

Localized Electronic Structure of Nitrogenase FeMoco Revealed by Selenium K-Edge High Resolution X-ray Absorption Spectroscopy

Justin T. Henthorn,^{‡,†} Renee J. Arias,^{‡,†} Sergey Koroidov,[§] Thomas Kroll,^{||} Dimosthenis Sokaras,^{||} Uwe Bergmann,[§] Douglas C. Rees,^{*,‡,†} and Serena DeBeer^{*,†}

[†]Max Planck Institute for Chemical Energy Conversion, Stiftstr. 34-36, D-45470 Mülheim an der Ruhr, Germany

[‡]Division of Chemistry and Chemical Engineering, California Institute of Technology, Pasadena, California 91125, United States

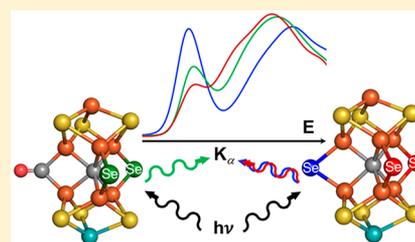
[§]PULSE Institute, SLAC National Accelerator Laboratory, Menlo Park, California 94025, United States

^{||}Stanford Synchrotron Radiation Lightsource, SLAC National Accelerator Laboratory, Menlo Park, California 94025, United States

[†]Howard Hughes Medical Institute, California Institute of Technology, Pasadena, California 91125, United States

Supporting Information

ABSTRACT: The size and complexity of Mo-dependent nitrogenase, a multi-component enzyme capable of reducing dinitrogen to ammonia, have made a detailed understanding of the FeMo cofactor (FeMoco) active site electronic structure an ongoing challenge. Selective substitution of sulfur by selenium in FeMoco affords a unique probe wherein local Fe–Se interactions can be directly interrogated via high-energy resolution fluorescence detected X-ray absorption spectroscopic (HERFD XAS) and extended X-ray absorption fine structure (EXAFS) studies. These studies reveal a significant asymmetry in the electronic distribution of the FeMoco, suggesting a more localized electronic structure picture than is typically assumed for iron–sulfur clusters. Supported by experimental small molecule model data in combination with time dependent density functional theory (TDDFT) calculations, the HERFD XAS data is consistent with an assignment of Fe2/Fe6 as an antiferromagnetically coupled diferric pair. HERFD XAS and EXAFS have also been applied to Se-substituted CO-inhibited MoFe protein, demonstrating the ability of these methods to reveal electronic and structural changes that occur upon substrate binding. These results emphasize the utility of Se HERFD XAS and EXAFS for selectively probing the local electronic and geometric structure of FeMoco.



1. INTRODUCTION

The nitrogenase family of enzymes is responsible for nearly all biological nitrogen fixation and has attracted intense mechanistic scrutiny due to its ability to reduce N_2 to NH_3 at ambient temperature and pressure. This is in sharp contrast to the high temperatures and pressures of the Haber–Bosch process, the industrial analogue accounting for approximately half of global nitrogen fixation.^{1,2} Of the known nitrogenases, molybdenum-dependent nitrogenases show by far the highest activity for N_2 reduction, while vanadium-dependent nitrogenases show relatively high activity for CO reduction.³ Hence, nitrogenases can enable both Haber–Bosch⁴ and Fischer–Tropsch⁵ chemistry with varied efficiencies. Due to the ability of the enzyme to tune the reactivity of the active site to selectively and efficiently carry out these challenging chemical transformations, there is great interest in understanding the atomic-level details of the enzymatic mechanisms.

To date, a multitude of spectroscopic techniques—including electron paramagnetic resonance (EPR),⁶ Mössbauer,⁷ and X-ray spectroscopy^{8–11}—have been used to interrogate the electronic structure of the FeMo cofactor (FeMoco, Figure 1). A major drawback for most of these techniques is their lack of spatial specificity, producing a signal averaged over the entire cluster that can be quite challenging to deconvolute in order to

extract local electronic structure information. These spectroscopies may be further complicated by the presence of the P- and F-clusters, requisite components of nitrogenase under native turnover conditions, further limiting their utility in characterizing intermediates in the reaction cycle. While EPR spectroscopy has proven to be one of the more selective probes in studies of the nitrogenase active site (particularly more reduced states) through effective spin state discrimination, the technique is still generally limited by the signal average problem and the requirement of odd-integer-electron species, potentially excluding a number of intermediates in the catalytic cycle (though under ideal conditions some information can still be extracted from non-Kramers systems).^{12–14} In specific instances, the cluster signal average limit has been overcome through single crystal experiments, as demonstrated by the determination of the geometric orientation of the g-tensor in FeMoco.¹⁵ More recently, a spatially resolved anomalous dispersion (SpReAD) study¹⁶ on single crystals of MoFe protein revealed atomistic electronic structure information, suggesting that Fe1, Fe3, and Fe7 were fully localized Fe^{2+} , while the remainder of the Fe centers in FeMoco were local

