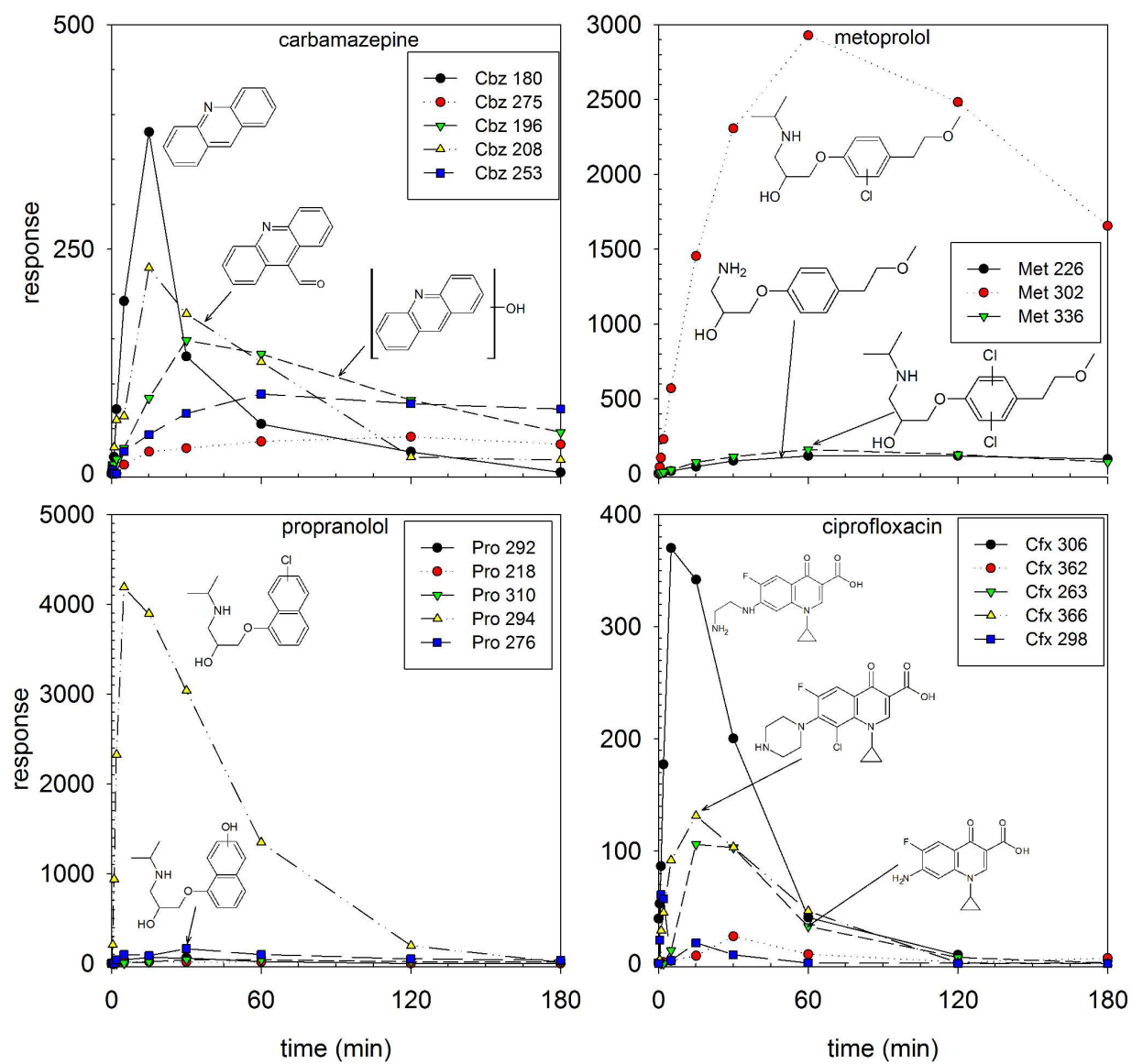


Figure SI 13d. Test compound transformation product formation during latrine wastewater electrolysis at 3.5 V with added NaCl. Transformation product IDs represent the nominal mass to charge ratio for each compound. Further transformation product information is given in Table SI 2.

