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Supporting Information for:

**Electrochemical Transformation of Trace Organic
Contaminants in Latrine Wastewater**

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42 pages
13 figures
2 tables

35 **Reaction Rate Constants for FAC and NH₂Cl.** Test compound (100 nM) removal was
36 followed in buffered solutions (pH 8.7; 20 mM borate) with excess FAC (0.03-5.0 mM) or
37 NH₂Cl (0.3-3.0 mM). NH₂Cl solutions were prepared by adding stock solutions of FAC
38 dropwise to excess NH₄Cl (1.2:1 NH₄Cl:HOCl). Samples were quenched (45 mM Na₂S₂O₃)
39 prior to analysis. Solution pH values and total chlorine concentrations did not change
40 significantly over the course of the experiments. Rate constants were calculated based on first-
41 order removal kinetics of the test compounds using measured steady-state total chlorine
42 concentrations. For ciprofloxacin, rate constants were measured at multiple NH₂Cl and FAC
43 concentrations to verify that observed ciprofloxacin transformation rates were not dependent on
44 oxidant concentrations, but rather represented decay of the rapidly formed N-chlorinated
45 ciprofloxacin intermediate as previously reported.¹ First-order ciprofloxacin decay rates were
46 therefore reported.

47 **Reaction Rate Constants for the Chlorine Radical Anion, $\cdot\text{Cl}_2^-$.** Second-order rate
48 constants for the reaction of test compounds with the dichlorine radical anion ($\cdot\text{Cl}_2^-$) were
49 measured by nanosecond transient absorption laser flash photolysis of solutions containing
50 $\text{Na}_2\text{S}_2\text{O}_8$ (25 mM), NaCl (100 mM), and test compounds (0-100 μM). Excitation at 266 nm (8 ns
51 FWHM, 10 Hz repetition rate) produced $\cdot\text{SO}_4^-$ that rapidly reacted with Cl^- to produce $\cdot\text{Cl}_2^-$
52 ($\sim 1 \mu\text{M}$). Solution pH values ranged from 5.5 to 6.0. The decay of $\cdot\text{Cl}_2^-$ was monitored at 340
53 nm over 10 ms, and data was averaged over 10 shots. $\cdot\text{Cl}_2^-$ decay was log-normalized and
54 plotted versus test compound concentrations to determine second-order rate constants. $\cdot\text{Cl}_2^-$
55 decay was log-normalized and fit according to:

$$56 \qquad \qquad \qquad A(340) = fe^{-gt} + h \qquad \qquad \qquad (\text{S1})$$

57 where $A(340)$ is the absorbance at 340 nm, f is the initial absorbance, g is the decay rate, and h is
58 an offset due to a longer lived species formed after $\cdot\text{Cl}_2^-$ decay.² $\cdot\text{Cl}_2^-$ decay rates were plotted
59 versus test compound concentrations to determine second-order rate constants:

$$60 \qquad \qquad \qquad g = k_{\text{cmpd},\cdot\text{Cl}_2^-}[\text{cmpd}] + b \qquad \qquad \qquad (\text{S2})$$

61 where $k_{\text{cmpd},\cdot\text{Cl}_2^-}$ is the second-order rate constant for the reaction of a test compound with $\cdot\text{Cl}_2^-$,
62 $[\text{cmpd}]$ is the test compound concentration, and b is the y-intercept that represents loss of $\cdot\text{Cl}_2^-$
63 due to other reactions (e.g., quenching by H_2O). Under these conditions $\cdot\text{Cl}$ is predicted to
64 contribute less than 5% to measured second-order rate constants greater than $10^7 \text{ M}^{-1} \text{ s}^{-1}$.

65 **Chronoamperometric Experiments.** Chronoamperometric experiments were
66 performed to verify direct electron transfer for compounds that underwent transformation in the
67 absence of Cl⁻ and were soluble enough dissolve rapidly (i.e., ranitidine and cimetidine). About
68 5 mM of a solid test compound was added to a solution of Na₂SO₄ (20 mM) undergoing
69 potentiostatic electrolysis (3.5 V cathode vs. anode), and the current response was observed. A
70 spike in the current upon test compound addition implied that the compound could be oxidized
71 directly at the anode.^{3,4} The possibility that current increased due to a change in electrolyte
72 strength was eliminated based on the current's return to near baseline values after test compound
73 oxidation was complete.

74 **Transformation Product Identification.** Test compound transformation product
75 formulae were calculated based on accurate masses and typically had a mass error of less than 2
76 mDa (Table SI 2). Transformation product halogen content was confirmed by characteristic
77 isotopic patterns using the $m/z+2$, $m/z+4$, etc. peaks. Isotopic peak ratios were typically within
78 10% of theoretical values for the proposed formulae. Transformation product structures were
79 proposed based on the literature when possible (see bold references in Table SI 2), by comparing
80 exact mass values and fragment ions. In other cases, structures were proposed simply based on
81 determined formulae. In these cases, the exact position of certain functional groups (e.g.,
82 hydroxyl and chlorine) could not be determined. When possible, proposed structures were
83 verified with authentic standards, as specified in Table SI 2.

84 **Analytical Methods.** Dissolved organic carbon and dissolved inorganic carbon were
85 measured by persulfate digestion using an Aurora TOC analyzer.⁵ NO₃⁻, Cl⁻, PO₄³⁻, SO₄²⁻, and
86 NH₄⁺ were analyzed by ion chromatography (Dionex ICS 2000; AS19G anions, CS12A
87 cations).⁵

88 Test compounds were separated by an Acquity BEH C18 column (2.1x50 mm; 1.7 μm
89 particles), eluted at 0.5 mL min⁻¹ using acetonitrile and water with 0.1% formic acid and 1%
90 acetonitrile. The following gradient was used: 0 min, 5% acetonitrile; 0.2 min, 5% acetonitrile;
91 3.2 min, 95% acetonitrile; 3.5 min, 95% acetonitrile; 3.6 min, 5% acetonitrile; 5 min, 5%
92 acetonitrile. Column temperature was held at 35 °C. pCBA were detected by UV absorbance
93 (270 nm). Trace organic compounds were detected by positive electrospray ionization (ESI+) in
94 resolution mode with a capillary voltage of 0.2 kV, a cone voltage of 50 V, and a source offset of
95 80 V. Source temperature was 120 °C and desolvation temperature was 400 °C. Cone gas was
96 40 L/h and desolvation gas was 800 L/h. Scan time was 0.3 s in continuum mode. A collision
97 energy of 1.0 eV was used. A second acquisition channel scanned collision energy from 0-30 eV
98 for fragment identification. Accurate mass values were corrected by the software using a leucine
99 lock-mass.

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101 **Calculation of $\cdot\text{Cl}_2^-$ Reaction with Propranolol.** The fraction of $\cdot\text{Cl}_2^-$ reacting with
102 propranolol ($F_{\cdot\text{Cl}_2^-, \text{propranolol}}$) in latrine wastewater (100 mg L^{-1} TOC) was calculated using $\cdot\text{Cl}_2^-$
103 measured second-order reaction rate constants with propranolol ($k_{\cdot\text{Cl}_2^-, \text{propranolol}}$) and TOC
104 ($k_{\cdot\text{Cl}_2^-, \text{TOC}}$) to be:

$$\begin{aligned} F_{\cdot\text{Cl}_2^-, \text{propranolol}} &= \frac{k_{\cdot\text{Cl}_2^-, \text{propranolol}}[\text{propranolol}]}{k_{\cdot\text{Cl}_2^-, \text{propranolol}}[\text{propranolol}] + k_{\cdot\text{Cl}_2^-, \text{TOC}}[\text{TOC}]} \\ &= \frac{1.9 \times 10^9 \text{ M}^{-1} \text{ s}^{-1} * 1 \times 10^{-6} \text{ M}}{1.9 \times 10^9 \text{ M}^{-1} \text{ s}^{-1} * 1 \times 10^{-6} \text{ M} + 1.9 \times 10^3 (\text{mg L}^{-1})^{-1} \text{ s}^{-1} * 100 \text{ mg L}^{-1}} \\ &= 0.0099 \end{aligned}$$

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106 **Test Compound Transformation Products.** Carbamazepine's predominant
107 transformation products were confirmed to be acridine, hydroxyacridine, and acridine-9-
108 carboxaldehyde (Table SI 2; Figures SI 12a and 13), based on authentic standards. No
109 chlorinated transformation products were identified during latrine wastewater or buffered Cl⁻
110 solution electrolysis. Transformation products were similar to those previously observed during
111 oxidation and chlorination of carbamazepine.⁶⁻¹⁰

112 Metoprolol and propranolol were predominantly transformed to chlorinated products that
113 underwent further transformation to form products not detected by LCMS (Figures SI 12b,c and
114 9). Hydroxylated and multiply halogenated transformation products were also identified (Table
115 SI 2). Most metoprolol transformation products were identified previously during reverse
116 osmosis retentate electrolysis¹¹ or other treatment processes.^{12,13} Some propranolol
117 transformation products observed in this study have been reported previously,¹⁴⁻¹⁷ but others
118 have not, including the predominant chlorinated product (m/z 294.1276 amu; m/z+2 amu
119 observed at ~33%).

120 Identified ciprofloxacin transformation products were typically transformed at the
121 piperazine group rather than chlorinated (Figures SI 12d, 13), as previously observed during
122 ciprofloxacin reaction with FAC and ClO₂.^{1,18}

123 Trimethoprim was transformed into a variety of hydroxylated and chlorinated
124 transformation products, some of which have been identified during treatment using ferrate,¹⁹
125 activated sludge,²⁰ and FAC.²¹

126 Only a few significant acetaminophen and cimetidine transformation products were
127 observed (Figures SI 12f and 13), all of which are produced by reaction with FAC.²²⁻²⁴ As with

128 trimethoprim, different acetaminophen transformation products were observed in the absence of
129 NH_4^+ , including formation of dichlorinated acetaminophen.

130 Ranitidine transformation products were chlorinated, demethylated, and oxidized, with
131 the latter two transformations likely occurring at the furan group.²⁵ Although a few of the
132 identified products have previously been observed during chloramination of ranitidine,²⁵ further
133 work is needed to confirm transformation product structures (Table SI 2). Many transformation
134 products observed during latrine wastewater electrolysis were not observed during electrolysis of
135 buffered Cl^- solutions (Figures SI 12g and 13; see main text).

136 Table SI 1. Comparison of Measured Test Compound Reaction rates with HOCl and NH₂Cl to Literature Values.

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compound	property					
	$k_{\text{FAC}} (\text{M}^{-1}\text{s}^{-1})^{a,b}$		$k_{\text{NH}_2\text{Cl}} (\text{M}^{-1}\text{s}^{-1})^a$		$k_{\text{Cl}_2} (\text{M}^{-1}\text{s}^{-1})$	
	measured	literature	measured	literature	measured	literature
carbamazepine	$8.9(\pm 1.0) \times 10^{-2}$	$2.7 \times 10^{-2} (10)$	$< 3 \times 10^{-3}$	n.a. ^c	$\sim 1 \times 10^5$	$6.7 \times 10^4 (16)$
metoprolol	$1.9(\pm 1.0) \times 10^{-2}$	$2 \times 10^{-2} (16)$	$< 4 \times 10^{-3}$	$< 0.1 (16)$	$\sim 4 \times 10^3$	$0.6-1.4 \times 10^3 (16)$
propranolol	$0.27(\pm 0.02)$	$0.3 (16)$	$< 6 \times 10^{-3}$	$< 0.1 (16)$	$\sim 2 \times 10^5$	$3.3 \times 10^5 (16)$
ciprofloxacin	$5.6(\pm 0.4) \times 10^{-4} \text{ s}^{-1} e$	$7 \times 10^{-4} \text{ s}^{-1} (1) e$	$9 \times 10^{-4} \text{ s}^{-1} e$	$7 \times 10^{-4} \text{ s}^{-1} (1) e$	$\sim 6 \times 10^3$	$4.3 \times 10^3 (1) f$
trimethoprim	$6.7(\pm 0.2)$	$8 (21)$	$< 1 \times 10^{-3}$	negligible ⁽²¹⁾	$\sim 3 \times 10^6$	$2 \times 10^5 - 2 \times 10^6 (21) g$
acetaminophen	$7.0(\pm 0.4) \times 10^2$	$0.7-1.4 \times 10^2 (16,23,27)$	$0.9(\pm 0.1)$	$1.1 \times 10^{-2} (16)$	$\sim 3 \times 10^4$	n.a. ^c
ranitidine	$> 8 \times 10^3$	n.a. ^c	$11(\pm 1)$	$12 (25)$	$\sim 8 \times 10^4$	n.a. ^c
cimetidine	$> 6.5 \times 10^3$	$> 1 \times 10^3 (24) g$	$47(\pm 1)$	$48 (24)$	$\sim 4 \times 10^6$	n.a. ^c

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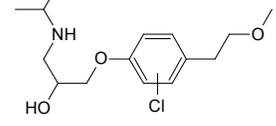
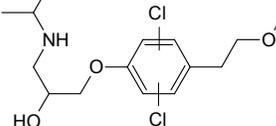
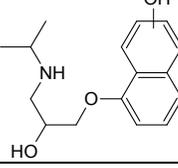
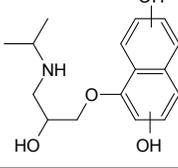
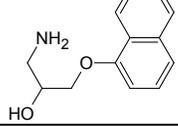
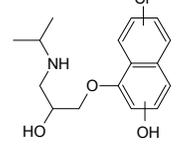
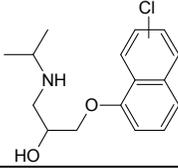
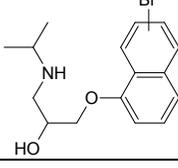
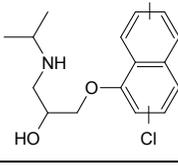
^a Measured or calculated at pH 8.7. ^b $\text{FAC} = [\text{HOCl}] + [\text{OCl}^-]$. ^c n.a.: not available. ^d Calculated at pH 1.3 based on reported acid catalyzed reactions with HOCl.

