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Enantioselective Total Synthesis of Nigelladine A via Late-Stage C–H Oxidation Enabled by an Engineered P450 Enzyme

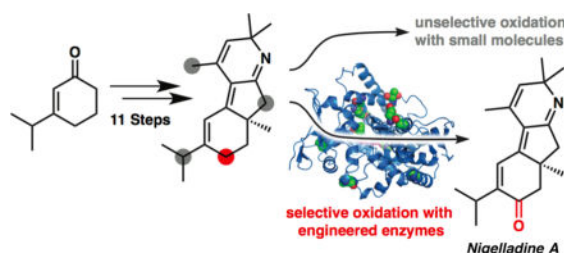
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Abstract

An enantioselective total synthesis of the norditerpenoid alkaloid nigelladine A is described. Strategically, the synthesis relies on a late stage C–H oxidation of an advanced intermediate. While traditional chemical methods failed to deliver the desired outcome, an engineered cytochrome P450 enzyme was employed to effect a chemo- and regioselective allylic C–H oxidation in the presence of four oxidizable positions. The enzyme variant was readily identified from a focused library of three enzymes, allowing for completion of the synthesis without the need for extensive screening.

Graphical abstract



The field of organic synthetic chemistry has benefited greatly from the many recent advances in selective C–H functionalization.¹ Indeed, chemical oxidations of C–H bonds are some of the most important transformations in synthetic chemistry because they allow for direct access to oxidized intermediates, without the need for synthetic handles or functional group interconversions. Despite recent advances, there remain significant limitations regarding the regioselectivity of non-directed C–H oxidation reactions. We envisioned that the recently isolated norditerpenoid alkaloids nigelladines A–C (**1–3**) and pyrroloquinoline alkaloid nigellaquinomine (**4**) (Figure 1)² would present a challenging test bed to evaluate the viability of late-stage C–H oxidation methods.

With respect to nigelladine A, we believed that late-stage installation of the C7 ketone would offer significant flexibility in how we chose to construct the highly conjugated core of

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alkaloid **1**. However, despite the abundance of chemical allylic oxidation conditions, there are relatively few examples of selective oxidation at a 2° carbon (e.g., C7) in the presence of a non-bridging 3° carbon (e.g., C10),³ since oxidation generally occurs at the position where hydrogen abstraction is most favorable.⁴ On the other hand, enzymes are well known to catalyze oxidation reactions with regioselectivity that defies conventional trends of chemical reactivity. Despite this, enzymatic methods are often overlooked in total synthesis efforts due to a typically narrow substrate scope deriving from their exceptional specificity. Fortunately, recent progress in the field of directed evolution has greatly increased the viability of biocatalysis in total synthesis by increasing the reactivity and selectivity of enzymes for non-native transformations.^{5,6} Biocatalytic C–H oxidations with engineered enzymes thus present an alternative approach that can circumvent the limitations of traditional chemical oxidations. Nonetheless, even though there have been examples of the use of engineered enzymes for the functionalization of complex molecules,⁷ their application in total syntheses is still very limited. Herein we report the first enantioselective total synthesis of tricyclic alkaloid **1**, enabled by the selective late-stage allylic oxidation of unsaturated imine **5** *via* an engineered P450 enzyme.

Retrosynthetically, we surmised that selective late-stage allylic oxidation of **5** at C7 would be essential to the synthesis of enone **1**. Imine **5** could be generated by the cross-coupling of vinyl boronic ester **6** and bromide dienone **7**. Vinyl bromide **7** was envisioned to arise via annulation of enone **8**, with the quaternary stereocenter constructed by an asymmetric allylic alkylation reaction from β -ketoester **9**.⁸ With this retrosynthetic analysis, we recognized that the strategically planned C–H oxidation would be particularly challenging due to issues of site selectivity. Imine **5** contains three different allylic carbon centers that could be susceptible to oxidation, a 1°, 2°, and 3°, in addition to a relatively acidic α -carbon adjacent to the imine. Together, the natural product combined with our strategic analysis unveiled a challenging problem for selective C–H functionalization that would need to be addressed in a successful synthesis.