Buffered water, 3.5 V, 30 mM Cl⁻

Buffered water, 3.5 V, 30 mM Cl⁻

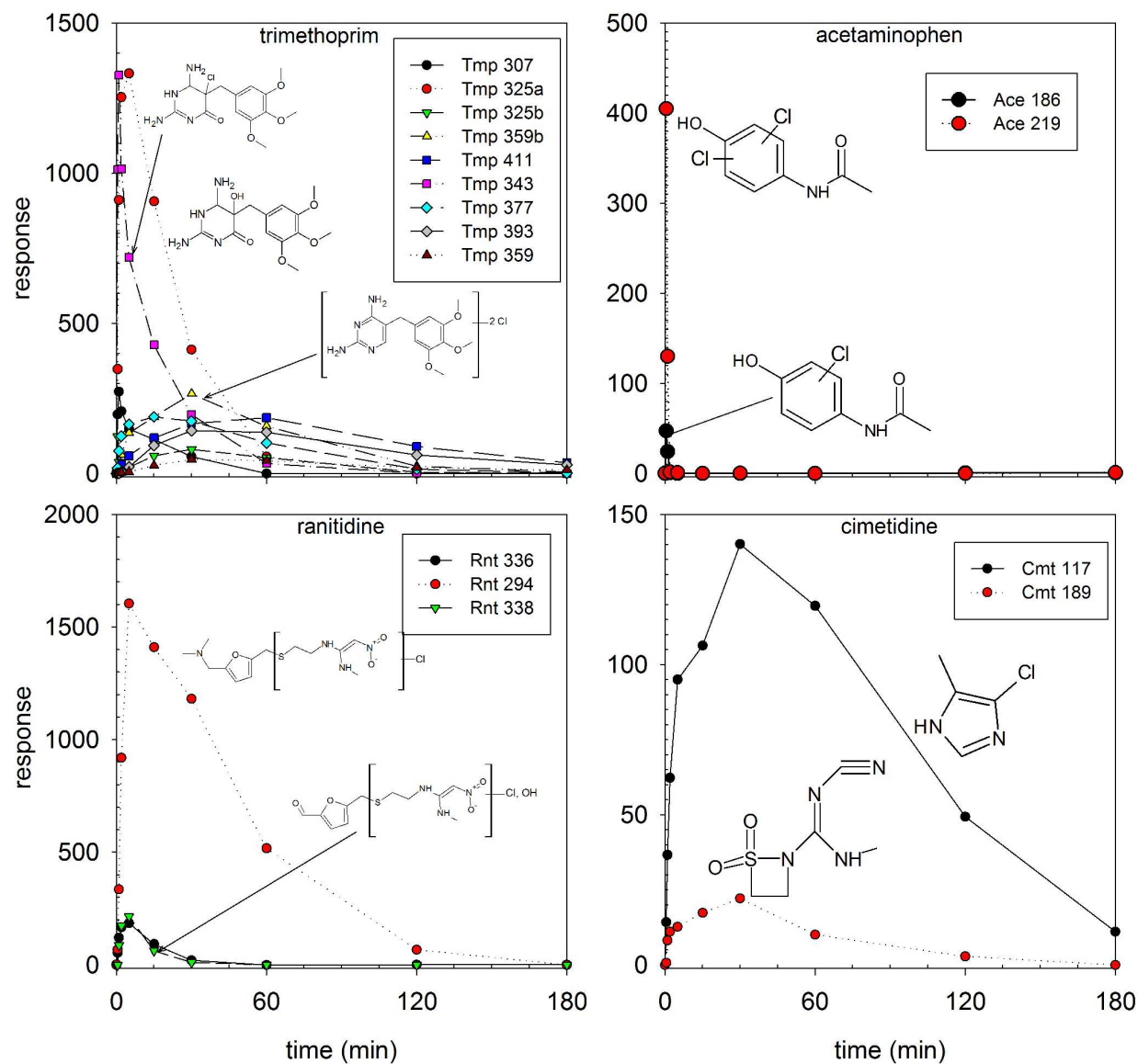


Figure SI 13e. Test compound transformation product formation during buffered Cl⁻ solution electrolysis. Transformation product IDs represent the nominal mass to charge ratio for each compound. Further transformation product information is given in Table SI 2.