^e First-order transformation rate is reported due to rapid quenching of the N-chlorinated ciprofloxacin intermediate by Na₂S₂O₃. ^f Reaction rate between HOCl

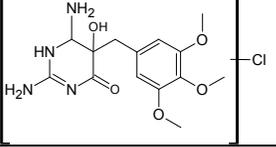
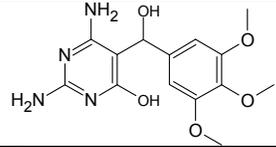
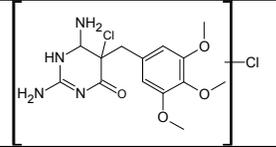
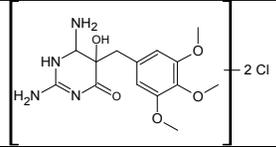
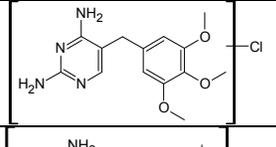
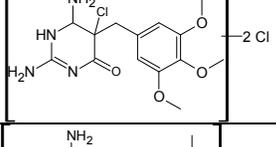
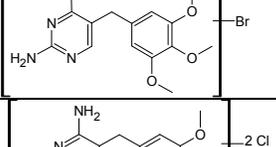
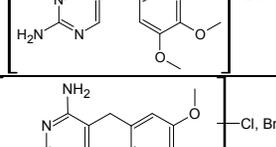
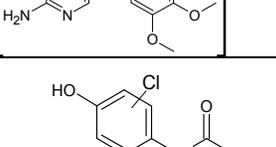
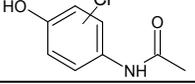
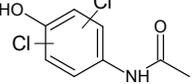
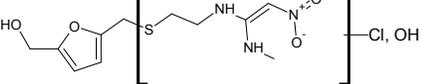
and the predominant fully protonated form of ciprofloxacin at pH 1.3. ^g Estimated based on complete removal within 10 s.

153 Table SI 2. Test Compound Transformation Products (TPs).

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Carbamazepine	224.0715 (Cbz 224)	164.920, 167.073, 180.081, 196.076	0.58	C ₁₄ H ₁₀ NO ₂	0.3		6,7
	180.0812 ^d (Cbz 180)	152.0621	1.17	C ₁₃ H ₁₀ N	0.3		6,8
	275.0800 (Cbz 275)	180.0800, 210.0919, 236.0712, 253.0976	1.58	C ₁₄ H ₁₃ NO ₃	0.6		
	253.0974 ^d (Cbz 253)		1.59	C ₁₅ H ₁₃ N ₂ O ₂	0.3		6,10
	196.0759 ^e (Cbz 196)	167.073, 180.0810	1.65	C ₁₃ H ₁₀ NO	0.3		6-9
	208.0761 ^d (Cbz 208)	152.062, 180.082, 196.0760	2.07	C ₁₄ H ₁₀ NO	0.1		8,9
Metoprolol	226.1436 (Met 226)	116.1067, 121.0636, 149.0583	0.82	C ₁₂ H ₂₀ NO ₃	0.7		11,13
	284.1853 (Met 284)	116.1063, 226.1420, 268.141	0.85	C ₁₅ H ₂₆ NO ₄	0.9		11,13
	254.1393 (Met 254)	105.0692, 151.0381, 177.0540, 212.0911, 236.1275	0.93	C ₁₃ H ₂₀ NO ₄	<0.1		
	238.1439 (Met 238)	105.0696, 133.0642, 149.0590, 161.0589, 196.0962	1.04	C ₁₃ H ₂₀ NO ₃	0.4		12
	282.1706 (Met 282)	105.0696, 133.0642, 149.0590, 161.0589, 196.0962	1.05	C ₁₅ H ₂₄ NO ₄	<0.1		11

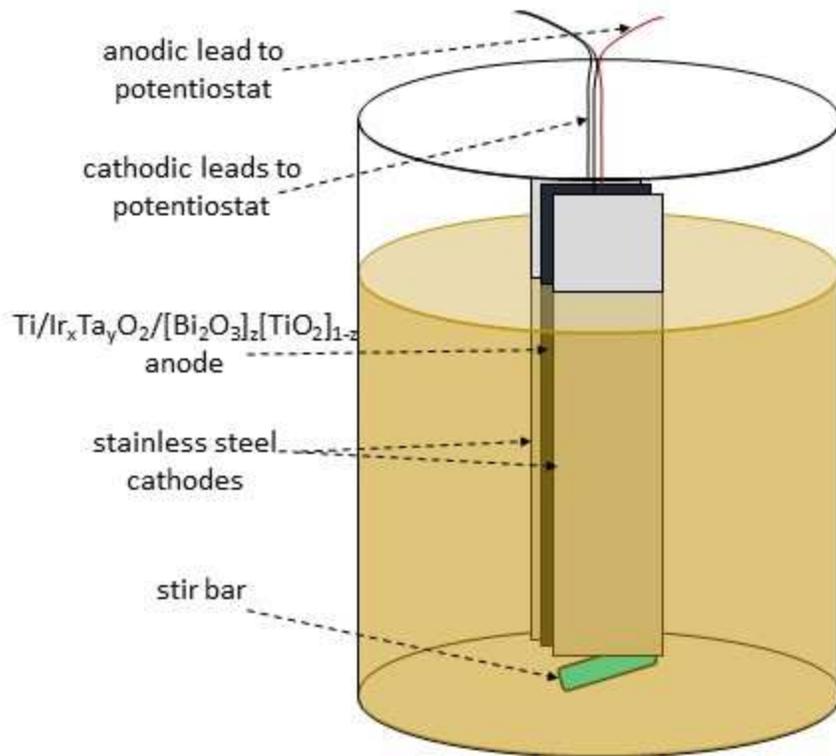
Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Metoprolol (cont.)	302.1515 (Met 302)	98.0963, 116.1066, 167.0249, 193.0407, 260.1044, 284.1411	1.51	C ₁₅ H ₂₅ NO ₃ Cl	0.8		11
	336.1121 (Met 336)	72.0817, 98.0965, 116.1064, 276.0549, 294.0649, 318.102	1.62	C ₁₅ H ₂₄ NO ₃ Cl ₂	1.2		11
Propranolol	276.1599 (Pro 276)	116.1074	1.0-1.3 ^f	C ₁₆ H ₂₂ NO ₃	<0.1		17
	292.1545 (Pro 292)	116.107, 123.092, 150.128, 274.144, 272.128,	1.29	C ₁₆ H ₂₂ NO ₄	0.4		15
	218.1180 (Pro 218)	116.1071, 183.0804	1.45	C ₁₃ H ₁₆ NO ₂	0.1		16
	310.1210 (Pro 310)	72.0816, 116.1071, 135.1038, 260.1653	1.53	C ₁₆ H ₂₁ NO ₃ Cl	<0.1		
	294.1276 (Pro 294)	150.128, 260.165	1.85	C ₁₆ H ₂₁ NO ₂ Cl	1.5		
	338.0739 (Pro 338)	116.1070, 182.0725, 217.0417	1.94	C ₁₆ H ₂₁ NO ₂ Br	1.7		
	328.0868 (Pro 328)	72.0819, 116.1068, 219.1751	1.98	C ₁₆ H ₂₀ NO ₂ Cl ₂	0.3		

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
Ciprofloxacin	306.1246 (Cfx 306)	217.0404, 268.1077, 288.1142	1.09	C ₁₅ H ₁₇ N ₃ O ₃ F	0.8		1,18
	366.1012 (Cfx 366)	225.0213, 245.1079, 279.0695, 322.1108	1.22	C ₁₇ H ₁₈ N ₃ O ₃ FCI	0.9		1
	362.1141 (Cfx 362)	344.1037	1.29	C ₁₇ H ₁₇ N ₃ O ₃ F	1.1		18
	291.0773 (Cfx 291)	245.0709, 273.0671	1.42	C ₁₄ H ₁₂ N ₂ O ₄ F	0.8		
	263.0828 (Cfx 263)	204.032, 245.0724	1.49	C ₁₃ H ₁₂ N ₂ O ₃ F	0.4		18
	298.0287 (Cfx 298)	280.0184	1.63	C ₁₃ H ₁₀ NO ₄ FCI	0.5		1
	289.0981 (Cfx 289)	271.0876	1.74	C ₁₅ H ₁₄ N ₂ O ₃ F	0.8		
	Trimethoprim	342.1327 (Tmp 342)	273.0978, 307.1397	0.98	C ₁₄ H ₂₁ N ₅ O ₃ Cl	0.6	
307.1391 (Tmp 307)		209.1644, 212.0686, 240.0634, 258.0742, 273.0975	0.98	C ₁₄ H ₁₉ N ₄ O ₄	1.5		19
322.1508 (Tmp 322)			1.04	C ₁₄ H ₂₀ N ₅ O ₄	0.7		
325.1499 (Tmp 325a)		173.0813, 219.0872, 233.1043, 293.1229	1.05	C ₁₄ H ₂₁ N ₄ O ₅	1.3		20
343.1164 (Tmp 343)		181.0852	1.08	C ₁₄ H ₂₀ N ₄ O ₄ Cl	0.9		
Trimethoprim							

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
(cont.)	359.1125 (Tmp 359a)	215.0465, 240.0414, 236.0905	1.19	C ₁₄ H ₂₀ N ₄ O ₅ Cl	0.3		
	323.1345 (Tmp 323)		1.26	C ₁₄ H ₁₉ N ₄ O ₅	1.0		
	377.0773 (Tmp 377)	217.0435, 236.0913, 255.0416, 257.0388, 314.0898	1.26	C ₁₄ H ₁₉ N ₄ O ₄ Cl ₂	1.0		21
	393.0719 (Tmp 393)	100.0503, 126.0529, 249.0075, 270.0524	1.32	C ₁₄ H ₁₉ N ₄ O ₅ Cl ₂	1.4		
	325.1059 (Tmp 325b)	145.0267, 271.0918, 249.008	1.39	C ₁₄ H ₁₈ N ₄ O ₃ Cl	0.8		21
	411.0391 (Tmp 411)	249.0080, 366.0616	1.39	C ₁₄ H ₁₈ N ₄ O ₄ Cl ₃	0.3		21
	369.0558 (Tmp 369)	255.0886, 273.092, 325.1078, 339.0081	1.42	C ₁₄ H ₁₈ N ₄ O ₃ Br	0.4		
	359.0672 (Tmp 359b)	181.0854, 199.097, 289.0480, 298.0371, 329.0196	1.55	C ₁₄ H ₁₇ N ₄ O ₃ Cl ₂	0.6		21
	403.0174 (Tmp 403)	140.0686, 180.8981, 257.1478, 359.0669	1.58	C ₁₄ H ₁₇ N ₄ O ₃ BrCl	0.1		
Acetaminophen	186.0312 (Ace 186)	109.0515, 144.0201, 155.9838	1.17	C ₈ H ₉ NO ₂ Cl	1.0		22,23
	219.9919 (Ace 219)	143.0123, 177.9810	1.42	C ₈ H ₈ NO ₂ Cl ₂	1.3		22,23
Ranitidine	338.0561 (Rnt 338)	109.0278, 186.0565, 202.9997, 214.0240, 292.0612	0.63	C ₁₁ H ₁₇ N ₃ O ₅ SCl	1.6		25

Parent Test compound	TP m/z ^a (ID)	Major Fragment ions m/z	RT (min)	Calculated Formula	Mass Error (mDa)	Proposed Molecular Ion Structure ^b	Refs. ^c
	336.0419 (Rnt 336)	125.0600, 181.0201, 203.0023, 214.0259, 290.0497	0.66	C ₁₁ H ₁₅ N ₃ O ₃ SCl	0.2		
	364.1194 (Rnt 364)	109.0278, 163.0854, 189.1012, 258.0579	0.76	C ₁₃ H ₂₃ N ₅ O ₃ SCl	1.6		25
	335.0928 (Rnt 335)		0.85	C ₁₂ H ₂₀ N ₄ O ₃ SCl	1.6		
	349.1083 (Rnt 349)	124.075, 132.0438, 164.0159, 210.0092, 223.0893	0.88	C ₁₃ H ₂₂ N ₄ O ₃ SCl	1.8		25
	270.0903 (Rnt 270)	97.0759, 165.1013, 191.1167, 224.0971	1.02	C ₁₁ H ₁₆ N ₃ O ₃ S	0.9		
	322.061 (Rnt 322)	165.0236, 240.1152	1.08	C ₁₁ H ₁₇ N ₃ O ₄ SCl	1.8		
	320.0459 (Rnt 320)	109.028, 165.0238, 208.0392, 234.0191, 274.0529	1.12	C ₁₁ H ₁₅ N ₃ O ₄ SCl	1.3		
	294.12575 (Rnt 294)	109.0294, 150.1284, 177.0489, 188.0054, 275.053	1.85	C ₁₃ H ₂₅ NO ₂ SCl	3.7		
Cimetidine	117.0210 (Cmt 117)	90.0104	0.53	C ₄ H ₆ N ₂ Cl	1.0		24
	189.0435 (Cmt 189)	82.0403	1.06	C ₅ H ₉ N ₄ O ₂ S	1.1		24

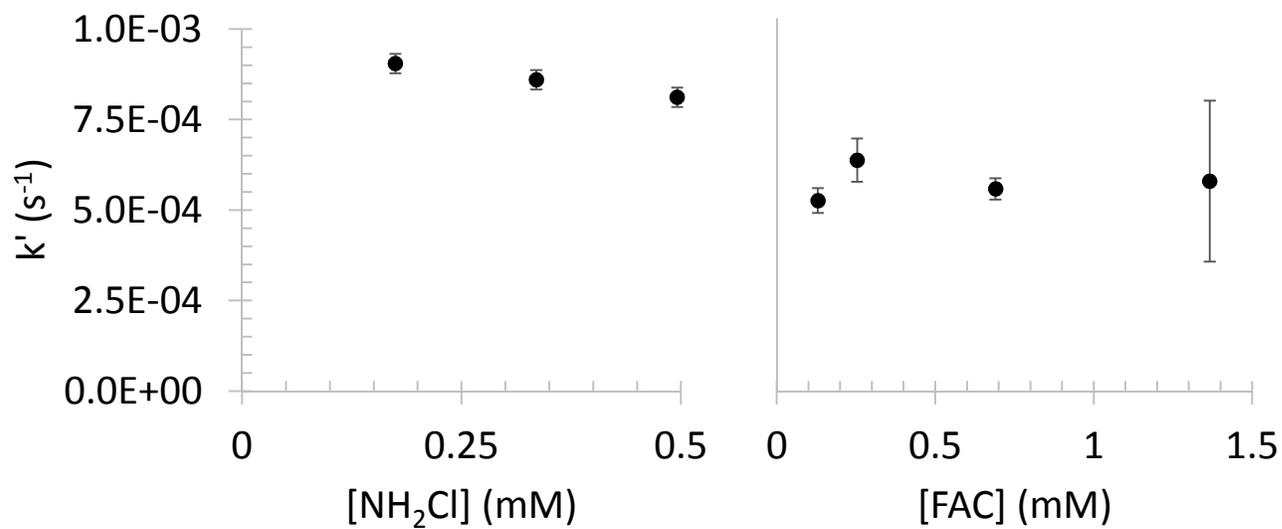
154 ^a Compounds with m/z ratios in bold exhibited isotopic patterns consistent with the halogen atoms specified in the
155 calculated formula (i.e., m/z+2, m/z+4, etc. peaks). ^b Structure of unprotonated TP. Structure based on calculated
156 formula, fragments, and literature. ^c References in bold proposed compound structure shown. References in italics
157 observed similar major fragment ions. ^d TP identity confirmed based on authentic standard retention time and mass
158 spectrum. ^e Authentic acridone standard (ketone product shown) retention time differed by ~0.03 min and had
159 different fragmentation pattern, suggesting that hydroxyacridone was formed during electrolysis. ^f Multiple peaks
160 observed.
161