In the forward direction, known enone **10** can be synthesized in excellent yield through a Stork-Danheiser transposition from 1,3-cyclohexanedione.^{9,10} Unfortunately, attempts to directly acylate the α -carbon of **10** resulted in low yields due to competing self-aldol additions. To avoid these issues, we chose to increase the reactivity and steric environment of the enolate by synthesizing the 1,1-dimethyl hydrazone **11**.¹¹ Hydrazone **11** was readily acylated then hydrolyzed, and the resulting β -ketoester was subsequently methylated, providing our desired asymmetric allylic alkylation substrate (**9**) in good yield (73% over 3 steps, Scheme 1). With β -ketoester **9** in hand, we evaluated a variety of reaction parameters for the key quaternary stereocenter-forming asymmetric allylic alkylation and determined that the use of (CF₃)₃-*t*-Bu-PHOX (**L1**)¹² in 2:1 hexanes–toluene with Pd₂(dba)₃ was optimal, generating desired enone **8** in 96% yield and 87% ee. Standard Tsuji–Wacker oxidation of enone **8** using PdCl₂ and CuCl under an atmosphere of oxygen generated the desired ketone,¹³ which subsequently underwent an aldol condensation to provide dienone **12** in good yield (76% yield over 2 steps).¹⁴ Enone **12** was then selectively mono-brominated to form vinyl bromide **7** in 83% yield on gram scale.¹⁵ The cross coupling of **6**¹⁶ and **7** was next evaluated for optimal reaction conditions. Ultimately, we found that

Buchwald's 2nd generation precatalyst with XPhos (XPhos Pd G2),¹⁷ allowed for the union of **6** and **7** in 88% yield using only a slight excess (1.1 equiv) of **6**. Furthermore, triene **13** provided crystals suitable for X-ray diffraction and allowed us to ascertain absolute stereochemistry.¹⁸

With triene **13** in hand, we turned our attention to the synthesis of the heterocyclic portion of the alkaloid and the final stages of the synthesis. Toward that end, triene **13** was treated with trifluoroacetic acid (TFA) in CH₂Cl₂ to deprotect the amine, which spontaneously cyclized to form the requisite imine upon quenching with aqueous sodium bicarbonate (Scheme 2). Fortuitously, upon removal of the aqueous layer, treatment with silica gel promoted isomerization of the exocyclic olefin into the ring, which, after purification, afforded the TFA salt of **5** in 75% yield (25% over 10 steps from enone **10**).¹⁹

With only an allylic oxidation separating tricycle **5** from nigelladine A (**1**), we initiated a broad exploration of oxidation methods for installing an oxidation handle at the desired C7 position. Unfortunately, the required site-selective C–H oxidation of **5** or the intermediate enone **13** proved intractable for traditional synthetic methods (Table 1). While numerous conditions were probed on both substrates,^{20,21} only a limited few effected oxidation at the desired position. These reactions also exhibited poor selectivity, generating an inseparable mixture of mono-oxidation products, along with over-oxidized byproducts that constituted the majority of the product mixture. We thus turned our attention to enzymes as a potential means to achieve the desired oxidation.

The cytochrome P450 enzyme from *Bacillus megaterium* (P450_{BM3}) is a workhorse of protein engineering studies because it is soluble and self-contained, exhibits one of the fastest reaction rates of any P450-catalyzed hydroxylation (17,000 min⁻¹ for arachidonic acid),^{5a} and possesses good stability (*t*_{1/2} = 68 min at 50 °C).²² Since the substrate scope of the wild-type protein is mostly limited to long-chain fatty acids, many engineering efforts have focused on expanding the substrate scope of P450_{BM3} to accept larger and more complex substrates. To this end, various approaches have been applied to develop biocatalysts derived from P450_{BM3} that perform regioselective C–H oxygenation of complex molecules for which the wild-type enzyme exhibits minimal activity.^{6c,23} However, these approaches involved extensive screening endeavors, often requiring high-throughput screening and limiting them to substrates that were readily available.