Received: July 2, 2019

Published: July 29, 2019

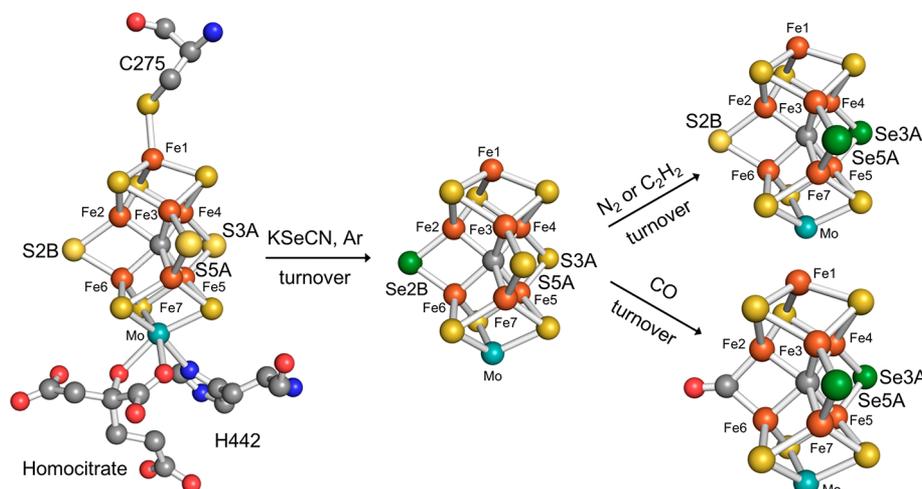


Figure 1. Summary of Se incorporation and migration reactions in FeMoco from *A. vinelandii* (PDB-IDs 3U7Q, 5BVG, 5BVH). Nitrogen atoms are shown in blue, oxygen atoms in red, and carbon atoms in gray.

Fe^{3+} (assuming the $[\text{MoFe}_7\text{S}_9\text{C}]^{1-}$ charge¹⁷ for FeMoco). This proposal of localized Fe oxidation states in FeMoco contrasts with the delocalization typically observed in larger (i.e. $>2\text{Fe}$) FeS clusters.^{18,19} Additionally, since this technique requires single crystals, it is challenging to extend beyond a select set of states. The highly localized electronic structure picture derived from the recent SpReAD study,¹⁶ together with the partially localized electronic structure calculated from recent broken symmetry QM/MM studies,²⁰ have raised questions as to how localized or delocalized the electronic structure is in FeMoco. These questions further motivate the development of selective probes of the local electronic structure of FeMoco that are not simply limited to the resting state, but can also be extended to more reduced states and catalytic intermediates.

It was previously reported that addition of KSeCN to MoFe protein under proton-reducing turnover conditions (argon atmosphere) results in incorporation of Se at the bridging positions of the FeMoco cluster,²¹ with initial selenium substitution occurring at the 2B position (Figure 1). Under turnover in the presence of nonproton substrate (N_2 or acetylene), it was observed that the Se migrates from the 2B position to the 3A and 5A positions. Additionally, it was found that exposure of the Se-substituted cofactor to CO under turnover also results in migration of the Se to the 3A and 5A positions with CO bridging Fe2 and Fe6. The discovery that the belt S atoms of FeMoco can be substituted by Se presents a new avenue for extracting localized electronic structure information from FeMoco, as introducing an element not natively found in the nitrogenase protein generates a selective probe for X-ray spectroscopy. The demonstrated site-selectivity of Se for the belt positions (with no incorporation observed elsewhere in the protein)²¹ allows selective interrogation of the six Fe's bound to the central carbide via the bridging Se atoms. By varying the extent of Se substitution, it is possible to further narrow the information content by deconvoluting the 2B position from the 3A and 5A positions. Additionally, through reaction with CO, it is possible to selectively probe the electronic structure of a substrate bound intermediate. Herein, we demonstrate the utility of selenium substitution in FeMoco as a selective probe of the cluster's local electronic structure.

2. METHODS

2.1. Protein Sample Preparations. Detailed descriptions of the preparation and analysis of the protein samples can be found in the Supporting Information. A succinct description of the four samples investigated can be found below.

Av1Se_{lo} and Av1Se_{hi}. Se-incorporated FeMoco was prepared with slight modifications to methods previously described.^{21,22} Two different Se-substituted FeMoco samples were prepared that differed in the concentration of KSeCN used. Av1Se_{lo} was prepared from a 250 μM KSeCN solution, while Av1Se_{hi} was prepared from a 10 mM KSeCN solution.