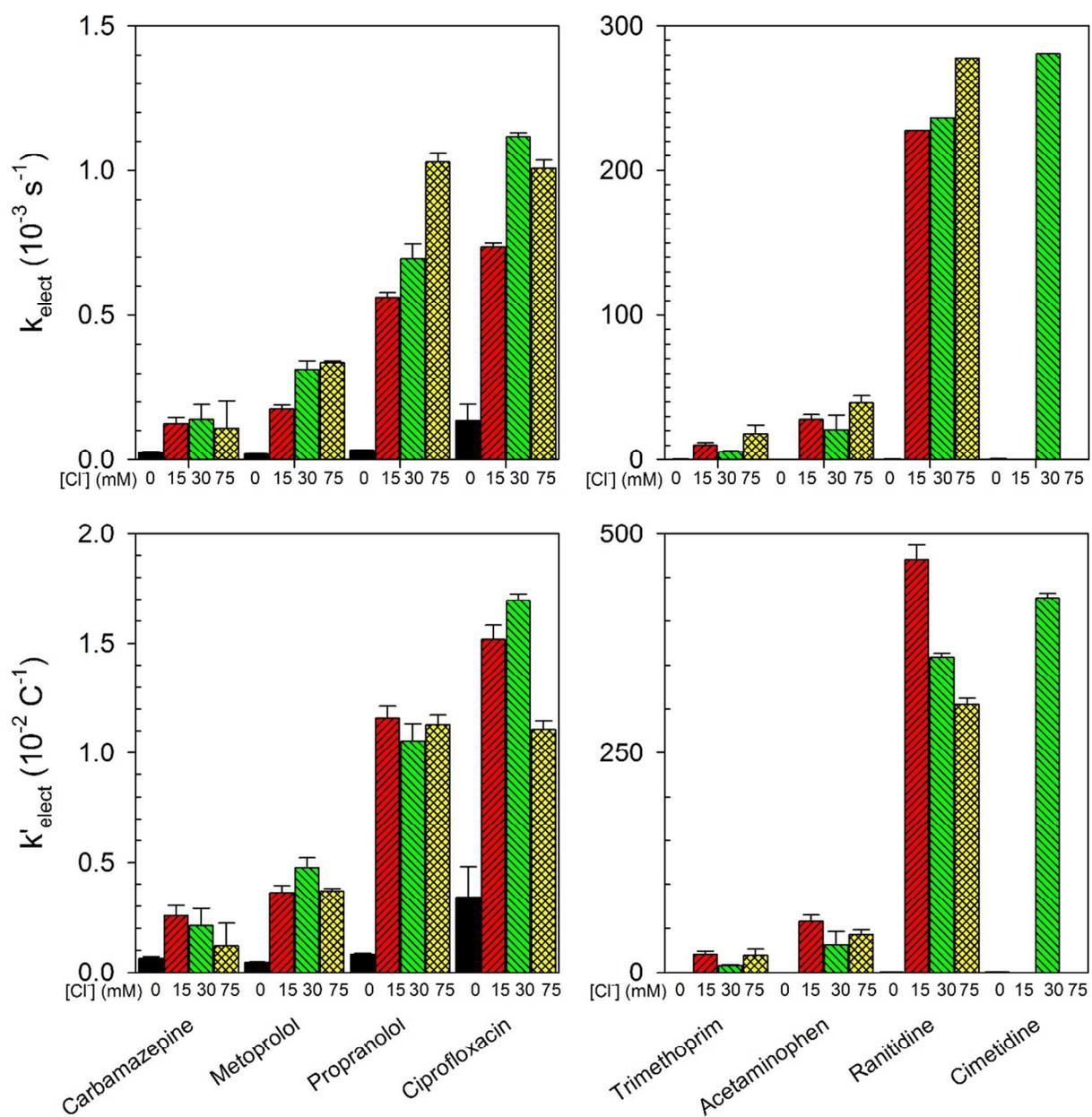
162

163 Figure SI 1. Schematic of electrochemical reactor used in this study.

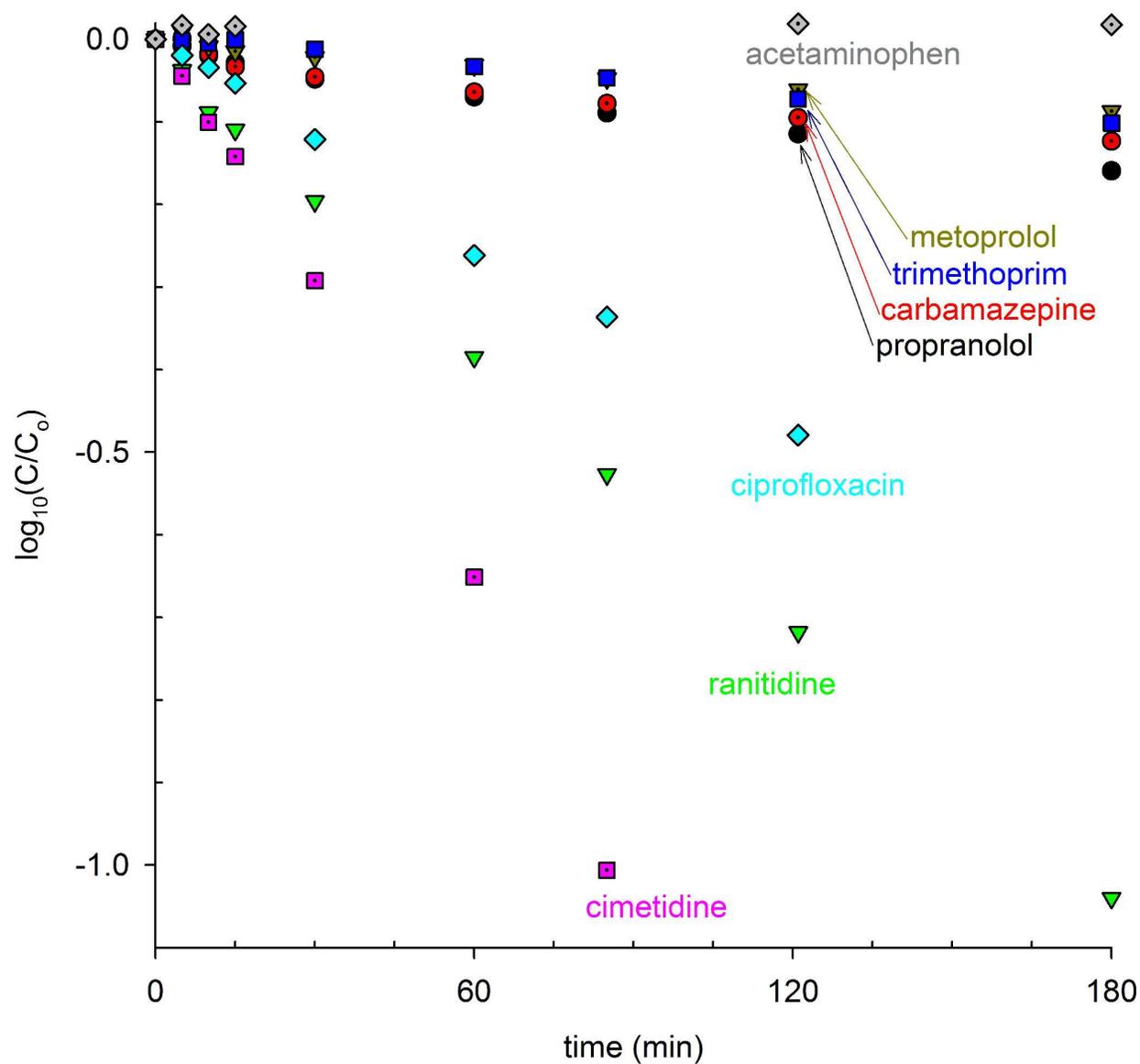
164



165 Figure SI 2. Measured first-order rate constants for the reaction of ciprofloxacin with various
166 concentrations of NH_2Cl and FAC. Error bars represent \pm one standard deviation.



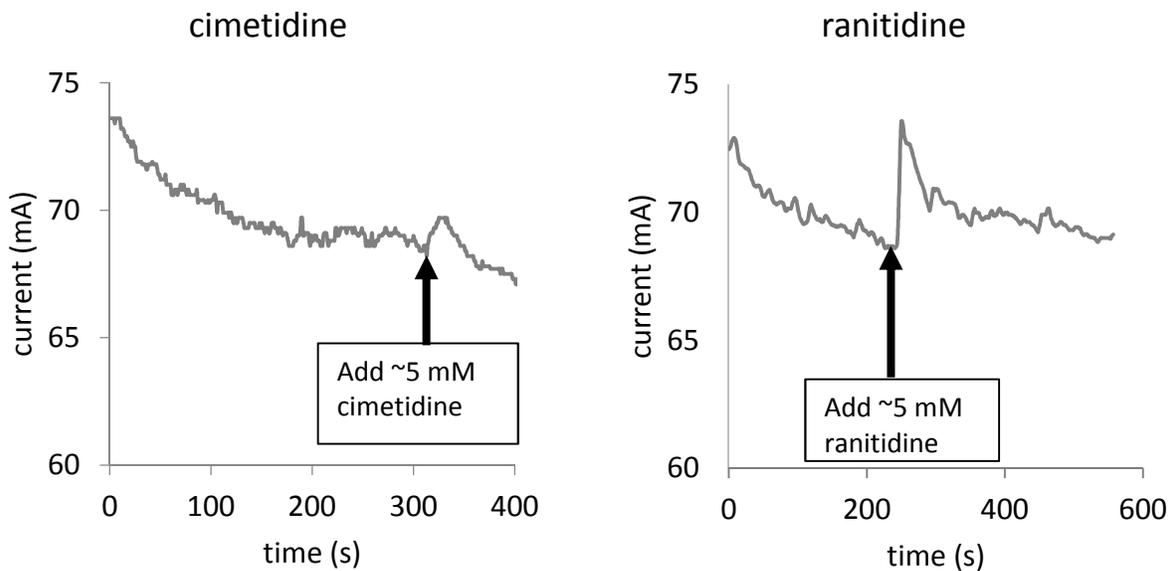
167
 168 Figure SI 3. Pseudo-first order test compound electrolysis rates (k ; top) and rates normalized for
 169 charge (k' ; bottom). Solutions were electrolyzed at an applied potential of 3.5 V in 0-75 mM
 170 NaCl solution buffered at pH 8.7 (20 mM borate). Average current densities: 0 mM, 0.6 A L^{-1} ;
 171 15 mM, 0.7 A L^{-1} ; 30 mM, 0.9 A L^{-1} ; 75 mM, 1.3 A L^{-1} . FAC production rates: 15 mM,
 172 $0.9 \mu\text{M s}^{-1}$; 30 mM, $1.6 \mu\text{M s}^{-1}$; 75 mM, $3 \mu\text{M s}^{-1}$. Error bars represent \pm one standard deviation.
 173



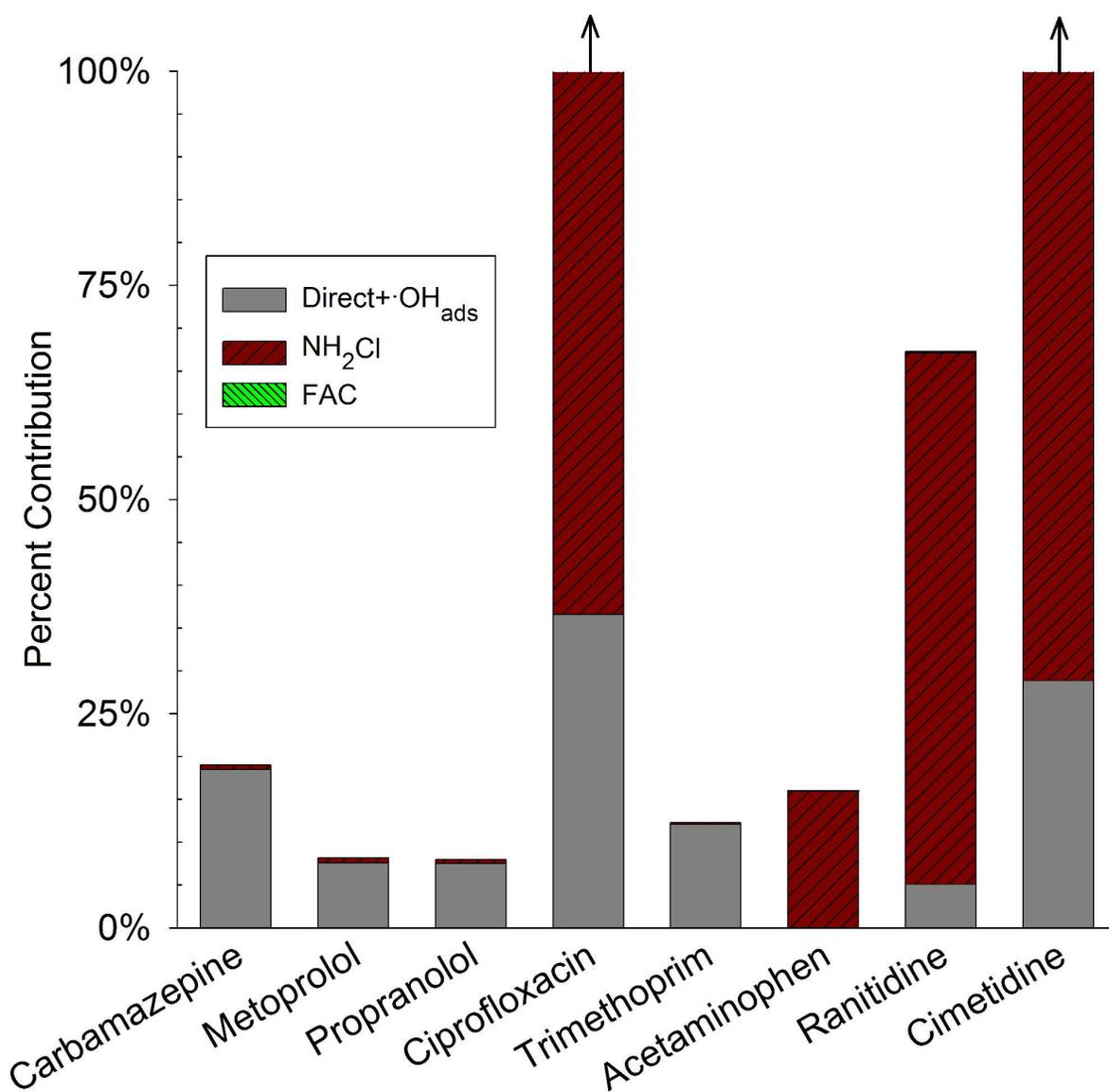
174

175 Figure SI 4. Test compound removal during electrolysis (3.5 V; 0.6 A L⁻¹) in water buffered at
 176 pH 8.7 (20 mM borate).