For biocatalysis to be a viable strategy for the late-stage of a total synthesis, it is desirable to identify a synthetically useful catalyst with minimal screening. Thus, we elected to screen a focused library of variants that had been previously engineered to accept large substrates,²³ but had otherwise never encountered substrate **5**. To compare the catalysts, we chose to allow the reactions to proceed to only low conversion and compare the ratio of desired to undesired oxidation products (Table 2). Encouragingly, wild-type P450_{BM3} exhibited some selectivity for C7, although oxidation of the isopropyl C–H bond was favored 1.2:1 (Table 2, entry 1). We then probed a small selection of engineered variants that all derive from the same parent (12 mutations from P450_{BM3}). These enzymes were previously evolved for regioselective oxidation of methoxymethyl-protected glycosides, but are distinct in that different combinations of active-site residues have been mutated to alanine. The variant **2A1**,

in which leucine residues 75, 181, and 437 are mutated to alanine, exhibited a worse selectivity compared to wild-type, favoring the isopropyl oxidation product, as well as an unidentified oxidation product (Table 2, entry 2). The two other variants, however, exhibited a reversal in selectivity, favoring oxidation at the desired position in excess of 2:1 (Table 2, entries 3–4). The best variant proved to be **8C7**, which is identical to **2A1**, but lacks the L437A mutation. This variant favors oxidation at the desired position in a 2.8:1 ratio (Table 2, entry 4).

Since enzyme active sites are well known to bind substrate enantiomers differently, we wished to test if the stereochemistry of **5** would affect the activity or selectivity of **8C7**. Using the Pd-catalyzed alkylation, we obtained both enantiomers of **5**, which were then enriched to enantiopurity by chiral preparative HPLC. Each enantiomer was then subjected to oxidation by catalyst **8C7**. Interestingly, at both short (15 min) and long (12 h) reaction times, the enantiomers gave nearly identical results, although the *R* enantiomer exhibited a slightly higher initial rate by a factor of 1.3.

Having achieved the desired regioselectivity, we then needed to improve the yield. Furthermore, we wished to supplant the stoichiometric use of NADPH, which is required if oxygen is used as the terminal oxidant. Fortunately, we could use NADPH in catalytic amounts by regenerating it *in situ* with an alcohol dehydrogenase (ADH) and isopropanol as the terminal reductant.²⁴ With these new conditions, we conducted the oxidation on a 160 mg scale, followed by subjecting the crude product mixture to oxidation with Dess-Martin periodinane, thus affording the natural product in 21% yield over both steps (43% yield based on recovered **5**, Scheme 3). The recovered starting material (**5**) was competent in the same two-step procedure with no loss in efficiency (26% yield).²⁵ Under these conditions, we observed up to 1700 turnovers to the C7 mono-oxygenation product.

In summary, we have completed the first total synthesis of nigelladine A in an expedient 12 steps and 5% overall yield (11% yield based on recovered **5**). The asymmetric allylic alkylation allowed for the construction of the quaternary center in high yield and enantioselectivity. Expedient identification of engineered enzymes allowed for a site-selective 2° allylic oxidation without the need for extensive generational screening or reaction optimization. These results demonstrate that enzymatic transformations are capable of defying standard chemical limitations and should be included in the repertoire of reactions that are traditionally considered for the late stages of total syntheses.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Funding Sources

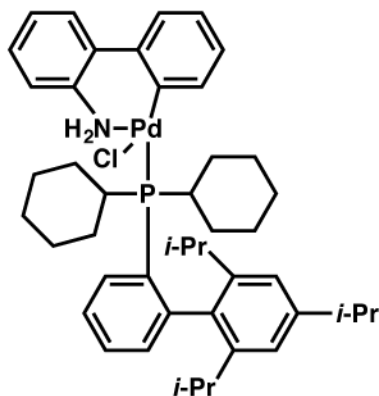
No competing financial interests have been declared.

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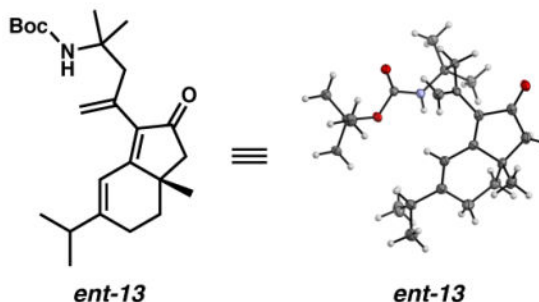
analytical equipment. The authors gratefully acknowledge Larry Henling and Dr. Michael Takase (CIT) for X-ray crystallographic structural determination, Dr. Mona Shahgholi and Naseem Torian (CIT) for mass spectrometry assistance, and Dr. David VanderVelde (CIT) for NMR experimental assistance and helpful discussions. The authors also acknowledge Dr. Sabine Brinkmann-Chen and Dr. Stephan Hammer (CIT) for helpful discussions regarding the enzymatic oxidation system.

