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Functional connectivity in the mouse brain imaged by B-mode photoacoustic microscopy

Mohammadreza Nasirivanaki, Wenxin Xing, Jun Xia, and Lihong V. Wang *

^aOptical Imaging Laboratory, Department of Biomedical Engineering, Washington University in St. Louis, St. Louis, MO 63130, USA

* Corresponding author. Email: LHWANG@WUSTL.EDU

ABSTRACT

The increasing use of mouse models for human brain disease studies, coupled with the fact that existing functional imaging modalities cannot be easily applied to mice, presents an emerging need for a new functional imaging modality. Utilizing acoustic-resolution photoacoustic microscopy (AR-PAM), we imaged spontaneous cerebral hemodynamic fluctuations and their associated functional connections in the mouse brain. The images were acquired noninvasively in B-scan mode with a fast frame rate, a large field of view, and a high spatial resolution. At a location relative to the bregma 0, correlations were investigated inter-hemispherically between bilaterally homologous regions, as well as intra-hemispherically within the same functional regions. The functional connectivity in different functional regions was studied. The locations of these regions agreed well with the Paxinos mouse brain atlas. The functional connectivity map obtained in this study can then be used in the investigation of brain disorders such as stroke, Alzheimer's, schizophrenia, multiple sclerosis, autism, and epilepsy. Our experiments show that photoacoustic microscopy is capable to detect connectivities between different functional regions in B-scan mode, promising a powerful functional imaging modality for future brain research.

Keywords: B-scan mode, Resting-state functional connectivity, Functional imaging, Acoustic-resolution photoacoustic microscopy.

1. INTRODUCTION

Functional magnetic resonance imaging (fMRI), the most common neuroimaging modality for human brain imaging, requires a very high magnetic field in order to obtain a sufficient signal to noise ratio (SNR) and spatial resolution for small animal imaging¹. Functional connectivity mapping with optical intrinsic signal imaging (fOIS) was recently introduced as an alternative method to image functional connectivity in mice^{2,3}. In fOIS, changes in local hemoglobin concentrations are determined based on changes in the reflected light intensity from the surface of the brain^{2,4}. Therefore, neuronal activity can be measured through the neurovascular response, similar to the method used in fMRI. However, due to the diffusion of light in tissue, the spatial resolution of fOIS is limited^{2,5}, and the experiment has, thus far, been performed using an exposed skull preparation, which adds complexity for longitudinal imaging. Photoacoustic imaging is an emerging technique that is based on the acoustic detection of optical absorption from tissue chromophores, such as oxy-hemoglobin (HbO₂) and deoxy-hemoglobin (Hb)⁶. This hybrid nature makes photoacoustic tomography capable of providing high resolution images of the brain while leaving the scalp intact⁷. In Figure 1, graphs of molar extinction coefficients of oxy (HbO₂) and deoxy (Hb) hemoglobin for wavelengths ranging from 250 nm to 950 nm are shown.

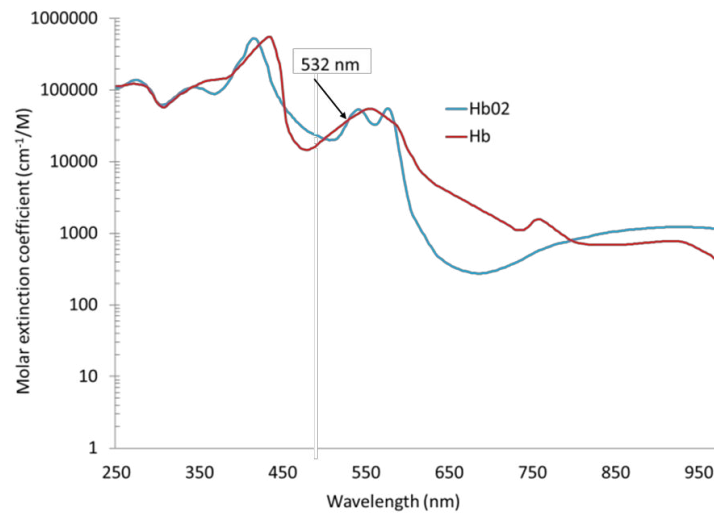


Figure 1. Graph showing molar extinction coefficients of oxy (HbO_2) and deoxy (Hb) hemoglobin. The molar extinction coefficients of HbO_2 and Hb are identical at 532 nm.