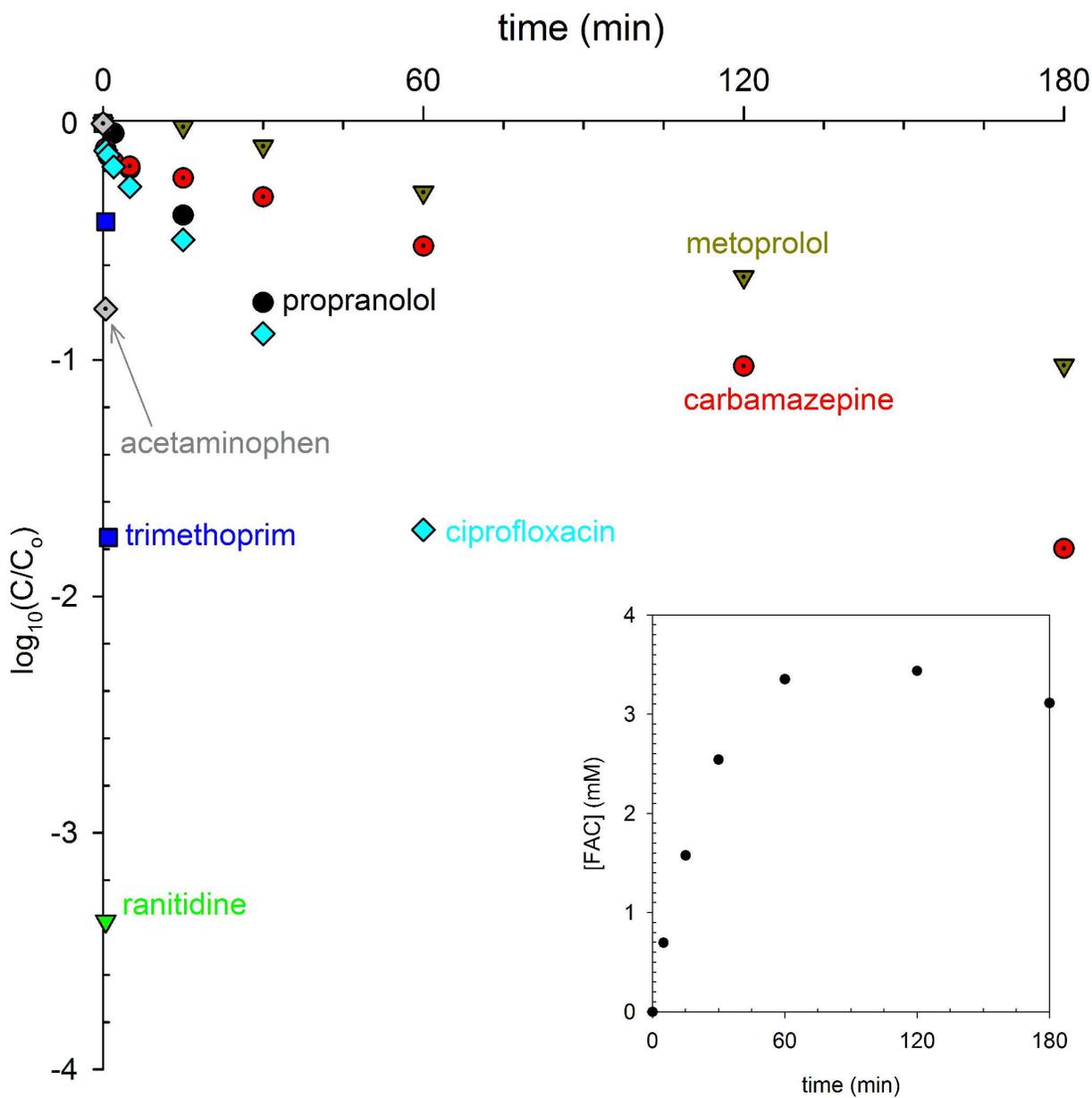
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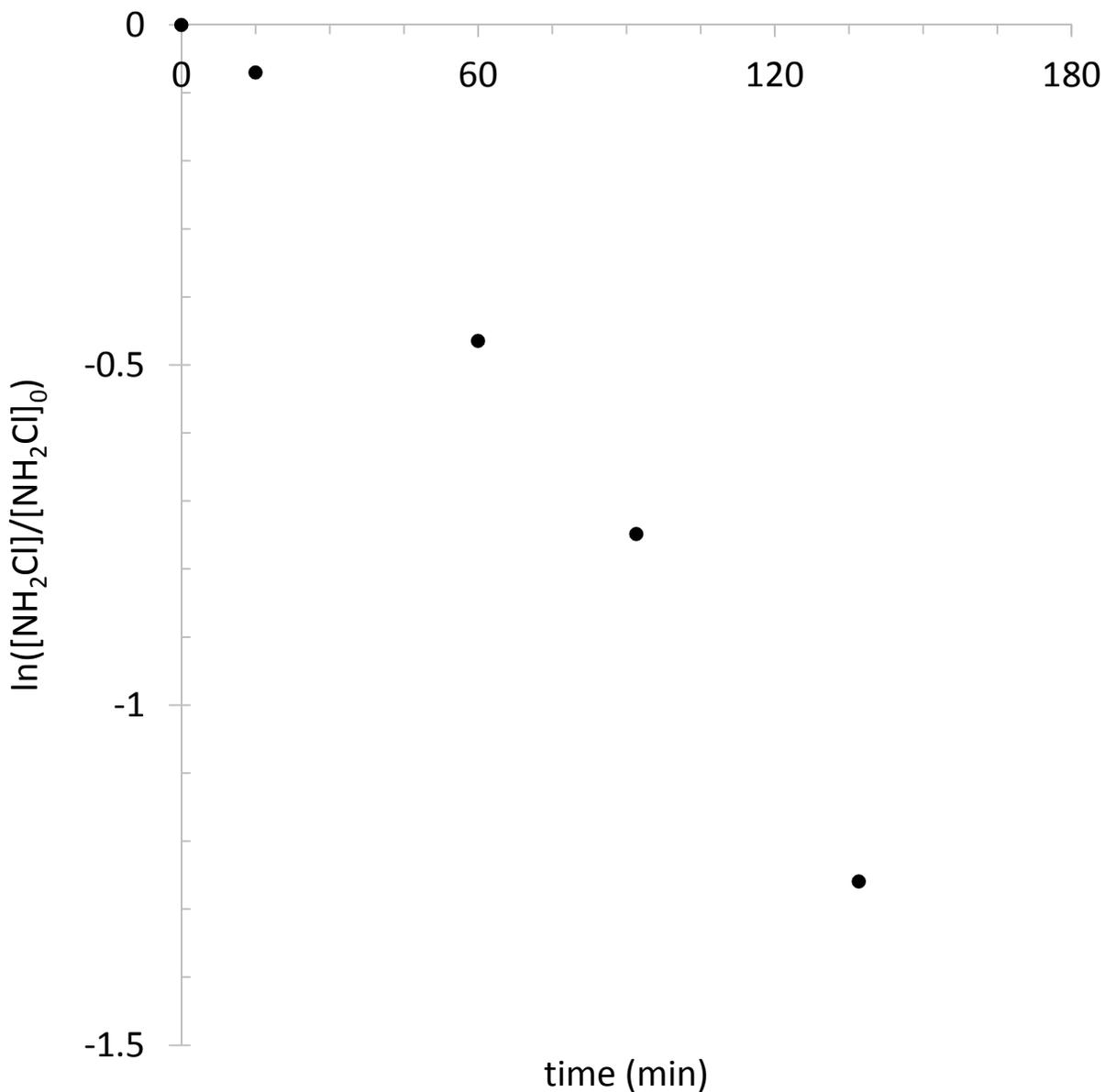
178
179 Figure SI 5. Current during addition of solid test compounds to solutions of Na_2SO_4 (20 mM)
180 undergoing electrolysis (3.5 V). Spikes in current upon test compound addition imply direct
181 electron transfer from the compound.
182



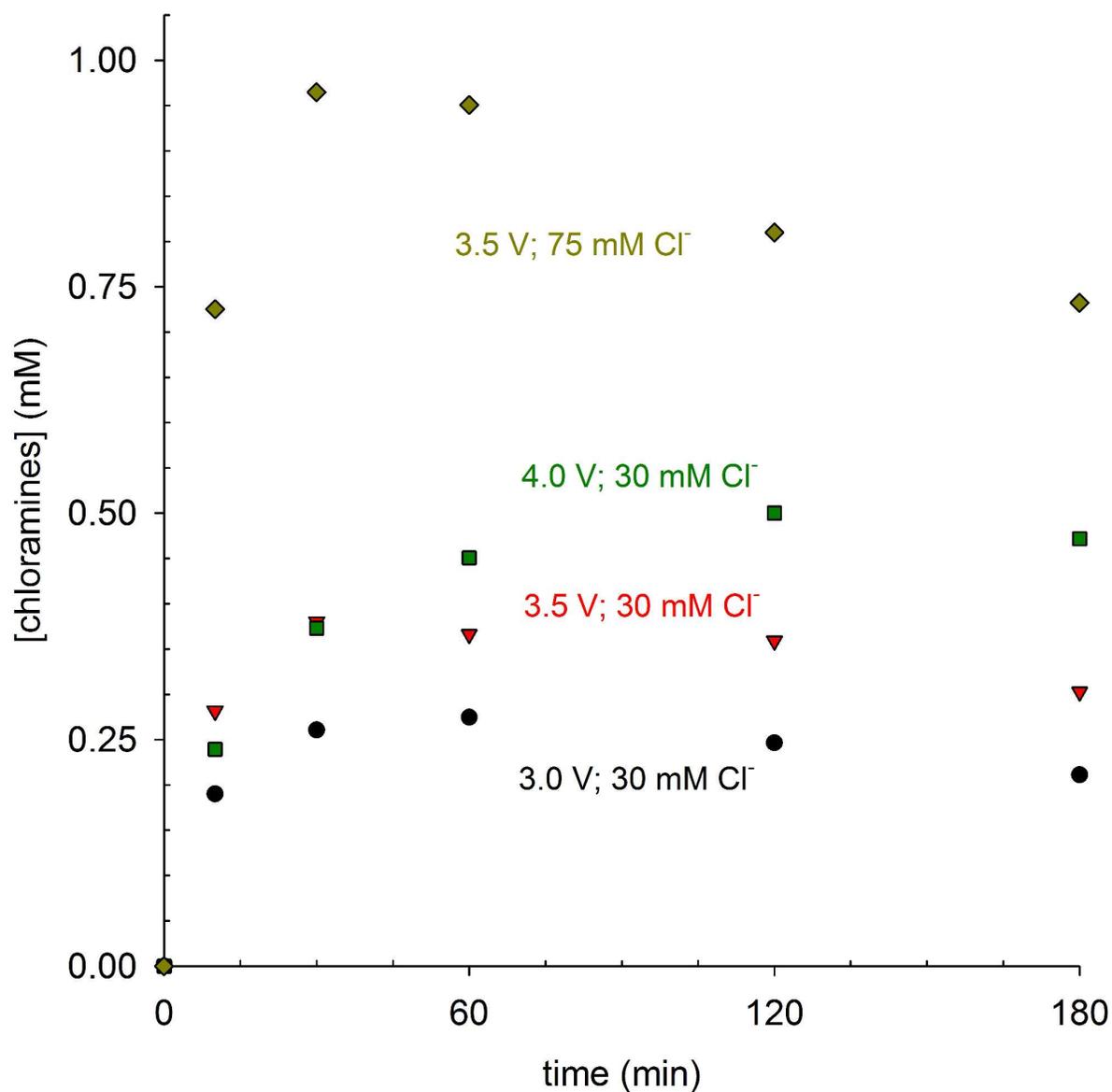
183
 184 Figure SI 6. Calculated contributions of transformation mechanisms to test compound removal
 185 in latrine wastewater electrolyzed at 3.5 V (0.7 A L⁻¹). Ciprofloxacin and cimetidine removal
 186 rates were over-predicted by reaction with NH₂Cl.



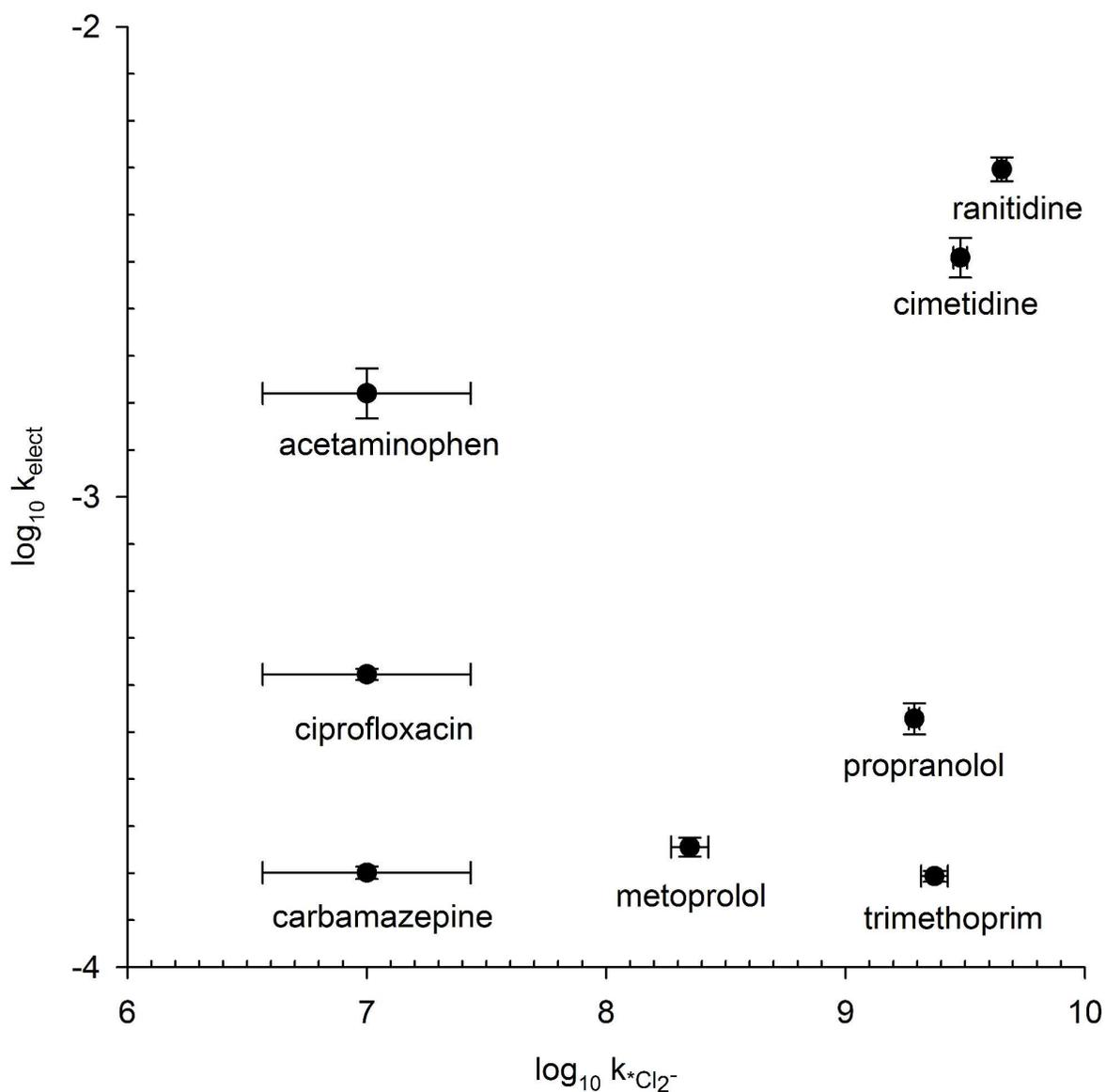
187
 188 Figure SI 7. Test compound removal during electrolysis (3.5 V; 0.9 A L⁻¹) in 30 mM NaCl
 189 solution buffered at pH 8.7 (20 mM borate). Inset: [FAC] ([HOCl]+[OCl⁻]) during electrolysis.
 190 Note that FAC concentrations plateaued after ~60 min due to FAC reduction on the cathode.²⁸
 191