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**XPhos Pd G2**

18. Crystals grown for X-ray crystallography were synthesized in the enantiomeric series using (S)-CF₃-tBu-PHOX for the asymmetric allylic alkylation of **9**.



19. The TFA salt of **5** can be deprotonated to the free imine by an aqueous wash with 10% potassium carbonate.
20. See SI for full list of conditions and results.
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25. The TFA salt of **5** was used to avoid deleterious aerobic oxidation of **5**. Both the TFA salt and the free base of **5** were competent substrates and reacted identically for the enzymatic oxidation.

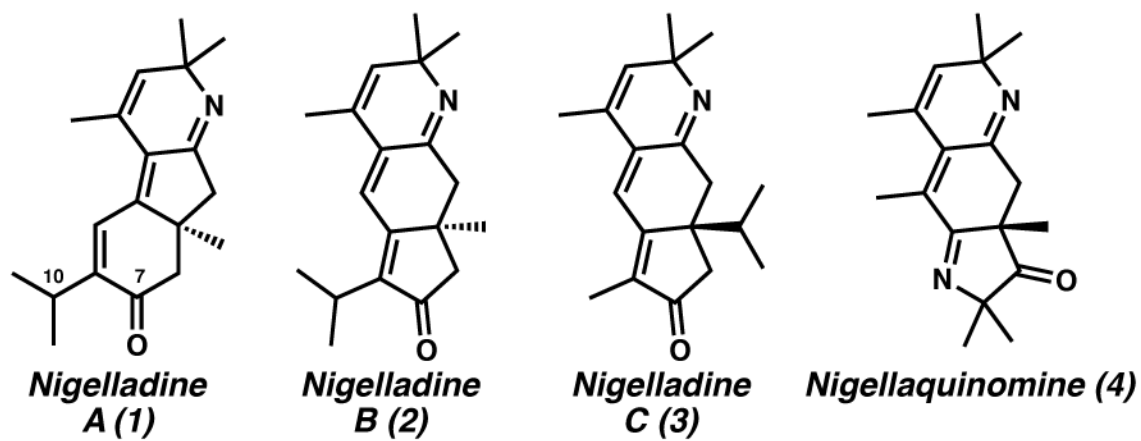


Figure 1.
Structures of nigelladines A–C and nigel-laquinomine.