Av1SeCO. CO-inhibition of Av1-Se was achieved with methods previously described with some adjustments.^{21–23} A sample of Se-substituted FeMoco was prepared (using 10 mM KSeCN as described above), and after successful selenocyanate-inhibition (as determined by acetylene reduction assays), the system was placed under an atmosphere of CO and allowed to react for 20 min. All further manipulation and purification steps were performed under an atmosphere of CO and using CO-saturated solutions.

Av1Se_{reac}. The protein was substituted with Se (10 mM KSeCN), inhibited with CO as described above, and then reactivated from CO. Reactivation from CO consisted of bubbling a fresh reaction mixture containing all activity components with Ar to replace excess CO. Reactivation from CO of native FeMoco (no Se labeling) has previously been reported,²³ with the reincorporation of S at the 2B position confirmed crystallographically.

Each protein sample was then divided into material for inductively coupled plasma mass spectrometric (ICP-MS), anomalous dispersion, and XAS studies (see Supporting Information for detailed analyses).

2.2. ICP-MS. Protein samples were analyzed for bulk Fe, Mo, and Se content using a quadrupole-based inductively coupled plasma-mass spectrometer (Agilent 8800 ICP-QQQ) at the Environmental Analysis Center at Caltech. Samples were analyzed in helium mode to avoid any argon–argon dimers that may form in the plasma. Calibrations were generated by comparing ion counts from selenium, molybdenum, and iron standards, all prepared in 2% HNO_3 .

2.3. Anomalous Dispersion. Protein single crystal diffraction data were collected at 7110 eV (1.743 Å) and 12 662 eV (0.979 Å) at the Stanford Synchrotron Radiation Lightsource (SSRL) beamline 12–2 equipped with a Dectris Pilatus 6 M detector. Se-anomalous density maps were calculated based on the data collected at 12 662 eV, while the S-anomalous maps were calculated based on the data collected at 7110 eV (see Supporting Information for detailed data processing).

2.4. HERFD XAS and EXAFS Data Collection. Se K-edge HERFD XAS and EXAFS measurements were performed at beamline 6–2 located at the Stanford Synchrotron Radiation Lightsource

facility, operated at 3 GeV with an electron beam current of 500 mA. Beamline 6–2 uses radiation from a 56-pole, 0.9 T wiggler, delivering $\sim 10^{12}$ photons/s (12.5 keV) at the sample position in a spot of $\sim 150 \mu\text{m} \times \sim 400 \mu\text{m}$ (vertical \times horizontal). A rhodium-coated silicon mirror was used to collimate the beam vertically, and a double-crystal monochromator equipped with Si(311) crystals was used to select the energy with a resolution ($\Delta E/E$) of $\sim 10^{-5}$. Energy selection of the emitted X-rays was accomplished using five spherically bent Si(844) crystals aligned on intersecting Rowland circles.²⁴ A helium-filled bag was placed between the sample, the analyzer crystals, and the detector to reduce attenuation of the fluorescence signal. Measurements were performed in a liquid helium flow cryostat maintained at ~ 20 K. Internal energy calibration was achieved by assigning the maximum of gray elemental selenium to 12 659.8 eV.

2.5. HERFD XAS and EXAFS Data Processing. HERFD XAS spectra were background corrected and normalized by fitting the edge-jump to 1. Experimental spectra were fit in an energy range from 12 640 to 12 670 eV as a sum of 1–3 pseudo-Voigt functions in the pre-edge region and 3–5 pure Gaussian functions in the edge region using an iterative least-squares Matlab script. In all cases, the fits converged to pre-edge pseudo-Voigt functions with $\leq 30\%$ Lorentzian composition. All energies and areas reported are from the corresponding fits, with pre-edge areas taken as the sum of the areas of the pre-edge functions multiplied by 100. Edge energies are reported as the white line maxima and pre-edge energies are reported as the maximum of the sum of the pre-edge functions. Deconvolutions were performed using the Se occupancies (Table S1) derived from the anomalous dispersions experiments (see Supporting Information). Se HERFD EXAFS spectra were initially averaged in Matlab 2014b and exported for further processing within Athena.²⁵ A second order polynomial was fit to the pre-edge region and subtracted throughout the entire EXAFS spectrum. A three-region cubic spline (with the AUTOBK function within Athena) was employed to model the background function to a minimum of $k = 12.9 \text{ \AA}^{-1}$ for all spectra. Fourier transforms were performed over a windowed k -range from 2 to 12 \AA^{-1} .