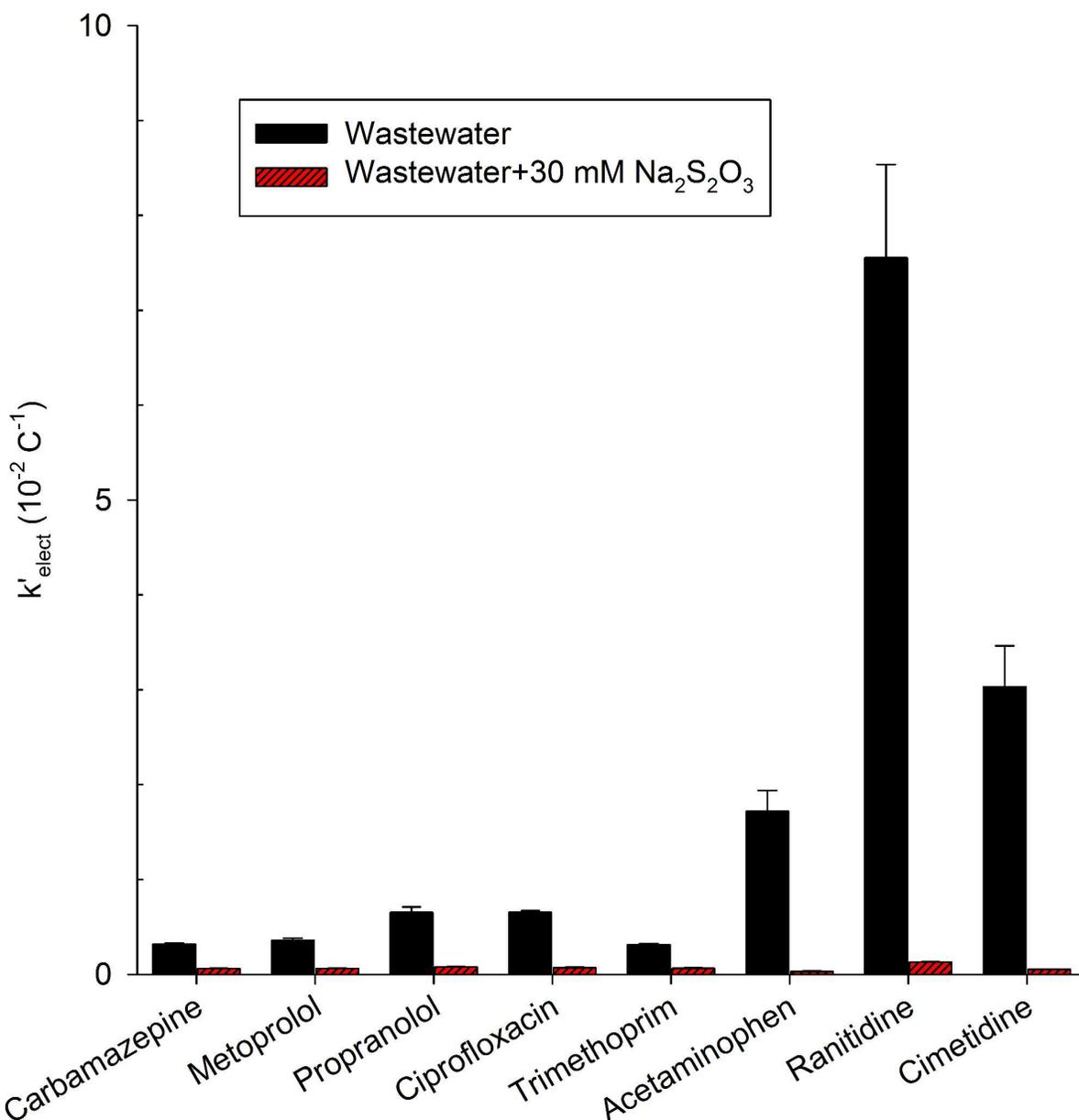
192
 193 Figure SI 8. NH_2Cl reduction in buffered solution (pH 8.7) on a stainless steel cathode
 194 ($k=0.02\ C^{-1}$). The experiment was performed in an H cell with anodic and cathodic chambers
 195 separated by a Nafion membrane to prevent formation of chloramines at the anode. The cathodic
 196 potential was controlled at a similar potential to that observed during wastewater electrolysis
 197 (i.e., about -1.5 V vs. an Ag/AgCl/Sat. NaCl reference electrode; 10 mA). $[NH_2Cl]_0=1\ mM$.
 198 Migration of NH_2Cl through the Nafion membrane was insignificant over the 2 hr experiment.



199
 200 Figure SI 9. Chloramine concentration during electrolysis of latrine wastewater at various
 201 applied potentials (3.0-4.0 V) and chloride concentrations (30 mM; 75 mM). Average current
 202 densities: 3.0 V, 0.5 A L⁻¹; 3.5 V, 0.7 A L⁻¹; 4.0 V, 1.3 A L⁻¹; 3.5 V 75 mM Cl⁻, 1.1 A L⁻¹.



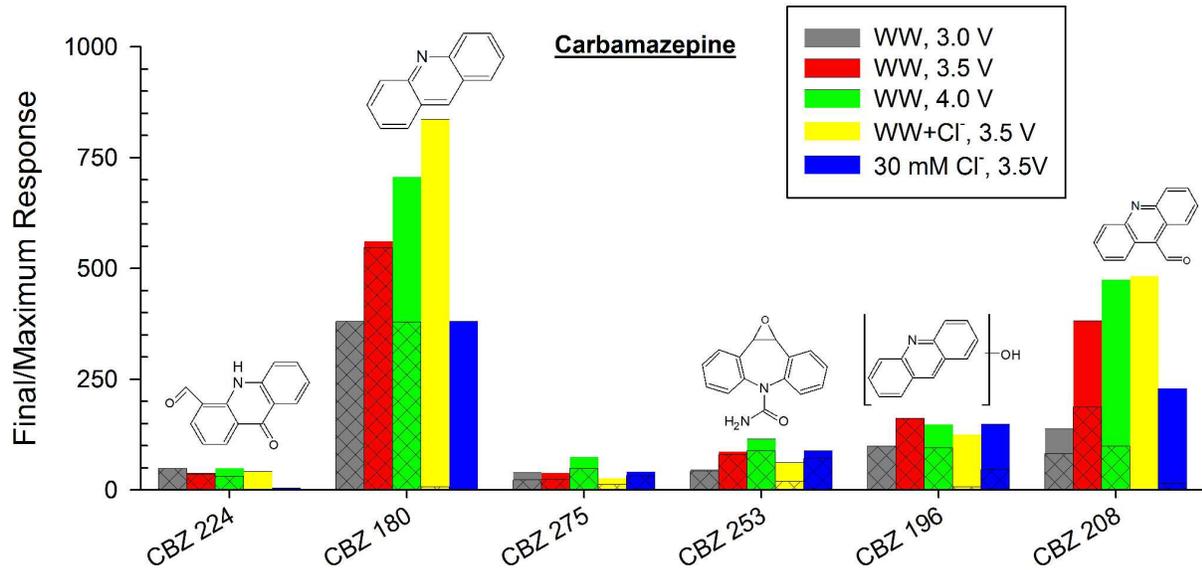
203
 204 Figure SI 10. Correlation between test compound pseudo-first order electrolysis rates (k_{elect} ; s^{-1})
 205 in latrine wastewater at an applied potential of 3.5 V and measured second-order reaction rates
 206 with $\cdot\text{Cl}_2^-$ ($k_{\text{Cl}_2^-}$; $\text{M}^{-1} \text{s}^{-1}$). $\cdot\text{Cl}_2^-$ reaction rates with compounds that were too slow to measure by
 207 flash photolysis (carbamazepine, ciprofloxacin, and acetaminophen) were assumed to be between
 208 $10^{6.5}$ and $10^{7.5} \text{ M}^{-1} \text{ s}^{-1}$. Error bars represent \pm one standard deviation.
 209



210
 211 Figure SI 11. Pseudo-first order test compound electrolysis rates normalized for charge (k') in
 212 latrine wastewater with or without added $\text{Na}_2\text{S}_2\text{O}_3$. Solutions were electrolyzed at an applied
 213 potential of 3.5 V. Average current densities: Wastewater, 0.7 A L^{-1} ; Wastewater+30 mM
 214 $\text{Na}_2\text{S}_2\text{O}_3$, 0.9 A L^{-1} . Average chloramine concentration: Wastewater, 0.3 mM; Wastewater+30
 215 mM $\text{Na}_2\text{S}_2\text{O}_3$, 0 mM. Error bars represent \pm one standard deviation.

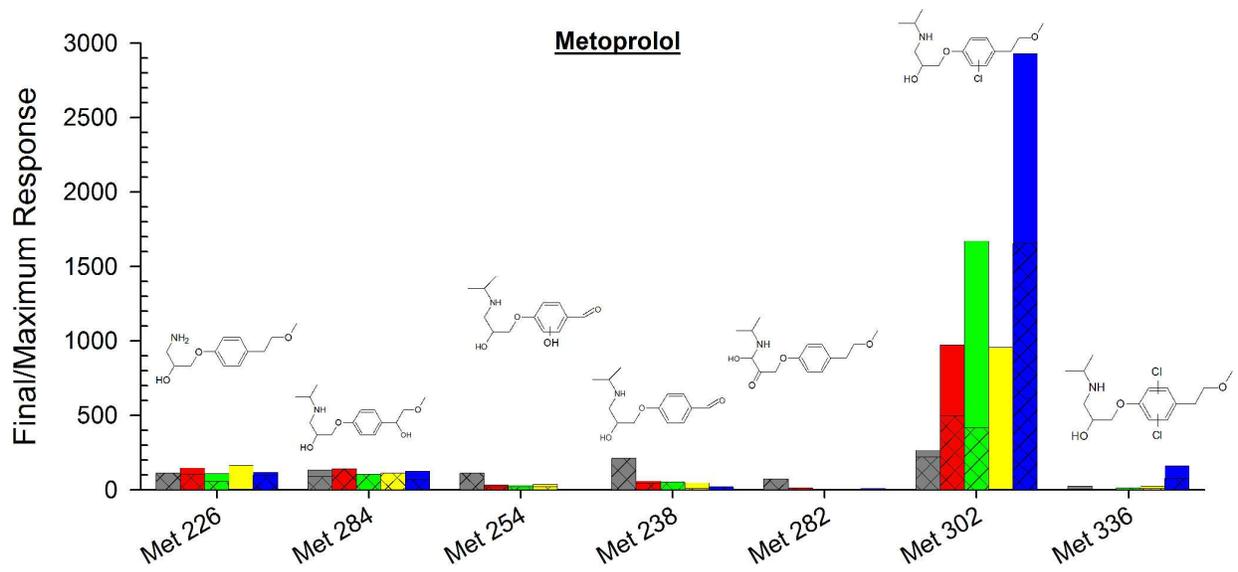
216

217 a



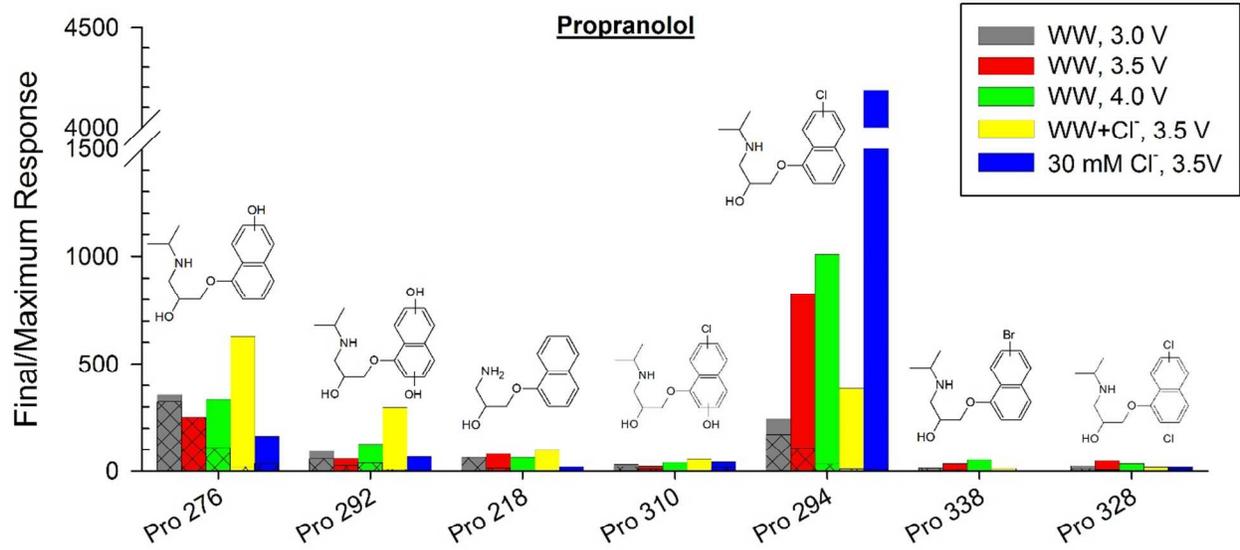
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219 b