Previously⁸, we developed a functional connectivity photoacoustic tomography (fcPAT) system, which, for the first time, allowed noninvasive imaging of resting-state functional connectivity in the mouse brain, with a large field of view and a high spatial resolution. Bilateral correlations were observed in eight functional regions, including the olfactory bulb, limbic, parietal, somatosensory, retrosplenial, visual, motor, and temporal regions, as well as in several subregions.

Here, we image the mouse brain in B-mode, and study the functional connectivity between the homologous functional regions in the left and right hemispheres of the mouse brain, non-invasively. We utilize acoustic-resolution photoacoustic microscopy (AR-PAM) imaging system to image the mouse brain non-invasively. The lateral and axial resolutions are $45\mu\text{m}$ and $30\mu\text{m}$, and the focal zone is around $+0.3\text{mm} \sim -0.7\text{mm}$. By changing the transducer or the numerical aperture, the spatial resolution and the maximum imaging depth of the AR-PAM are saleable within the reach of the excitation photons. An imaging depth of more than 3 mm in live animals has already been demonstrated using our AR-PAM⁹. This is much larger penetration depth compared to other optical imaging modalities such as optical coherence tomography^{10,11}. The acquisition time in this system is 250 ms. A typical image acquired by the AR-PAM from the mouse brain is given in Figure 2.

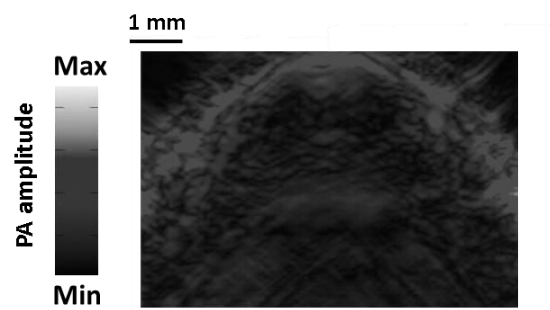


Figure 2. B-scan image of a mouse brain (Bregma 0) acquired non-invasively using the AR-PAM.

For the experiments, we used 3-4 month old Swiss Webster mice. Before imaging, the animal was briefly anesthetized with 2% isoflurane, and the hair was removed with a hair remover lotion. The animal was then mounted on the imaging system. We changed the anesthesia from isoflurane to the mixture of ketamine and xylazine, because they provide stronger brain activity. 100 mg/kg ketamine and 10 mg/kg xylazine were mixed and administered intraperitoneally. For brain imaging, all experimental animal procedures were carried out in conformity with the guidelines of the US National Institutes of Health. The laboratory animal protocols for this work were in accordance with those approved by the Animal Studies Committee of Washington University in St. Louis.

2. SEED-BASED ANALYSIS OF FUNCTIONAL CONNECTIVITY

In each experiment, the animal was imaged using the AR-PAM system for 5 minutes, generating 1200 temporal images. To process the images, the seed-based method is utilized. For the seed-based analysis, the images are processed in order with the following procedures: (a) Smoothing, (b) DC removal, (d) resting-state functional connectivity frequency components (between 0.009 and 0.08 Hz) selection, and (e) global regression. The regression algorithm is performed following the procedure described in [23]. Briefly speaking, for a brain data B including s temporal images and $m \times n$ pixels in each image, the global signal is $B_g = g^{-1} \times B$, where g is a $1 \times s$ vector that contains the average of each brain image, and $g^{-1} = (g \times g')^{-1} \times g'$ (g' is the transpose of g). The global signal is then regressed out from each pixel's temporal signal using $B_p = B - g \times B_g$. In Figure 3, the time traces of seed locations in the left and right secondary motor cortex are shown. As one can see, there is a strong correlation between the signal from these homologous functional regions.