271 **References**

- 272 (1) Alegre, M. L.; Geronés, M.; Rosso, J. A.; Bertolotti, S. G.; Braun, A. M.; Mártire, D. O.;
273 Gonzalez, M. C. Kinetic Study of the Reactions of Chlorine Atoms and Cl₂•- Radical
274 Anions in Aqueous Solutions. 1. Reaction with Benzene. *J. Phys. Chem. A* **2000**, *104* (14),
275 3117–3125.
- 276 (2) Kesselman, J. M.; Weres, O.; Lewis, N. S.; Hoffmann, M. R. Electrochemical Production
277 of Hydroxyl Radical at Polycrystalline Nb-Doped TiO₂ Electrodes and Estimation of the
278 Partitioning between Hydroxyl Radical and Direct Hole Oxidation Pathways. *J. Phys.*
279 *Chem. B* **1997**, *101* (14), 2637–2643.
- 280 (3) Zhi, J.-F.; Wang, H.-B.; Nakashima, T.; Rao, T. N.; Fujishima, A. Electrochemical
281 Incineration of Organic Pollutants on Boron-Doped Diamond Electrode. Evidence for
282 Direct Electrochemical Oxidation Pathway. *J. Phys. Chem. B* **2003**, *107* (48), 13389–
283 13395.
- 284 (4) American Public Health Association. *Standard Methods for the Examination of Water and*
285 *Wastewater*, 19th ed.; American Public Health Association,, A. W. W. A., Water
286 Environment Federation: Washington, DC, 1995.
- 287 (5) Wang, W.-L.; Wu, Q.-Y.; Huang, N.; Wang, T.; Hu, H.-Y. Synergistic effect between UV
288 and chlorine (UV/chlorine) on the degradation of carbamazepine: Influence factors and
289 radical species. *Water Res.* **2016**, *98*, 190–198.
- 290 (6) Hu, L.; Martin, H. M.; Arce-Bulted, O.; Sugihara, M. N.; Keating, K. A.; Strathmann, T. J.
291 Oxidation of Carbamazepine by Mn(VII) and Fe(VI): Reaction Kinetics and Mechanism.
292 *Environ. Sci. Technol.* **2009**, *43* (2), 509–515.
- 293 (7) Vogna, D.; Marotta, R.; Andreozzi, R.; Napolitano, A.; d’Ischia, M. Kinetic and chemical
294 assessment of the UV/H₂O₂ treatment of antiepileptic drug carbamazepine. *Chemosphere*
295 **2004**, *54* (4), 497–505.
- 296 (8) Geus, J. *Biotreatment of drinking water resources polluted by pesticides, pharmaceuticals*
297 *and other micropollutants*; BIOTREAT; European Commission, 2014.
- 298 (9) Soufan, M.; Deborde, M.; Delmont, A.; Legube, B. Aqueous chlorination of
299 carbamazepine: Kinetic study and transformation product identification. *Water Res.* **2013**,
300 *47* (14), 5076–5087.
- 301 (10) Radjenovic, J.; Escher, B. I.; Rabaey, K. Electrochemical degradation of the β-blocker
302 metoprolol by Ti/Ru_{0.7}Ir_{0.3}O₂ and Ti/SnO₂-Sb electrodes. *Water Res.* **2011**, *45* (10),
303 3205–3214.
- 304 (11) Veloutsou, S.; Bizani, E.; Fytianos, K. Photo-Fenton decomposition of β-blockers atenolol
305 and metoprolol; study and optimization of system parameters and identification of
306 intermediates. *Chemosphere* **2014**, *107*, 180–186.
- 307 (12) Svan, A.; Hedeland, M.; Arvidsson, T.; Jasper, J. T.; Sedlak, D. L.; Pettersson, C. E.
308 Identification of transformation products from β-blocking agents formed in wetland
309 microcosms using LC-Q-ToF. *J. Mass Spectrom.* **2016**, *51* (3), 207–218.
- 310 (13) Marcourrea, E.; Pereztrujillo, M.; Vicent, T.; Caminal, G. Ability of white-rot fungi to
311 remove selected pharmaceuticals and identification of degradation products of ibuprofen
312 by *Trametes versicolor*. *Chemosphere* **2009**, *74*, 765–772.
- 313 (14) Liu, Q.-T.; Williams, H. E. Kinetics and Degradation Products for Direct Photolysis of β-
314 Blockers in Water. *Env. Sci Technol* **2007**, *41* (3), 803–810.