2.6. EPR Spectroscopy. EPR spectra were recorded at the Caltech Electron Paramagnetic Resonance Facility with a Bruker EMX X-band CW-EPR spectrometer with an Oxford ESR 900 liquid helium/nitrogen flow-through cryostat. EPR spectra were recorded at 10K with microwave power 2 mW and microwave frequency 10 GHz.

2.7. Computational Details. All geometry optimizations and TDDFT^{26–28} calculations were executed using ORCA²⁹ version 4.1. Computations were performed using the hybrid TPSSH^{30,31} functional with the D3BJ^{32,33} dispersion correction and CPCM solvation model. The ZORA^{34,35} relativistic approximation was used and employed the relativistically contracted def2 Ahlrichs^{36,37} basis set. A triple- ζ ZORA-def2-TZVP basis set was used for all Se, Fe, S, and N atoms, while a double- ζ def2-SVP basis set was used for all other atoms. The RJCOSX^{38,39} approximation was used to speed up Coulomb and exchange integrals. For the complexes discussed in this work, appropriate antiferromagnetic ground states were achieved starting from a “high spin” ferromagnetic solution and employing spinflip to access the broken symmetry solution.

The hybrid TDDFT calculations were performed using 100–400 roots (depending on the system) to ensure the maximum of the rising edge was calculated. S and Se spectra were plotted with applied broadenings of 1.9 and 2.0 eV (fwhm) and shifted by constant values of +41.2 and -77 eV, respectively. Calculated S and Se XAS spectra were normalized by dividing by 37 and 4.95, respectively, to reproduce pre-edge intensities observed experimentally for a $[\text{Fe}_2\text{S}_2]^{n+}$ series⁴⁰ and $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPH})_4]$ and selenocystine. Sample input files for calculations can be found in the Supporting Information.

3. RESULTS AND DISCUSSION

3.1. S vs Se K-Edge X-ray Absorption Spectroscopies.

Sulfur K-edge XAS has been extensively employed to quantify metal–ligand covalency and probe electronic effects in protein

FeS clusters.^{41–49} S XAS studies on nitrogenase protein have been prohibited by the high number of S atoms present in FeMoco, the P-cluster, F-cluster, as well as the cysteine and methionine residues of the MoFe and Fe proteins, all of which contribute to the total signal.⁵⁰ Additionally, the only existing S XAS study on extracted cofactor is complicated by the presence of dithionite and related decomposition products.⁵¹ While Se-substitution in iron–sulfur clusters is by no means novel, we are unaware of any previous studies wherein Se K-edge XAS was used to probe Se-substituted iron–sulfur clusters.⁵² Though the information content available through Se XAS is analogous to that of S, in practice the achievable energy resolution in conventional Se XAS is much lower due to the significantly shorter 1s core hole lifetime,⁵³ resulting in significant line-broadening in Se K-edge spectra that can lead to a loss of chemical information.

This broadening effect can be observed in Figure 2, where the S and Se K-edge XAS spectra of cystine and selenocystine

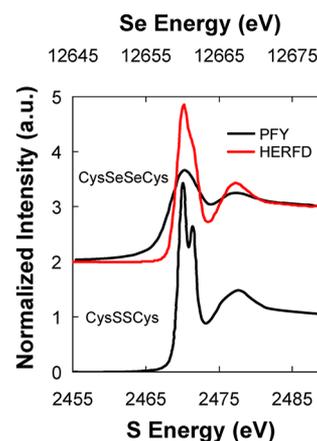


Figure 2. Comparison of S and Se K-edge partial fluorescence yield (PFY) XAS spectra of cystine (bottom) and selenocystine (top, black) with high energy resolution fluorescence detected (HERFD) Se K-edge XAS spectra of selenocystine (top, red). HERFD data are from the present study. PFY data are adapted from ref 54 with permission from the Society of Biological Inorganic Chemistry (1999).

are presented.⁵⁴ The S XAS spectrum of cystine reveals two well-resolved features corresponding to the S 1s to $\text{S}-\text{S}'\sigma^*$ and S 1s to $\text{S}-\text{C}\sigma^*$ transitions at ~ 2470 eV. In the partial fluorescence yield (PFY) Se XAS spectrum of selenocystine, the two expected features are no longer resolvable due to significant line broadening and instead a single unresolved feature is observed. This limitation can be overcome through the use of high energy resolution fluorescence detected (HERFD) XAS.^{9,11,55–57} By selectively measuring the $K_{\alpha 1}$ fluorescence line ($2p_{3/2} \rightarrow 1s$) using high-resolution Bragg optics,²⁴ the effective broadening of the 1s core hole lifetime can be suppressed. Employing this technique, the chemical information lost in the PFY spectrum of selenocystine can be recovered, as demonstrated in the HERFD Se spectrum (Figure 2, red). Using the Se HERFD technique, it is thus possible to acquire chemical information analogous to S K-edge XAS of native protein from Se-substituted FeS protein. Furthermore, selective Se substitution at different bridge positions in FeMoco allows for selective interrogation of a given bridging interaction in FeMoco, free from F- or P-cluster contributions.