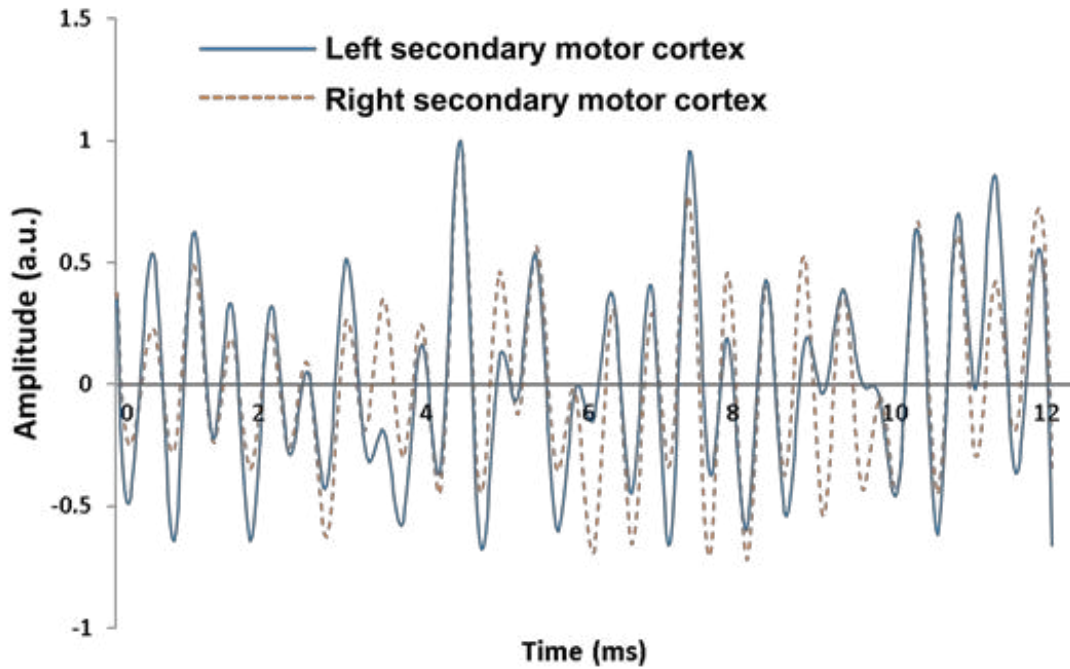


Figure 3. Time traces of seed locations in the left and right secondary motor cortex

3. RESULTS AND DISCUSSION

The mice were imaged by using the AR-PAM imaging system. Images were analyzed using the seed-based method as described in section 3. The imaging depth was 3 mm to 3.5 mm. We co-registered the functional regions on the Paxinos atlas with those on the PAM image. Then, a seed was placed in retrosplenial granular cortex. A strong correlation was observed between the left retrosplenial and right retrosplenial regions (Figure 4). We also observed a strong correlation between the cingulate regions in the left and right hemispheres of the mouse brain. Moreover, simultaneously we observed an anti-correlation between some of the functional regions in left and right hemispheres. We are still investigating the anti-correlated regions. The anti-correlated regions are believed to have opposing functions, but their origins are still being debated. We plan to use electrical stimulation to test the correctness of the results¹².

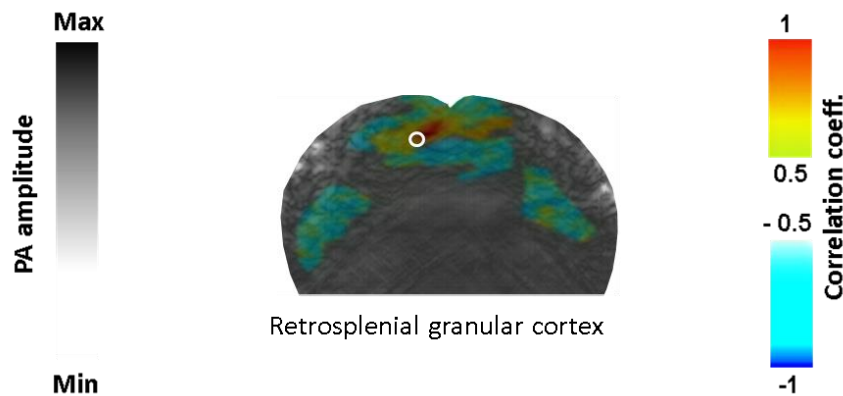


Figure 4. B-mode resting-state functional connectivity maps in a live mouse brain acquired noninvasively by AR-PAM system. Correlation map of retrosplenial granular cortex. White circles: seed regions.

4. CONCLUSION

Utilizing acoustic-resolution photoacoustic microscopy (AR-PAM), we imaged spontaneous cerebral hemodynamic fluctuations and their associated functional connections in the mouse brain. The images were acquired noninvasively in B-scan mode with a fast frame rate, a large field of view. At different locations relative to the bregma, correlations can be investigated inter-hemispherically between bilaterally homologous regions, as well as intra-hemispherically within the same functional regions up to several millimeters in depth. The B-scan functional connectivity maps at different brain locations can be combined to eventually form a three dimensional correlation map of the entire mouse brain. These are in our future plan which is ongoing.

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