A Redox-Switchable α -Cyclodextrin-Based [2]Rotaxane

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General. All reagents, including α -cyclodextrin, 5-hydroxyisophthalic acid (S2), propargyl bromide, 2-bromoethanol (S7), and propargyl alcohol (S9) were purchased commercially and were used without further purification. The monotosylated tetrathiafulvalene derivative (S1) was prepared according to literature procedures.^{S1} Dimethyl 5-hydroxyisophthalate (S3) was prepared from S2, according to literature procedures.^{S2} 2-Azidoethanol (S8) was prepared from 2bromoethanol (S7) and NaN₃, also according to literature procedures.^{S3} Thin laver chromatography (TLC) was carried out using silica gel 60 F254 (E. Merck). Column chromatography was performed on silica gel 60F (Merck 9385, 0.040-0.063 mm). Deuterated solvents (Cambridge Isotope Laboratories) for nuclear magnetic resonance (NMR) spectroscopic analyses were used as received. NMR spectra were recorded on a Brüker Avance 500 or 600 spectrometer at 25 °C. Chemical shifts were reported in parts per million (ppm) downfield from the Me₄Si resonance which was used as the internal standard when recording ¹H NMR spectra. The addition of K_2CO_3 in the NMR experiments of the [2]rotaxane is to destroy the strong hydrogen bonds between α -cyclodextrin and stoppers, making the movement of α -cyclodextrin doable.^{S4} High-resolution matrix-assisted laser desorption/ionization spectra (HR-MALDI) were measured on an AppliedBiosystems DE-STR MALDI time-of-flight mass spectrometer. The reported molecular mass (m/z) values were the most abundant monoisotopic mass. Electrochemical experiments were carried out at room temperature in argon-purged aqueous solutions, with a Princeton Applied Research 263A Multipurpose instrument interfaced to a PC. Cyclic voltammetry experiments were performed using a glassy carbon working electrode (0.018) cm^2 , Cypress Systems). Its surface was polished routinely with 0.05 μ m alumina-water slurry on a felt surface immediately before use. The counter electrode was a Pt coil and the reference electrode was a standard calomel electrode (SCE). The concentration of the sample and

supporting electrolyte (lithium perchlorate (LiClO₄)) were ca. 1.0×10^{-3} and 0.1 mol L⁻¹, respectively. UV/Vis spectra were recorded at room temperature on a Varian 100 Bio instrument. Circular dichroism (CD) spectra were recorded on a Jasco J-715 Spectropolarimeter at 25 °C. The microcalorimetric titrations were performed by an isothermal titration microcalorimeter (MicroCal Inc., Model No.: VP-ITC) at the atmospheric pressure and 25 °C in potassium carbonate-potassium borate-potassium hydroxide buffer solution (pH 10). In each run, a buffer solution of α -cyclodextrin host in a 0.250 mL syringe was sequentially injected with stirring at 300 rpm into a buffer solution of guest in the sample cell (1.4 mL volume). A control experiment was performed to determine the heat of dilution by injecting a host buffer solution into a pure buffer solution, containing no guest. The dilution enthalpy was subtracted from the apparent enthalpy obtained in each titration run, and the net reaction enthalpy was analyzed by using a single-site binding model. The Origin software (MicroCal) was used to simultaneously determine the binding constant (K_a) and reaction enthalpy (ΔH^0) with the standard derivation on the basis of the scatter of data points from a single titration experiment. Three independent titration experiments were performed to afford self-consistent parameters and give the averaged values.

S4: A solution of **S1** (0.59 g, 1.00 mmol), **S3** (0.21 g, 1.00 mmol), K₂CO₃ (1.00 g, 7.24 mmol), LiBr (10.0 mg, 0.12 mmol), and [18]crown-6 (10.0 mg, 0.04 mmol) in anhydrous MeCN (50 mL) was heated under reflux and an atmosphere of argon for 16 h. After cooling, the reaction mixture was filtered and the solid was washed with Me₂CO. The combined filtrates were concentrated, and the residue was purified by column chromatography (SiO₂: CH₂Cl₂ / EtOH 99:1) to give compound **S4** (0.41 g, 65%) as a yellow oil. ¹H NMR (500 MHz, CD₂Cl₂, 25 °C, TMS): δ 3.58–3.72 (m, 14H, OCH₂), 3.98 (s, 6H, CO₂Me), 4.13–4.15 (m, 2H, Ar-OCH₂), 4.29 (s, 4H,

TTF-CH₂), 6.28–6.29 (d, 2H, TTF-H), 7.87 (s, 2H, Ar-H), 8.32 (s, 1H, Ar-H). ¹³C NMR (125 MHz, CD₂Cl₂, 25 °C): δ 52.3, 62.1, 70.1, 71.2, 79.3, 117.5, 118.6, 121.5, 127.6, 131.1, 138.4, 158.3, 166.6. MS (HR-MALDI): Calcd for C₂₆H₃₂O₁₀S₄ *m*/*z* = 632.0878, found *m*/*z* = 632.0821.

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