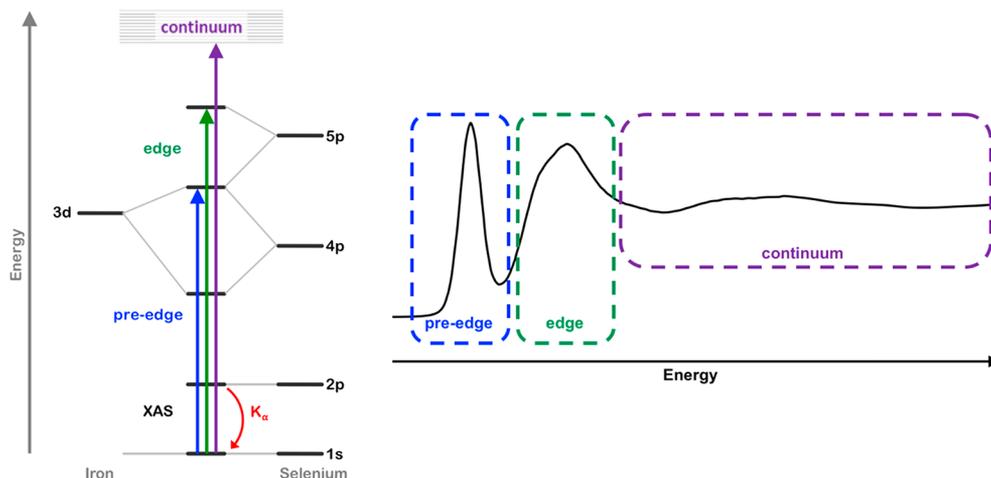


Figure 3. Simplified molecular orbital diagram (left) of Se–Fe interaction showing electronic transitions accessed in Se K-edge XAS (blue, green, and purple), as well as the K_{α} fluorescence measured as part of the HERFD technique (red), and corresponding regions of Se K-edge XAS spectrum (right).

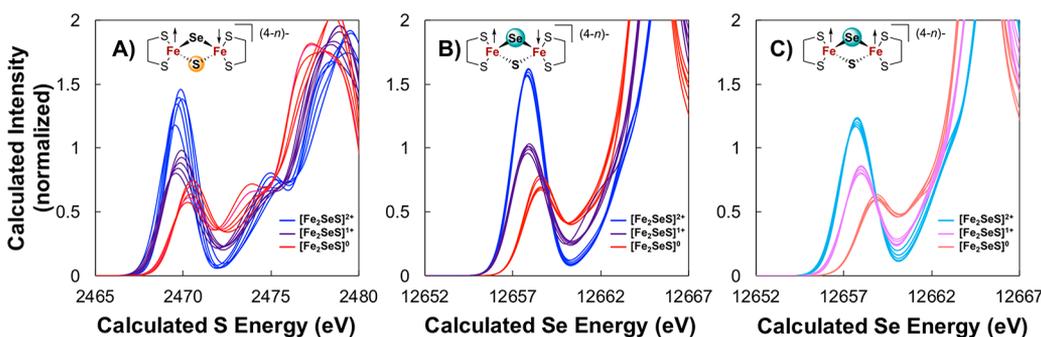


Figure 4. TDDFT calculated S XAS spectra (A) and corresponding Se XAS spectra (B) of a fictitious antiferromagnetically coupled $[\text{Fe}_2\text{SeS}]^{n+}$ series ($n = 2, 1, 0$) as a function of the Fe–Se–Fe angle (from 70° to 90° in intervals of 5°). Se XAS spectra (C) of the analogous ferromagnetically coupled $[\text{Fe}_2\text{SeS}]^{n+}$ series as a function of the Fe–Se–Fe angle (from 70° to 90° in intervals of 5°). Relative coupling is indicated by arrows over the Fe centers. Relevant quantitative data from these calculations are included in Table 1.