- (15) Pinkston, K. E.; Sedlak, D. L. Transformation of Aromatic Ether- and Amine-Containing Pharmaceuticals during Chlorine Disinfection. *Environ. Sci. Technol.* **2004**, *38* (14), 4019–4025.
- (16) Jasper, J. T.; Jones, Z. L.; Sharp, J. O.; Sedlak, D. L. Biotransformation of wastewater-derived trace organic contaminants in open-water unit process treatment wetlands. *Environ. Sci. Technol.* **2014**, *48* (9), 5136–5144.
- (17) Dodd, M. C.; Shah, A. D.; von Gunten, U.; Huang, C.-H. Interactions of Fluoroquinolone Antibacterial Agents with Aqueous Chlorine: Reaction Kinetics, Mechanisms, and Transformation Pathways. *Environ. Sci. Technol.* **2005**, *39* (18), 7065–7076.
- (18) Wang, P.; He, Y.-L.; Huang, C.-H. Oxidation of fluoroquinolone antibiotics and structurally related amines by chlorine dioxide: Reaction kinetics, product and pathway evaluation. *Water Res.* **2010**, *44* (20), 5989–5998.
- (19) Anquandah, G. A. K.; Sharma, V. K.; Knight, D. A.; Batchu, S. R.; Gardinali, P. R. Oxidation of Trimethoprim by Ferrate(VI): Kinetics, Products, and Antibacterial Activity. *Environ. Sci. Technol.* **2011**, *45* (24), 10575–10581.
- (20) Eichhorn, P.; Ferguson, P. L.; Pérez, S.; Aga, D. S. Application of Ion Trap-MS with H/D Exchange and QqTOF-MS in the Identification of Microbial Degradates of Trimethoprim in Nitrifying Activated Sludge. *Anal. Chem.* **2005**, *77* (13), 4176–4184.
- (21) Dodd, M. C.; Huang, C.-H. Aqueous chlorination of the antibacterial agent trimethoprim: Reaction kinetics and pathways. *Water Res.* **2007**, *41* (3), 647–655.
- (22) Bedner, M.; MacCrehan, W. A. Transformation of Acetaminophen by Chlorination Produces the Toxicants 1,4-Benzoquinone and N-Acetyl-p-benzoquinone Imine. *Environ. Sci. Technol.* **2006**, *40* (2), 516–522.
- (23) Cao, F.; Zhang, M.; Yuan, S.; Feng, J.; Wang, Q.; Wang, W.; Hu, Z. Transformation of acetaminophen during water chlorination treatment: kinetics and transformation products identification. *Environ. Sci. Pollut. Res.* **2016**, *23* (12), 1–9.
- (24) Buth, J. M.; Arnold, W. A.; McNeill, K. Unexpected Products and Reaction Mechanisms of the Aqueous Chlorination of Cimetidine. *Environ. Sci. Technol.* **2007**, *41* (17), 6228–6233.
- (25) Le Roux, J.; Gallard, H.; Croué, J.-P.; Papot, S.; Deborde, M. NDMA Formation by Chloramination of Ranitidine: Kinetics and Mechanism. *Environ. Sci. Technol.* **2012**, *46* (20), 11095–11103.
- (26) Acero, J. L.; Benitez, F. J.; Real, F. J.; Roldan, G. Kinetics of aqueous chlorination of some pharmaceuticals and their elimination from water matrices. *Water Res.* **2010**, *44* (14), 4158–4170.
- (27) Gallard, H.; von Gunten, U. Chlorination of Phenols: Kinetics and Formation of Chloroform. *Environ. Sci. Technol.* **2002**, *36* (5), 884–890.
- (28) Park, H.; Vecitis, C. D.; Hoffmann, M. R. Electrochemical Water Splitting Coupled with Organic Compound Oxidation: The Role of Active Chlorine Species. *J. Phys. Chem. C* **2009**, *113* (18), 7935–7945.