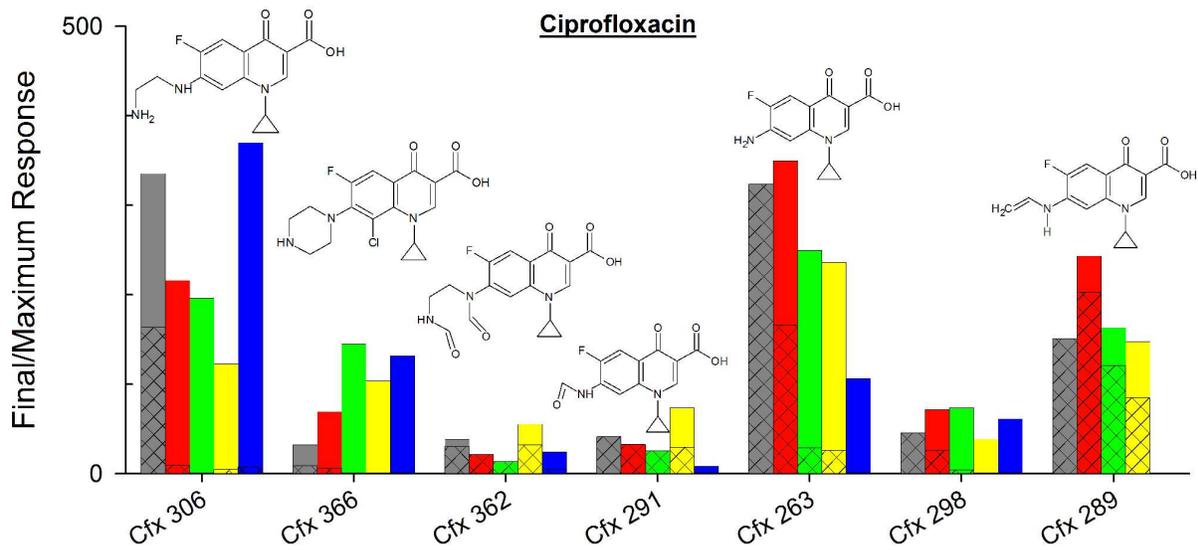
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221 c



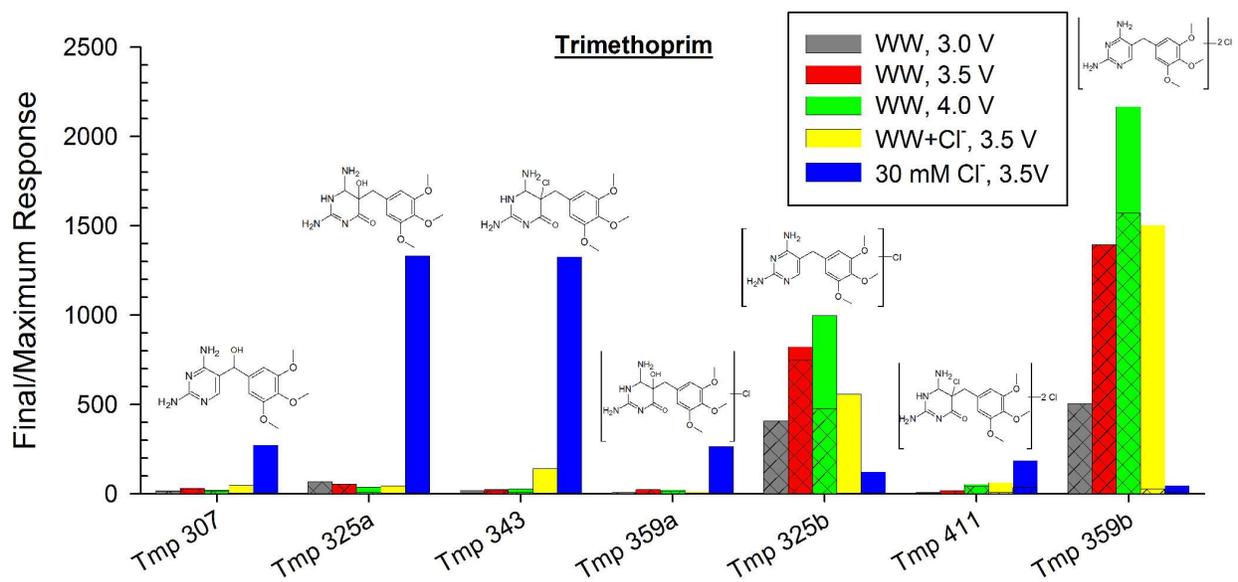
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223 d



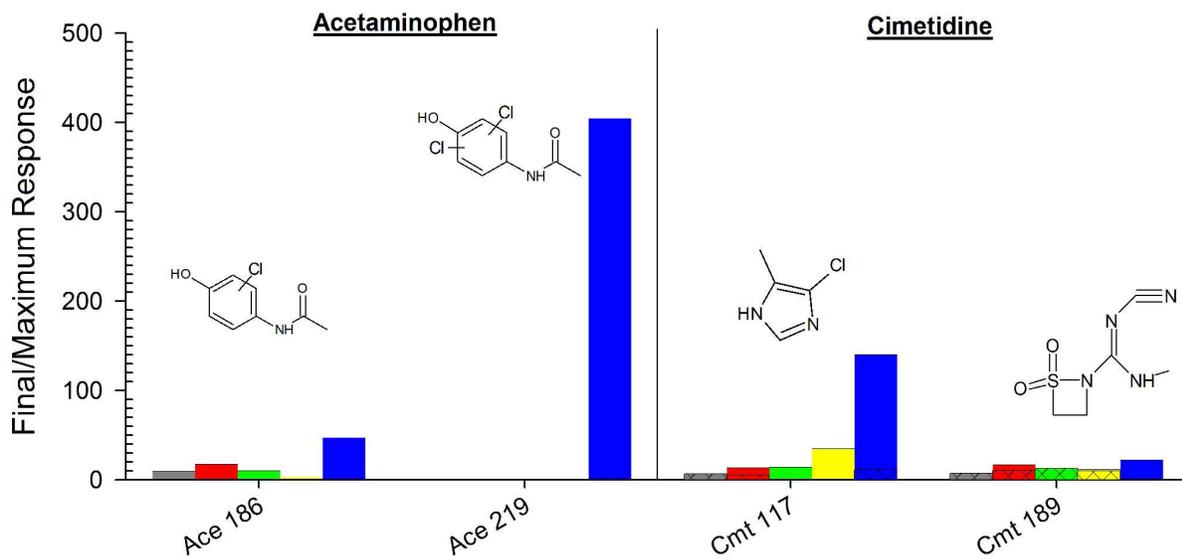
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225 e

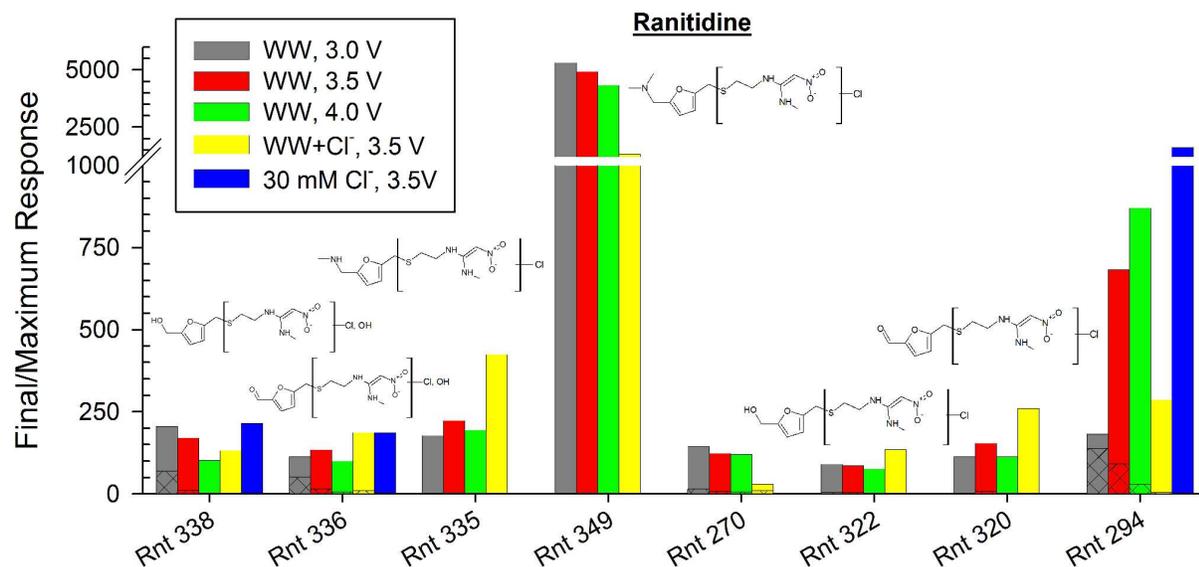


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227 f

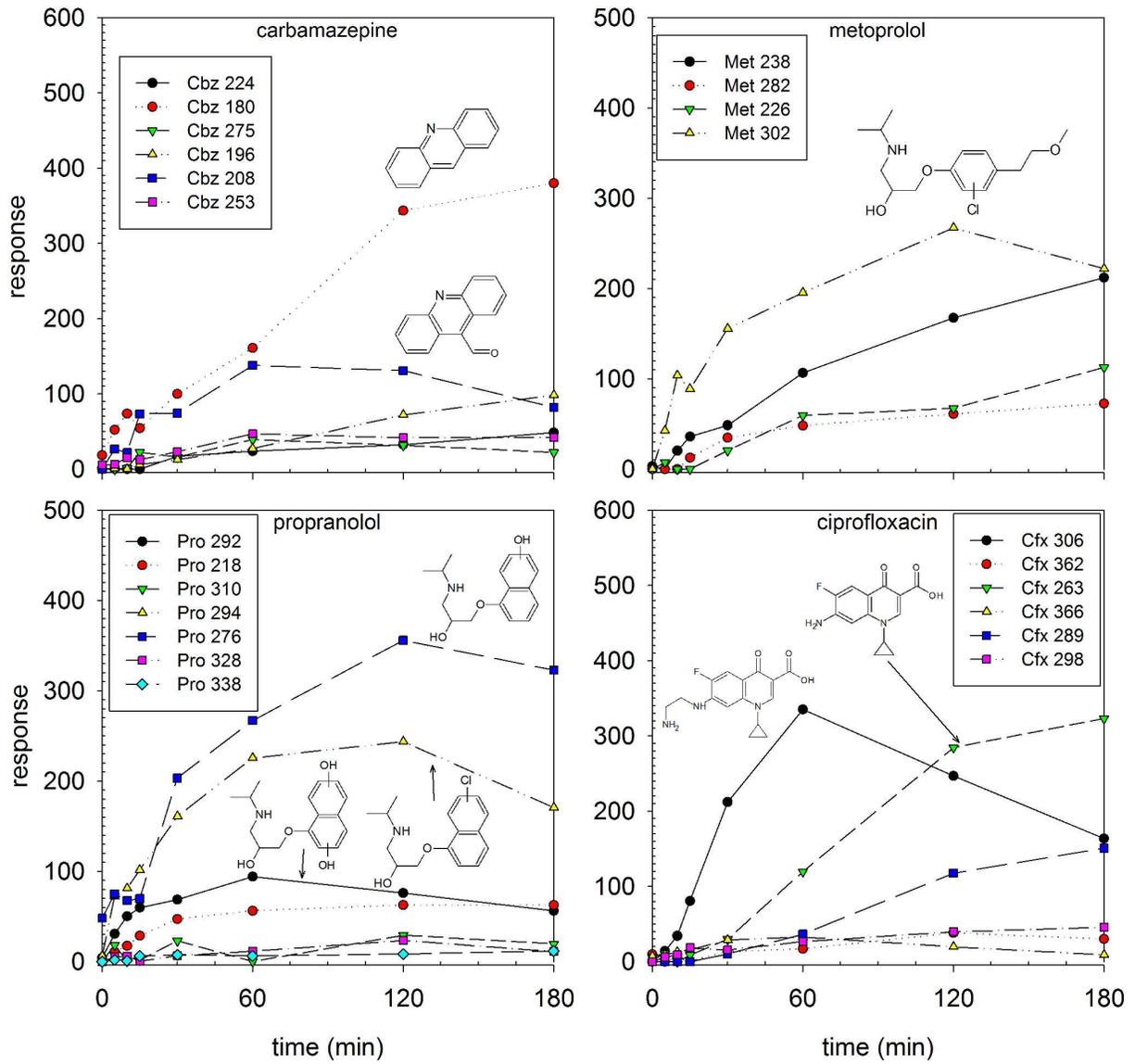


228



230
 231 Figure SI 12. Maximum responses of predominant transformation product formed during
 232 electrolysis (entire bar) and final amount remaining at end of electrolysis cycle (cross-hatched
 233 portion). Electrolysis cycle was 180 min. Transformation product IDs represent the nominal
 234 mass to charge ratio for each compound. Structures are proposed based on accurate mass to
 235 charge ratios, fragments, and transformation products reported in the literature. Further
 236 transformation product information is given in Table SI 2.
 237