Table 1. Calculated and Experimental S and Se XAS Pre-edge Areas and Relative Fe 3d-Manifold Energies

| sample | spin-state | element | calculated pre-edge area (a.u.) | calculated $ \delta_d $ (eV) | experimental pre-edge area (a.u.) | experimental $ \delta_d $ (eV) |
|--|------------|---------|---------------------------------|------------------------------|-----------------------------------|--------------------------------|
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{2-}$ | 0 | S | 280 ± 15^a | 9.50 ± 0.20 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{3-}$ | 1/2 | S | 230 ± 10 | 9.05 ± 0.20 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{4-}$ | 0 | S | 140 ± 15 | 7.65 ± 0.55 | – | – |
| $[\text{Fe}_2\text{S}_2]^{2+ b}$ | 0 | S | 350 | 8.90 | 350 | 7.25 |
| $[\text{Fe}_2\text{S}_2]^{+ b}$ | 1/2 | S | 265 | 8.70 | 260 | 6.68 |
| $[\text{Fe}_2\text{S}_2]^0 b$ | 0 | S | 170 | 6.70 | 180 | 6.27 |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{2-}$ | 0 | Se | 350 ± 5 | 8.00 ± 0.05 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{3-}$ | 1/2 | Se | 275 ± 5 | 7.45 ± 0.05 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{4-}$ | 0 | Se | 150 ± 10 | 6.40 ± 0.05 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{2-}$ | 5 | Se | 285 ± 5 | 7.95 ± 0.10 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{3-}$ | 9/2 | Se | 195 ± 5 | 7.10 ± 0.05 | – | – |
| $[\text{Fe}_2\text{SeS}(\text{S}_2\text{C}_2\text{H}_4)_2]^{4-}$ | 4 | Se | 110 ± 5 | 6.15 ± 0.05 | – | – |
| $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{2-}$ | 0 | Se | 375 | 8.10 | 385 | 6.05 |
| Secystine | 0 | Se | 805 | – | 815 | – |

^aReported values are the mean \pm standard deviation of the 5 geometries calculated (Fe–Se–Fe angle from 70° to 90°). ^bSpectra originally from ref 40; data derived from fittings of experimental spectra and calculations are from present study (Figure S3).

3.2. Se K-Edge HERFD XAS. The Se HERFD XAS spectra presented in this study can be divided into two main regions: (1) the pre-edge ($\sim 12\,655$ – $12\,659$ eV) and (2) the edge ($\sim 12\,662$ – $12\,665$ eV). Pre-edge features arise primarily from a Se 1s to 4p dipole-allowed transition that results from bonding/antibonding interactions of the Se photoabsorber,

while the edge arises primarily from the dipole-allowed Se 1s to 5p transition and reflects the effective nuclear charge of the Se (Figure 3). Beyond the edge, the electron is excited into the continuum where it approximates a free photoelectron and can interact with electrons of neighboring atoms. The postedge region, often referred to as the extended X-ray absorption fine

structure (EXAFS) region, can be used to obtain structural information about the photoabsorber. The studies presented herein will focus on the pre-edge and edge regions, collectively referred to as the HERFD XAS.

In the context of a Se–Fe interaction, as the formal oxidation state of the bound Fe atom increases from Fe²⁺ to Fe³⁺, the corresponding Se XAS pre-edge feature is anticipated to increase in intensity (as additional holes are generated in the Fe 3d-manifold and the Se–Fe interaction becomes more covalent leading to greater orbital overlap), and the edge is anticipated to shift to higher energy (as the effective nuclear charge of Se is increased). The energy of the pre-edge is not as easily predicted a priori, as an increase in the oxidation state of Fe will result in stabilization of both the Fe 3d orbitals and the Se 1s orbital, shifting both the donor and acceptor orbitals lower in energy. However, if the edge energy is used to estimate the continuum, the difference in energy between the edge and pre-edge approximates the relative energy of the Fe 3d-manifold. This difference, which we will refer to as $|\delta_d|$ (and define as $|\delta_d| = |\text{pre-edge energy} - \text{edge energy}|$), is expected to increase with increasing Fe oxidation state. These qualitative trends anticipated for Se HERFD XAS have been quantitatively observed at the S K-edge in a synthetic $[\text{Fe}_2\text{S}_2]^n$ ($n = 2+, 1+, 0$) redox series (Figures S3–S4),⁴⁰ as well as other Cl and S K-edge XAS studies⁴⁷ on synthetic monometallic and dimeric Fe complexes (Figures S5–S7).