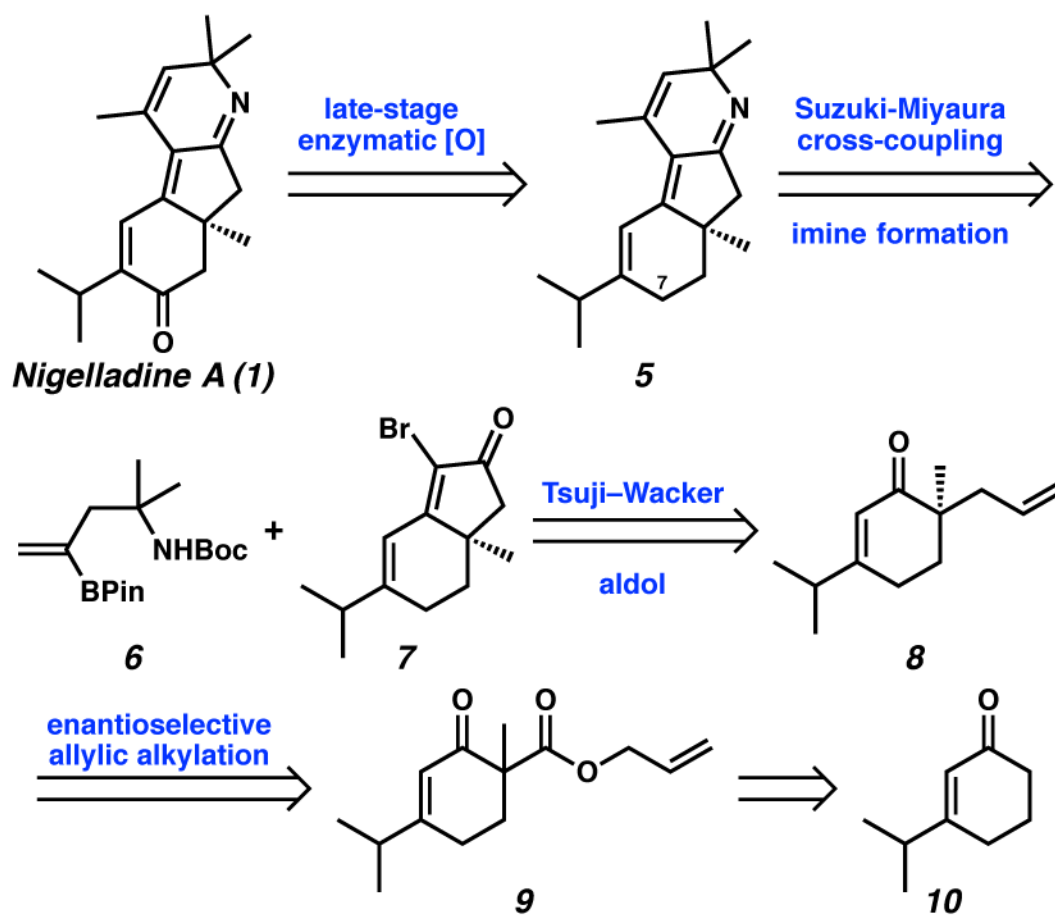
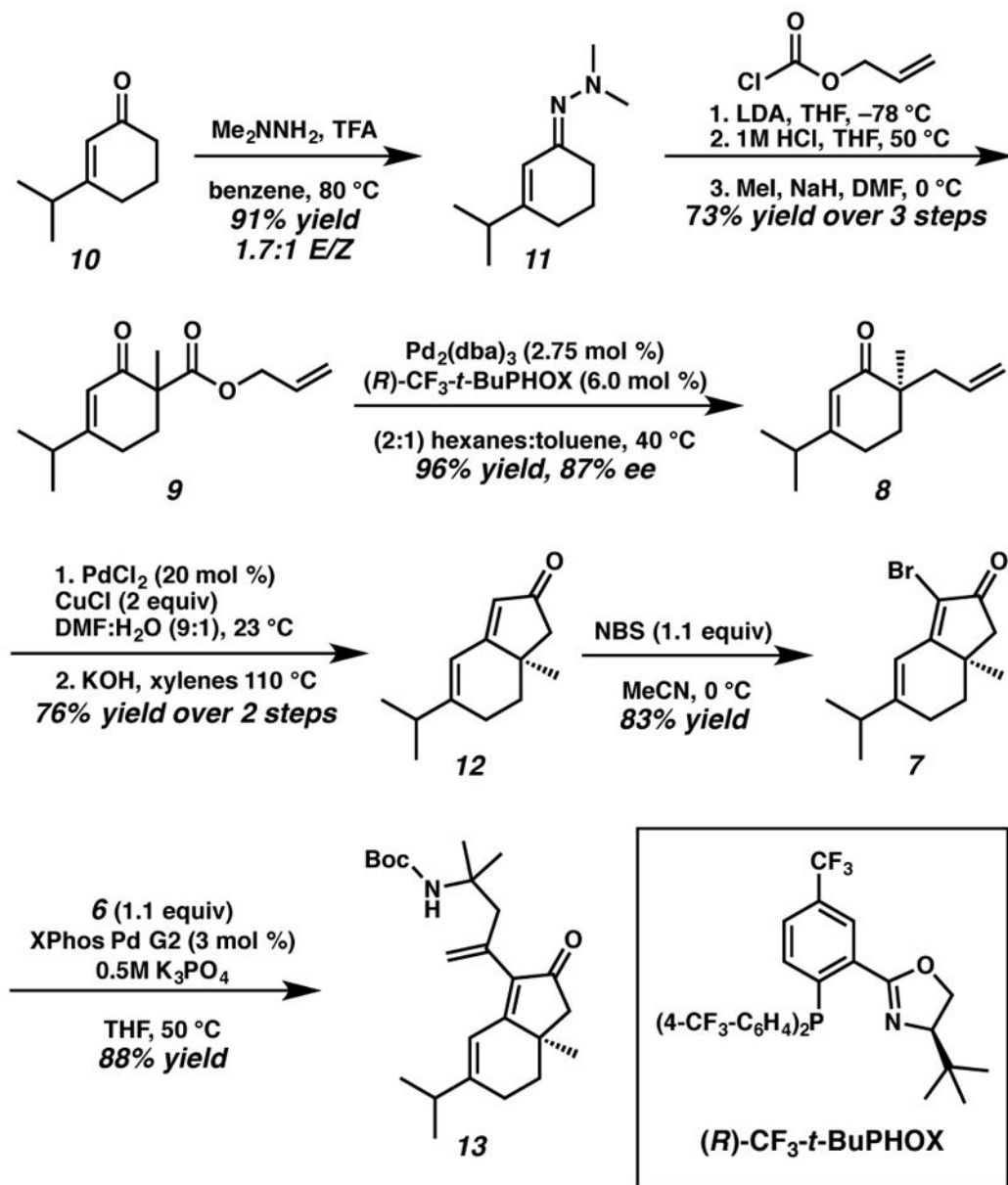
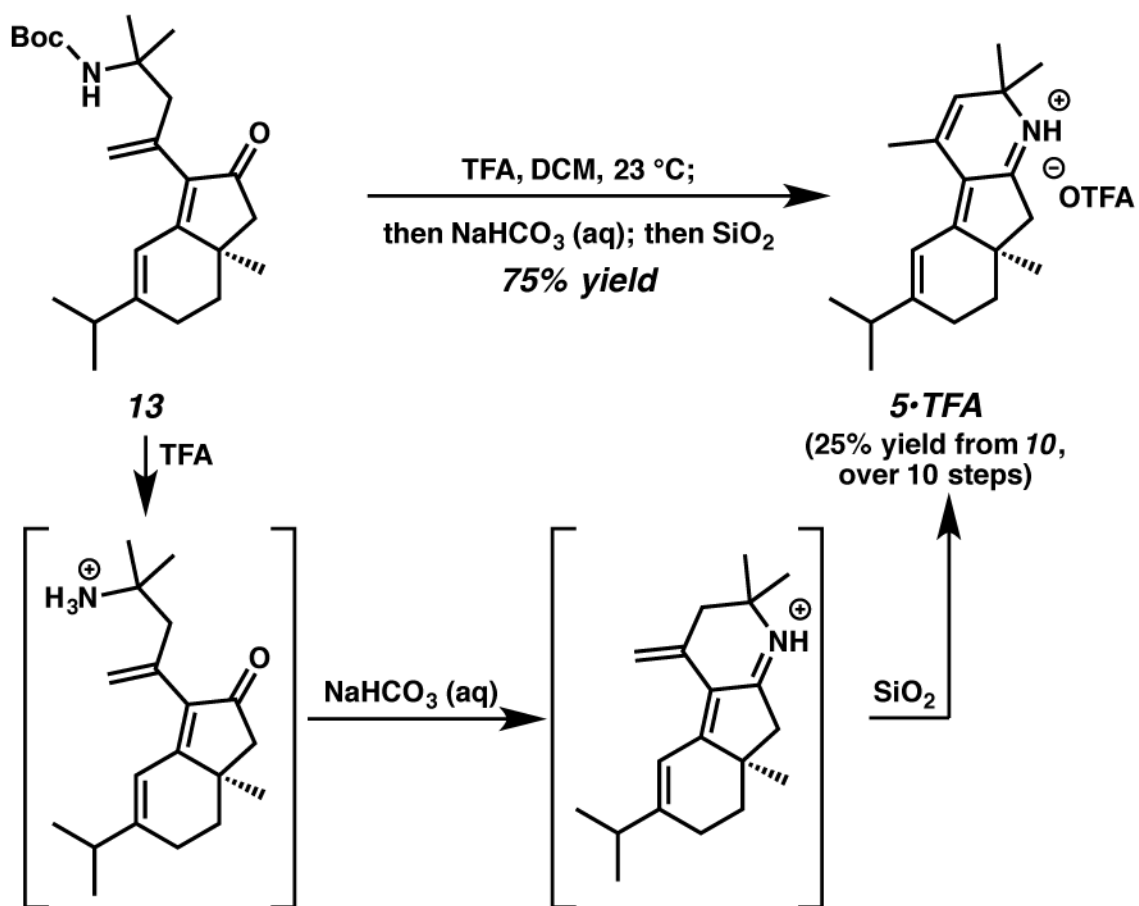