3.0 V, 30 mM Cl⁻

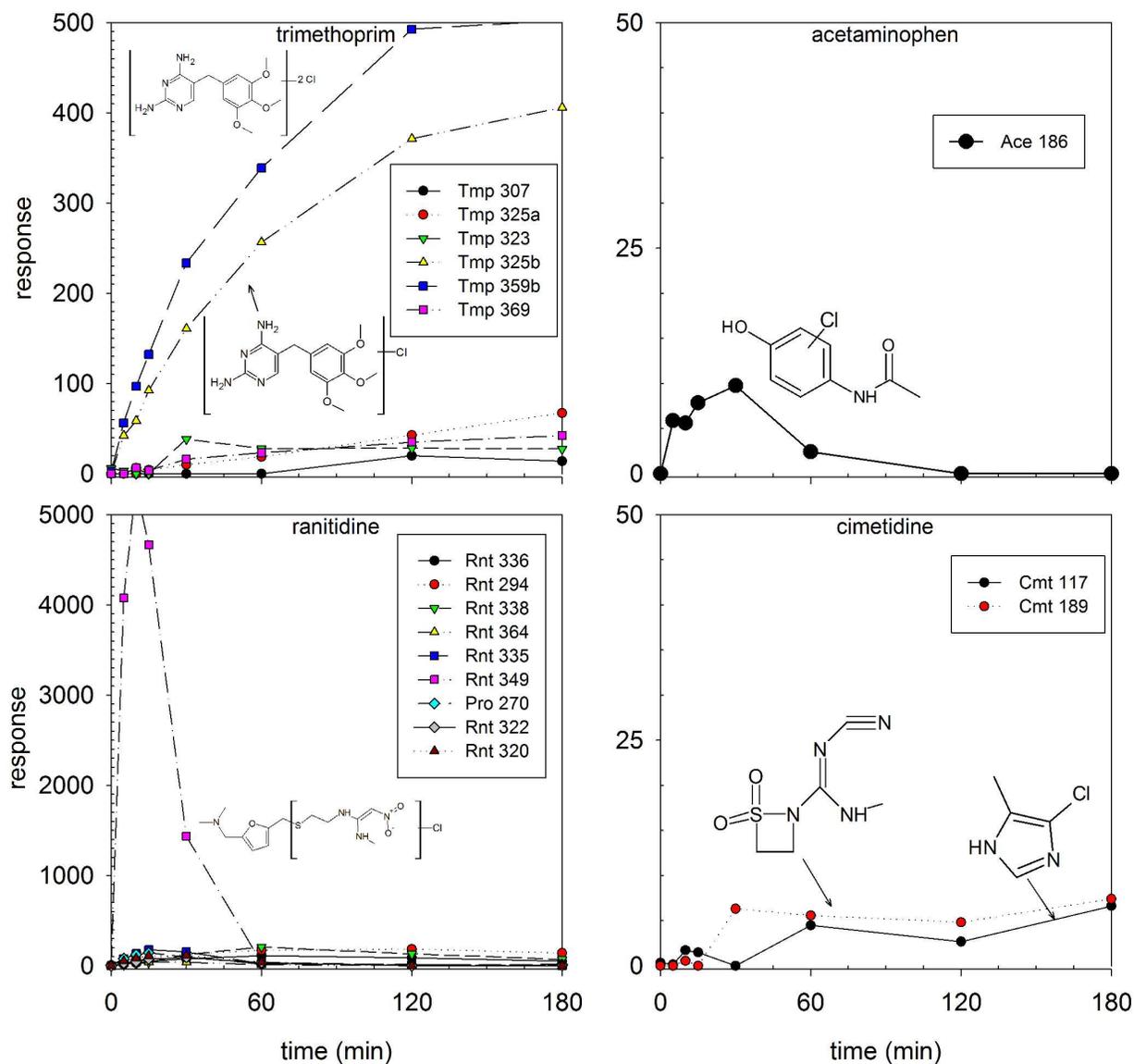


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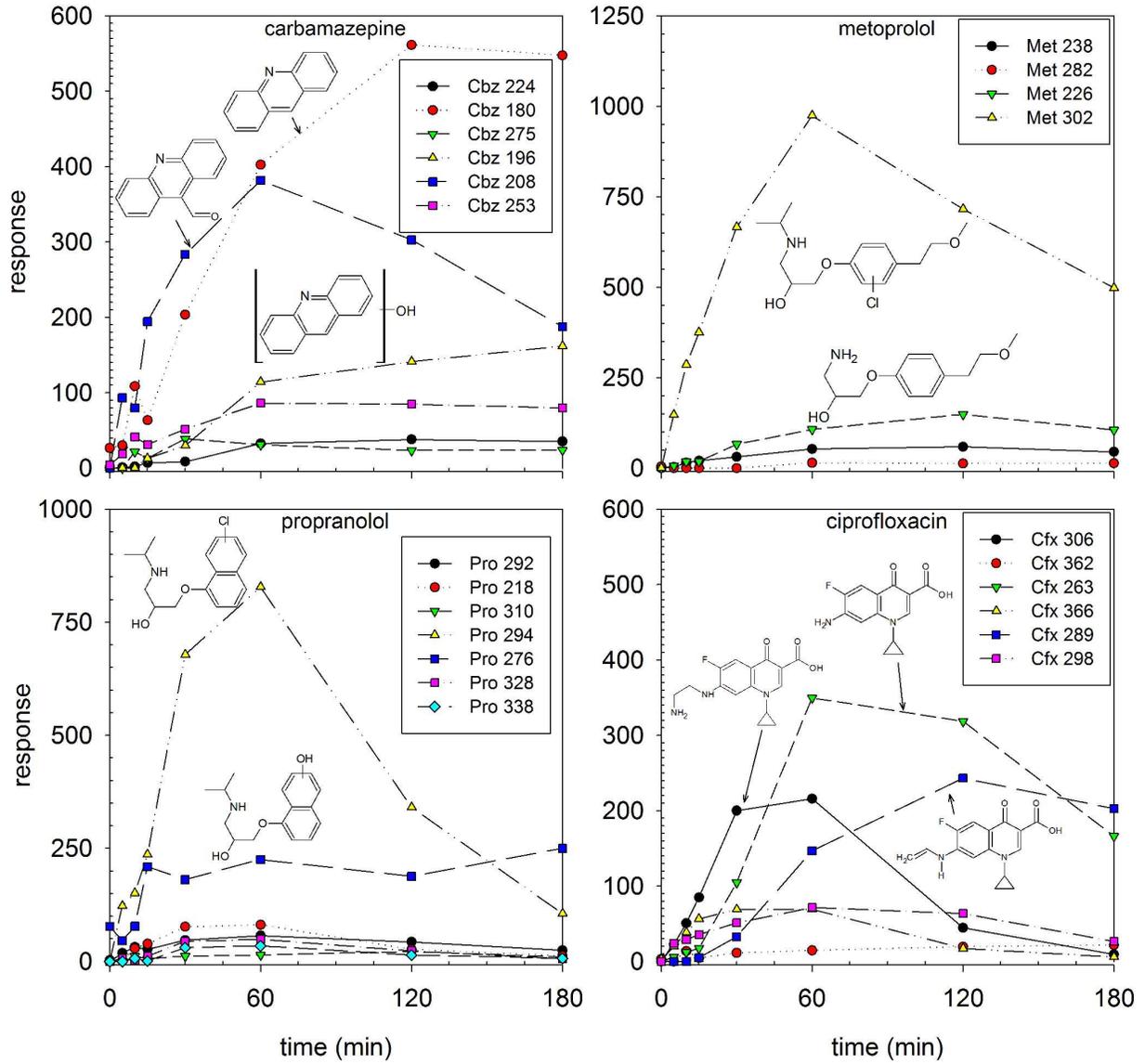
241

3.0 V, 30 mM Cl⁻

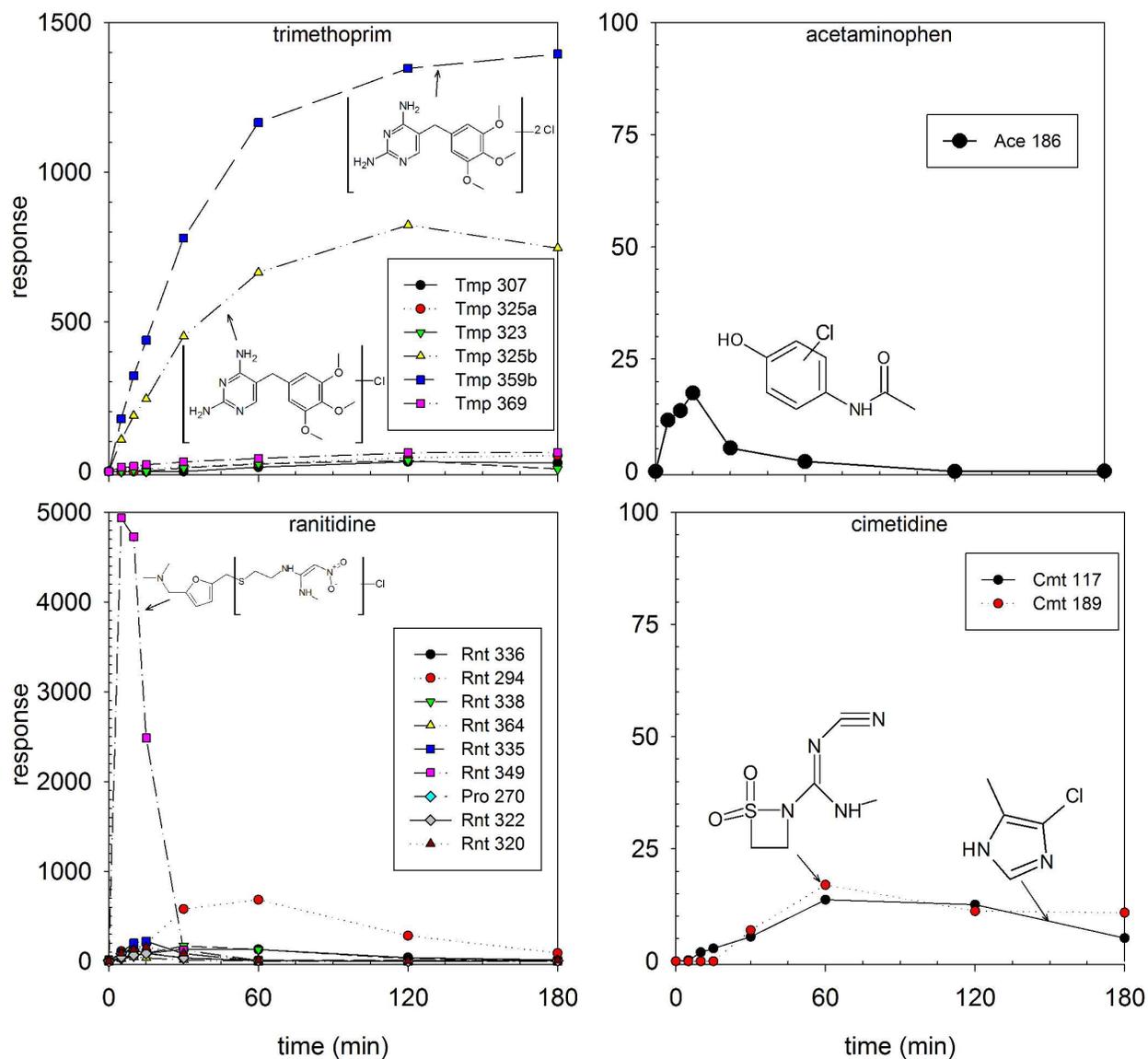


242
 243 Figure SI 13a. Test compound transformation product formation during latrine wastewater
 244 electrolysis at 3.0 V. Transformation product IDs represent the nominal mass to charge ratio for
 245 each compound. Further transformation product information is given in Table SI 2.