These anticipated trends are further supported by time-dependent density functional theory (TDDFT) calculations on a fictitious $[\text{Fe}_2\text{SeS}]^{n+}$ ($n = 0, +1, +2$) series in which analogous S (Figure 4A) and Se K-edge (Figure 4B) XAS spectra are calculated within a single complex. In Figure 4B and Table 1, it can be clearly seen that the Se XAS spectra are sensitive to the oxidation state of the bound Fe's, demonstrating well-separated pre-edge and rising edge features dependent on the oxidation state of the dimer. These calculations indicate that Se is less sensitive to geometric changes compared to S (each line in Figure 4 corresponds to a distinct geometry optimization in which the Fe–Se–Fe angle was fixed, ranging from 70 to 90°, while all other parameters are relaxed), suggesting that electronic rather than geometric effects dominate in Se XAS. This is intuitively consistent, as the larger Se would require a greater geometric distortion to similarly perturb the orbital interaction with Fe relative to the smaller S. In addition, the spectra also appear sensitive to the relative coupling between the bound Fe centers (Figure 4C), with antiferromagnetic coupling resulting in greater pre-edge intensity than the corresponding ferromagnetically coupled system. These results suggest that under the localized limit it may be possible to assign the coupling interaction between bridged Fe centers, absent any significant perturbative interactions (see section 3.7).

Finally, experimental data (Figure 5 and Table 1) on an antiferromagnetically coupled dimer complex $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$ further supports these trends and the direct analogy between Se and S XAS. The Se HERFD XAS spectrum of $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$ reveals an intense pre-edge feature (area = 385) consistent with the previously reported diferric $[\text{Fe}_2\text{S}_2]^{2+}$ complex (S XAS pre-edge area = 350) as well as the TDDFT calculations of our fictitious $[\text{Fe}_2\text{SeS}]^{2+}$ complex (Se XAS pre-edge area 350 ± 5). On the basis of these analyses, the Se K-edge pre-edge intensity and $|\delta_d|$ will be used to evaluate the nature of the Se–Fe interaction and thus reveal local Fe electronic structure information.

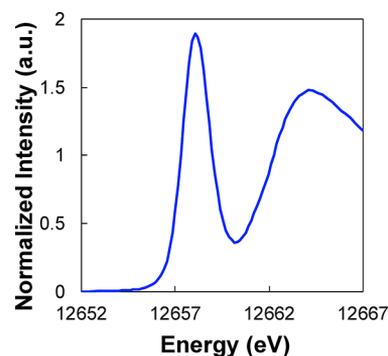


Figure 5. Experimental Se HERFD XAS spectrum of $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$.

3.3. Se Incorporation in FeMoco Samples. Four different protein samples were prepared for the Se HERFD XAS studies with varied Se incorporation in the FeMoco cluster (see Supporting Information for detailed preparations): three samples of Se-incorporated FeMoco, Av1Se_{lo}, Av1Se_{hi}, and Av1Se_{reac} (with the labeling corresponding to how the samples were prepared: low concentration of KSeCN, high concentration of KSeCN, and reactivation after CO-inhibition) and one CO-inhibited Se-incorporated FeMoco sample, Av1SeCO. Each of the four samples were then divided into material for anomalous dispersion, mass spectrometric, and XAS analysis. The Se incorporations, as determined by anomalous dispersion studies (Table S1), indicate that for all four samples the difference in Se occupancy between the 3A and 5A positions was found to be $\leq 5\%$, precluding any meaningful deconvolution of these two positions. Hence herein, 3A and 5A will be treated and discussed together as 3A/5A. Additionally, the anomalous dispersion data indicate Av1Se_{lo} and Av1Se_{hi} exhibit similar Se distributions that are majority 2B, Av1Se_{reac} is more evenly distributed between 2B:3A/5A, and Av1SeCO is majority 3A/5A.

Se incorporation in the protein samples was also quantified by inductively coupled plasma mass spectrometric (ICP-MS) studies in an effort to correlate the single crystal occupancies to the bulk sample (Table S1). Overall, the two measurements were either consistent within error or revealed lower Se:FeMoco ratios in the bulk than was found in the anomalous dispersion experiment of the single crystals. This discrepancy is attributed to the variation observed in f' at 12 662 eV (see Supporting Information), which can affect the absolute Se occupancy values, but will have minimal effect on the relative 2B:3A/5A ratios. We conclude from these studies that samples contain negligible Se outside of the crystallographically observed FeMoco incorporation, and thus the Se HERFD XAS measurements are selectively probing the FeMoco core.