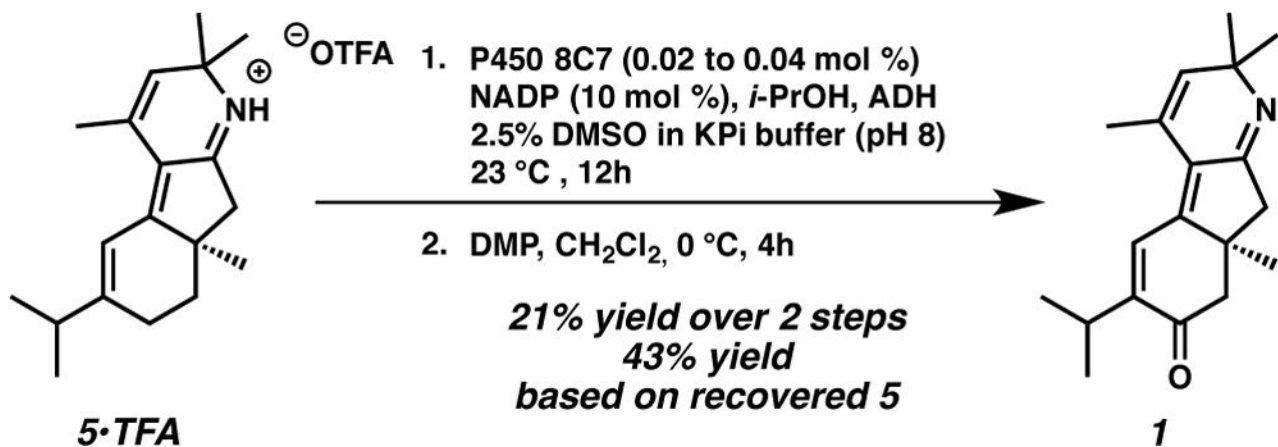
Figure 2.
Retrosynthetic analysis of nigelladine A (1).