3.5 V, 30 mM Cl⁻



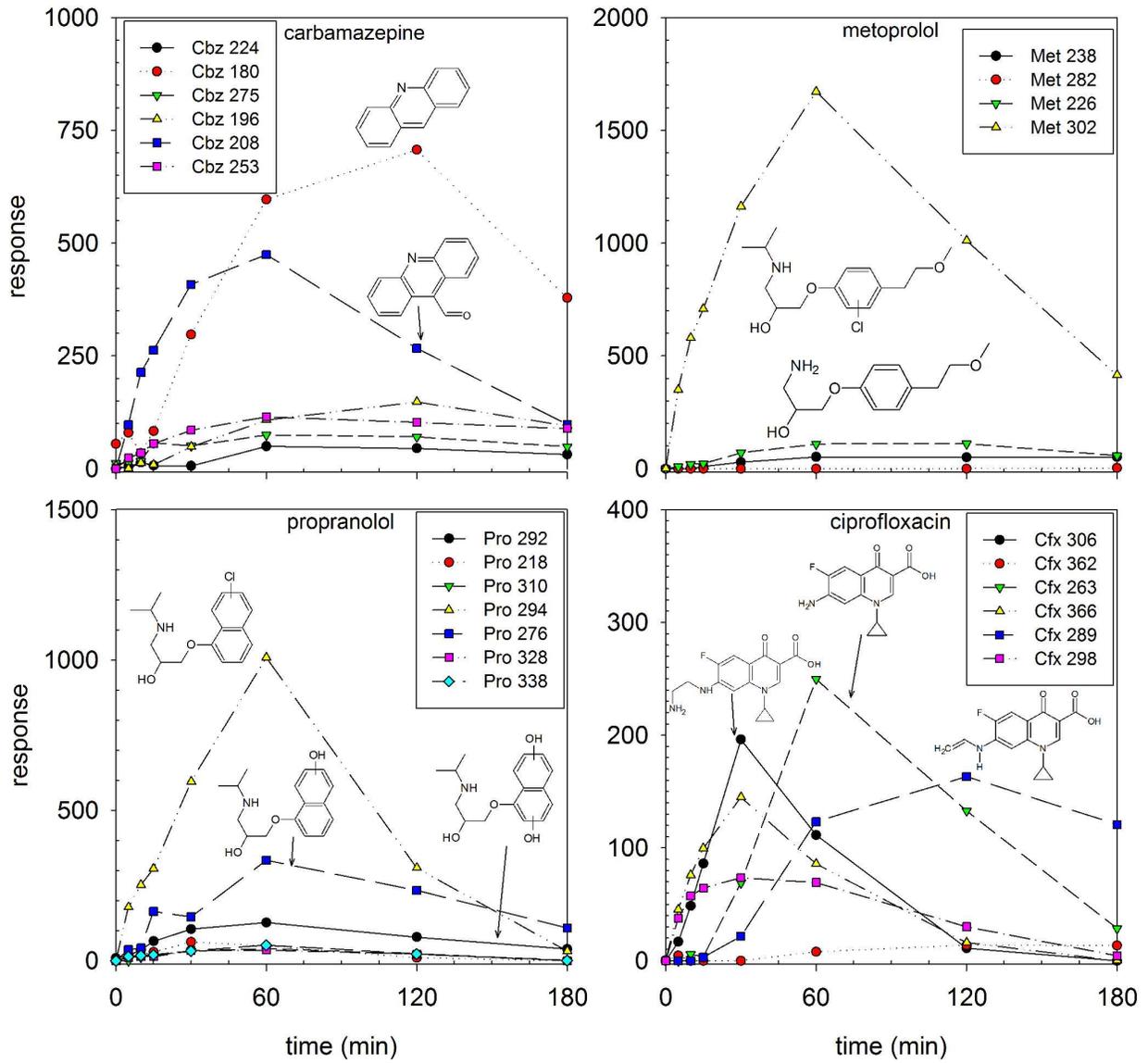
3.5 V, 30 mM Cl⁻



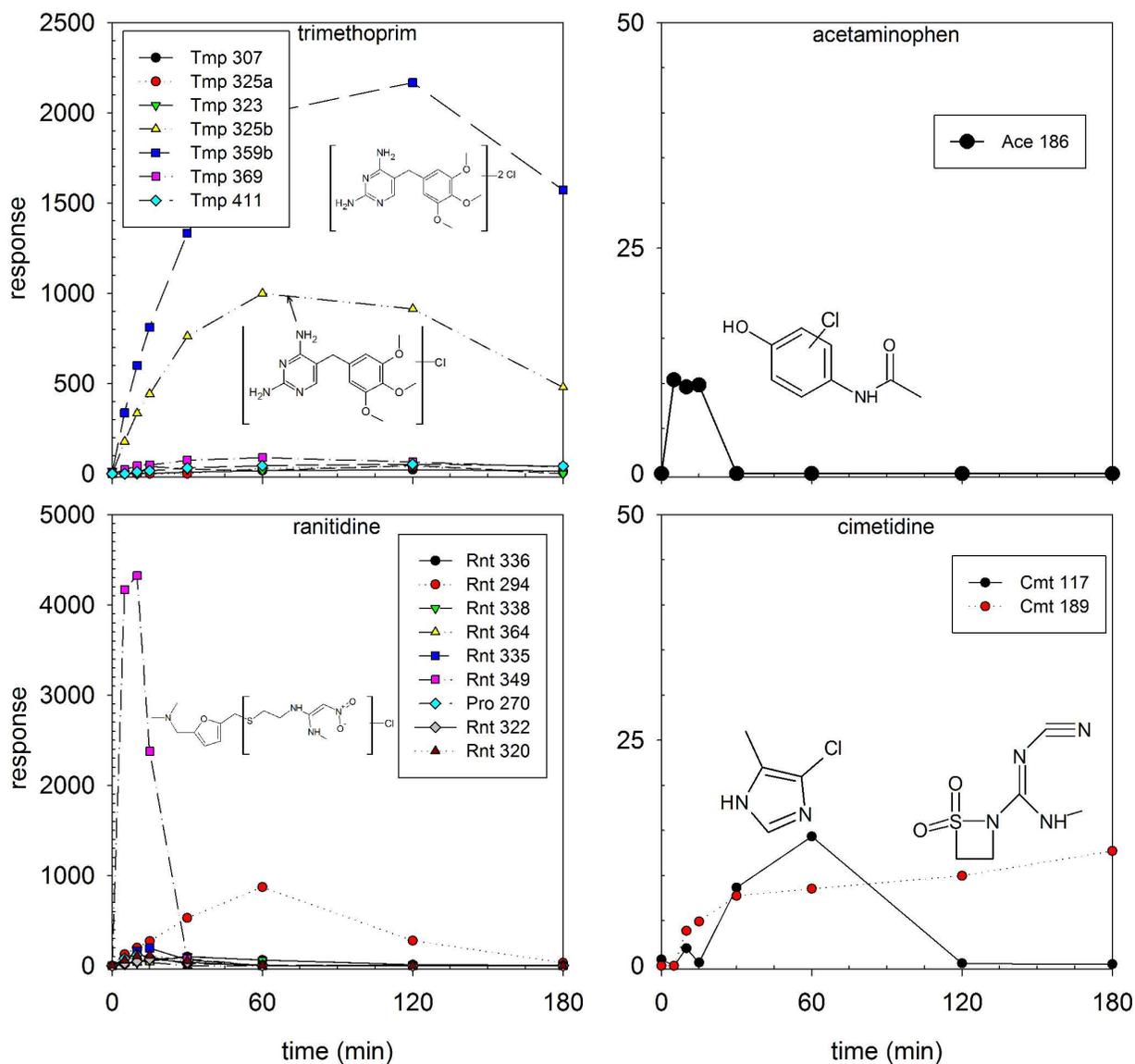
248

249 Figure SI 13b. Test compound transformation product formation during latrine wastewater
250 electrolysis at 3.5 V. Transformation product IDs represent the nominal mass to charge ratio for
251 each compound. Further transformation product information is given in Table SI 2.

4.0 V, 30 mM Cl⁻



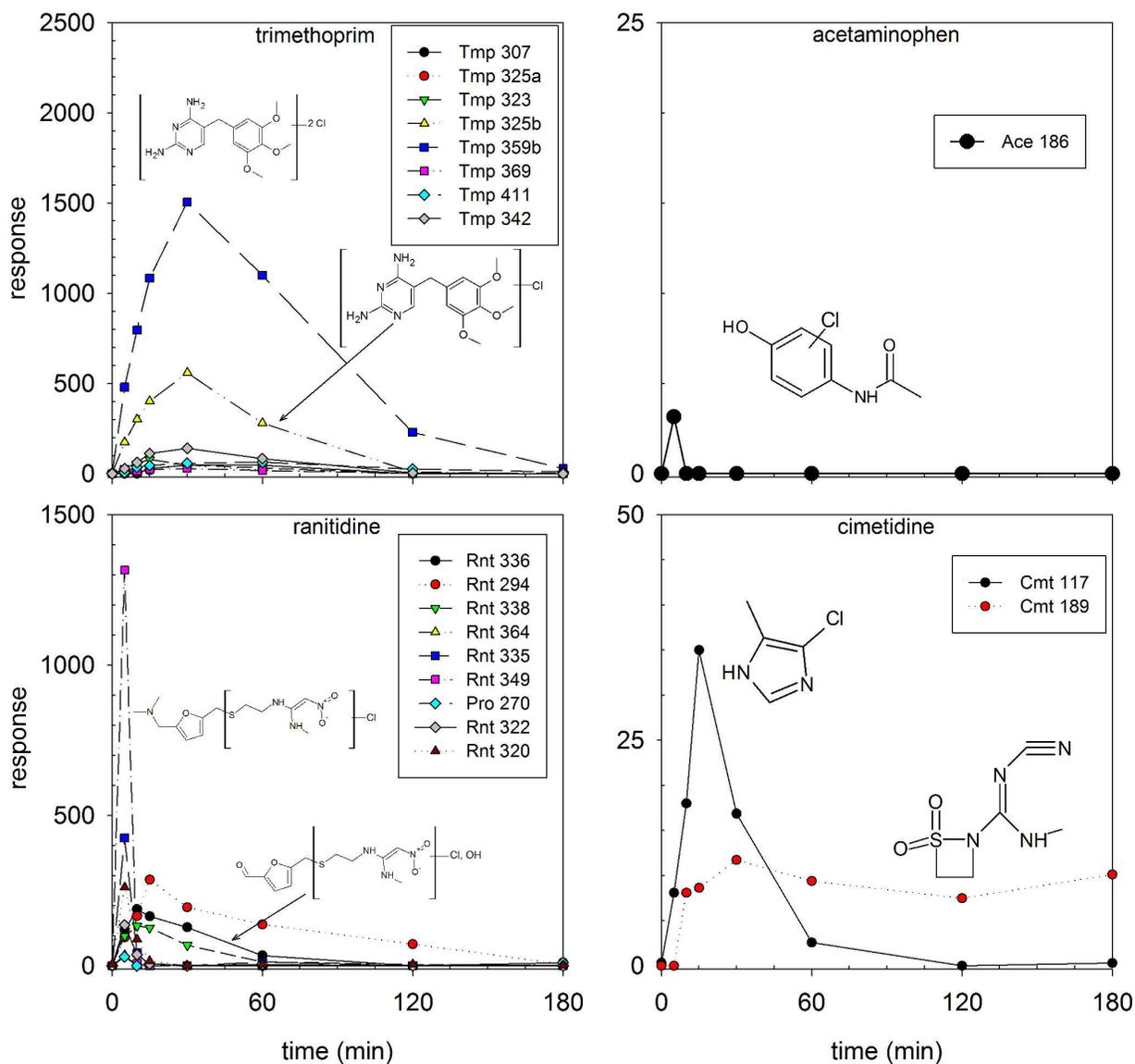
4.0 V, 30 mM Cl⁻



254

255 Figure SI 13c. Test compound transformation product formation during latrine wastewater
 256 electrolysis at 4.0 V. Transformation product IDs represent the nominal mass to charge ratio for
 257 each compound. Further transformation product information is given in Table SI 2.

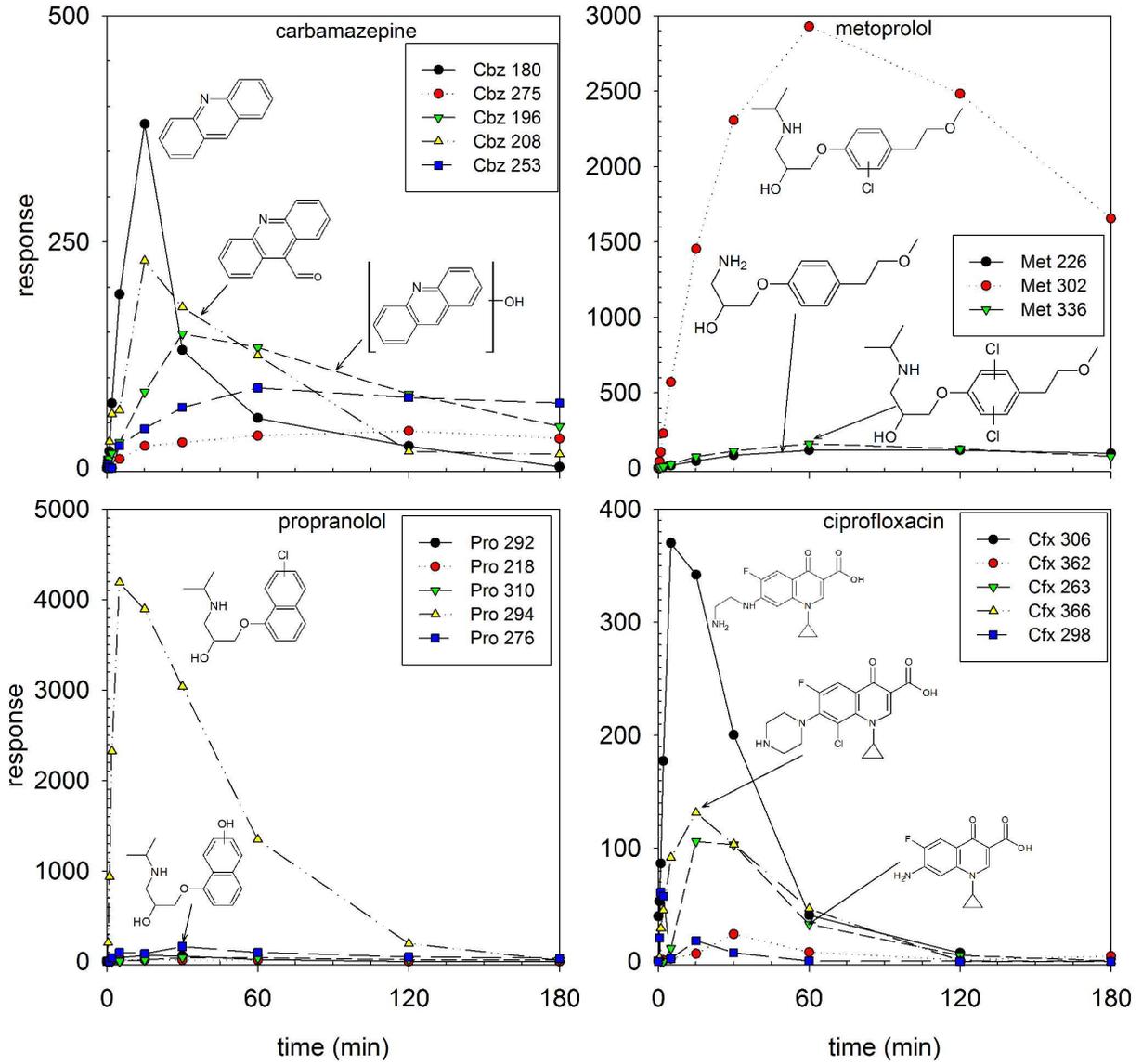
3.5 V, 75 mM Cl⁻



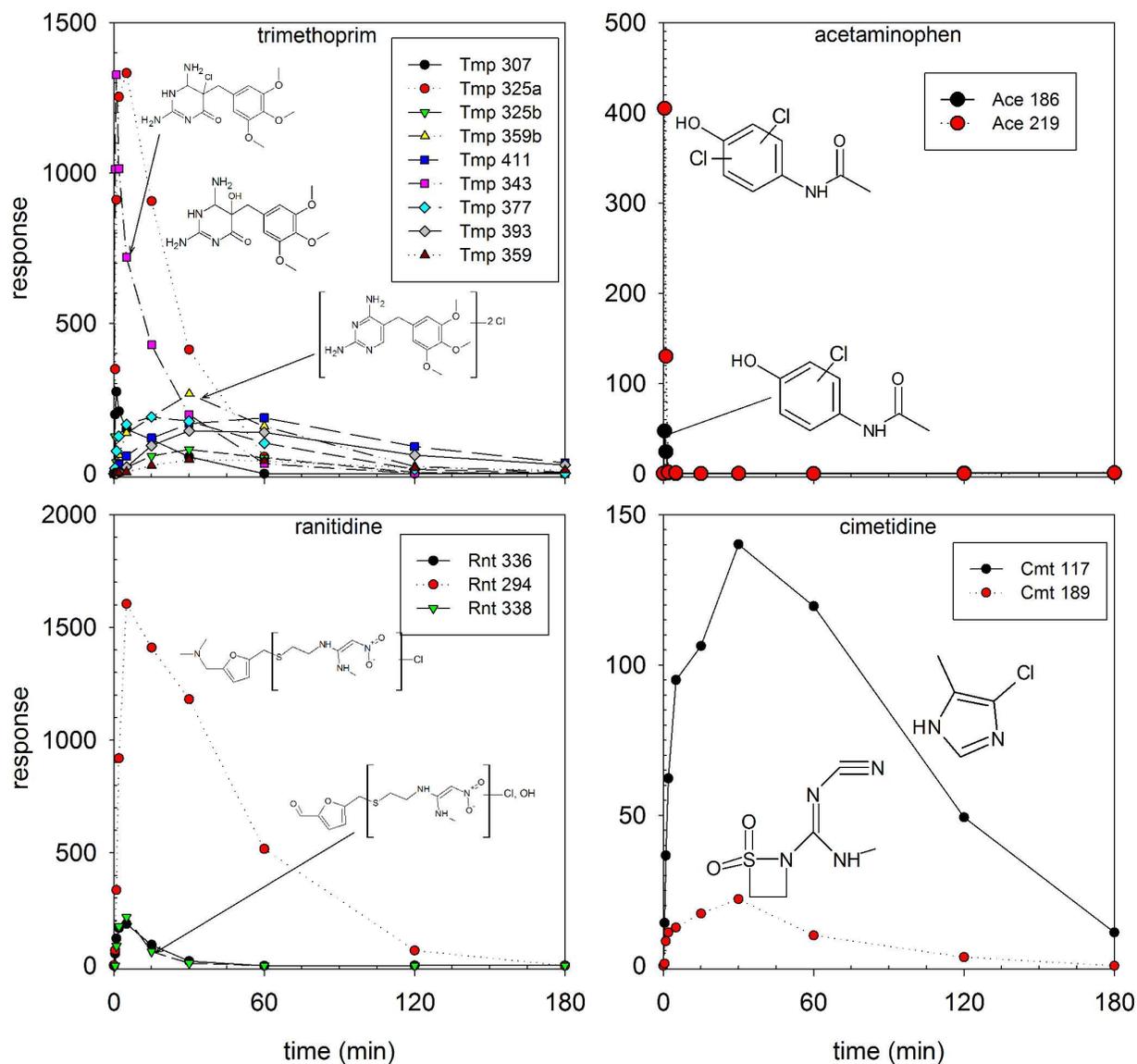
260

261 Figure SI 13d. Test compound transformation product formation during latrine wastewater
 262 electrolysis at 3.5 V with added NaCl. Transformation product IDs represent the nominal mass
 263 to charge ratio for each compound. Further transformation product information is given in Table
 264 SI 2.

Buffered water, 3.5 V, 30 mM Cl⁻



Buffered water, 3.5 V, 30 mM Cl⁻



267
 268 Figure SI 13e. Test compound transformation product formation during buffered Cl⁻ solution
 269 electrolysis. Transformation product IDs represent the nominal mass to charge ratio for each
 270 compound. Further transformation product information is given in Table SI 2.

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