Scheme 1.
Synthesis of dienone **13**

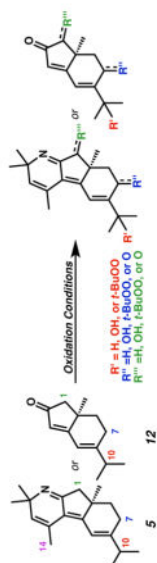


Scheme 2.
Condensation of imine 5

**Scheme 3.**Completion of the synthesis of nigelladine A^a^aADH, alcohol dehydrogenase; KPi, potassium phosphate; DMP, Dess-Martin periodinane.

Attempted chemical oxidations^a

Table 1



Substrate	Conditions	HPLC Yield of Oxygenation Products (%) ^{b,c,d}			
		SM			Polyoxygenation
5	SeO ₂ , DCM, 40 °C	21	79 (57) ^e	–	–
12	SeO ₂ , DCM, 40 °C	5	26 (22)	–	15
5	Pd/C (10 wt%), THPP, 0 °C	7	–	3	44
12	Pd/C (10 wt%), THPP, 0 °C ^f	46	–	2	42
5	Cr(V) (1 equiv), MnO ₂ CF ₃ C ₆ H ₅ , 15-crown-5, 80 °C	55	3	35 ^g	–
5	Rh ₂ (esp) ₂ , T-HYDRO, 23 °C	2	8	21(10)	–
12	Rh ₂ (esp) ₂ , T-HYDRO, 23 °C	–	–	15	16
12	Rh ₂ (esp) ₂ , T-HYDRO, 23 °C	–	–	15	43

^aSee supporting information for reaction conditions and description of the product distribution.

^bApproximated by separating the products on UHPLC-MS and comparing ion count of the various oxidation products.

^cRemaining percent balance remaining were unidentified side products.

^dNumbers given in parentheses indicate isolated yields of the oxidized product.

^eProtection of **5** to as an N-acyl enamide results in oxidation at C-14.

^fShortened reaction time used to observe inherent regioselectivity without over oxidation.

^gOxidized product **S14** was isolated.

Table 2

Screening of focused library for allylic oxidation

Entry	Catalyst ^a	Active-Site Ala Substitutions	Desired/Undesired ^b
1	P450 _{BM3}	–	0.86
2	2A1	L75A, L181A, L437A	0.37
3	4H5	L75A, L177A, L181A	2.1
4	8C7	L75A, L181A	2.8

^aSee supporting information for full list of mutations.^bApproximated by separating the products on UHPLC-MS and comparing ion count of C-7 oxidation product to all other mono-oxygenation products ($m/z = 286$). See supporting information for more details.