Additionally, EPR spectroscopy was employed to assess the relative perturbation of the Se substitution on the overall electronic structure of the FeMoco, as it has been previously observed that complete Se substitution in $[\text{Fe}_4\text{S}_4]^+$ cubanes can result in stabilization of alternate spin ground states ($S > 1/2$).^{58,59} The EPR spectrum of Se-substituted FeMoco (Figure S2) reveals a broadened $S = 3/2$ signal qualitatively similar to native FeMoco (ref 22), suggesting minimal perturbation of the FeMoco electronic structure upon Se substitution. The lack of other observable spin states by EPR, coupled with the previous finding that Se-substituted FeMoco demonstrates similar acetylene and N₂ reduction activity compared to native

protein,²¹ suggests Se to be a relatively innocent probe of the FeMoco electronic structure. Additionally, the EPR spectrum of CO-inhibited Se-substituted FeMoco confirms the near-complete loss of the $S = 3/2$ resting state signal (consistent with the low occupancy of Se2B as determined by the anomalous dispersion experiments) and reveals a new $S = 1/2$ signal consistent with the lo-CO species previously identified in the literature.⁶⁰

3.4. FeMoco Se HERFD XAS. The K-edge Se HERFD XAS spectra for the four protein samples examined in this study are shown in Figure 6 and the relevant quantitative

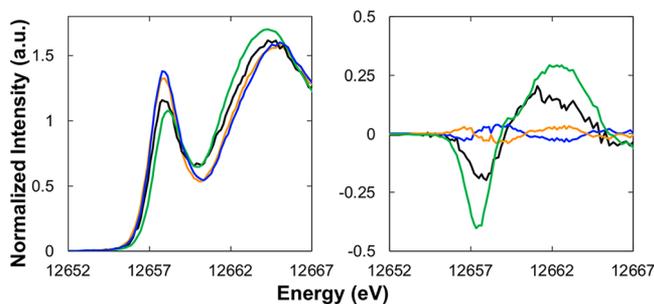


Figure 6. Experimental Se HERFD XAS (left) and corresponding difference spectra (right) of Av1Se₁₀ (blue), Av1Se_{hi} (orange), Av1Se_{reac} (black), and Av1SeCO (green). The difference spectra are taken relative to the average of Av1Se₁₀ and Av1Se_{hi}.

metrics are presented in Table 2. Comparing first the non-CO samples (Av1Se₁₀, Av1Se_{hi}, and Av1Se_{reac}), it is immediately apparent that the spectra vary depending on the Se2B:Se3A/5A ratios. The spectra of Av1Se₁₀ and Av1Se_{hi} (blue and orange, Figure 6) most closely overlap, consistent with the more similar Se occupancies observed in the two samples as determined from the anomalous dispersion studies, while the spectrum of Av1Se_{reac} (black, Figure 6) exhibits a less intense pre-edge feature and an edge shifted to lower energy. These experimental results thus suggest that the Se2B and Se3A/5A positions in FeMoco are electronically distinct. This becomes more obvious when looking at the difference spectra (Figure 6, right). As Av1Se₁₀ and Av1Se_{hi} demonstrate near-identical relative Se occupancies as determined by anomalous dispersion, the difference spectra are given relative to the average of Av1Se₁₀ and Av1Se_{hi}. Thus, the difference spectra for Av1Se₁₀ and Av1Se_{hi} are mirror images and represent an estimate of the absolute error over both the anomalous dispersion and Se HERFD XAS measurements, while the difference spectra for Av1Se_{reac} and Av1SeCO reveal the significant electronic changes observed by increasing the

relative amount of Se3A/5A in these samples compared to Av1Se_{10/hi}.

From the HERFD XAS of these three samples (Av1Se₁₀, Av1Se_{hi}, and Av1Se_{reac}), we can observe that as the fraction of Se2B decreases going from Av1Se₁₀ and Av1Se_{hi} to Av1Se_{reac} (0.74 to 0.53), the pre-edge intensity similarly decreases, with Av1Se₁₀ ≥ Av1Se_{hi} > Av1Se_{reac}. Correspondingly, as the fraction of Se2B decreases from Av1Se₁₀ and Av1Se_{hi} to Av1Se_{reac}, $|\delta_d|$ also decreases, with Av1Se_{hi} ≥ Av1Se₁₀ > Av1Se_{reac}. Together these two trends suggest that Se2B engages in a more covalent interactions with Fe2/Fe6 compared to the average of the Se3A/5A interactions with Fe4/Fe5/Fe3/Fe7, resulting in a more intense pre-edge feature and greater stabilization of the Fe 3d-manifold. Comparing next Av1Se_{reac} to Av1SeCO (Figure 6, green), the two spectra exhibit similar pre-edge areas (285 vs 260) and $|\delta_d|$ values (6.33 vs 6.15), though Av1SeCO contains a significantly higher fraction of Se3A/5A contributing to the spectrum (0.88 in Av1SeCO vs 0.48 in Av1Se_{reac}). This observation suggests not only that the Se2B and the Se3A/5A are electronically distinct, but also that the Se3A/5A position is electronically perturbed upon CO binding to the cofactor.

Using the Se-occupancy populations derived from the anomalous dispersion studies, the pure Se_{2B} spectrum could be deconvoluted from Se_{3A/5A} (Figure 7, blue and red spectra).

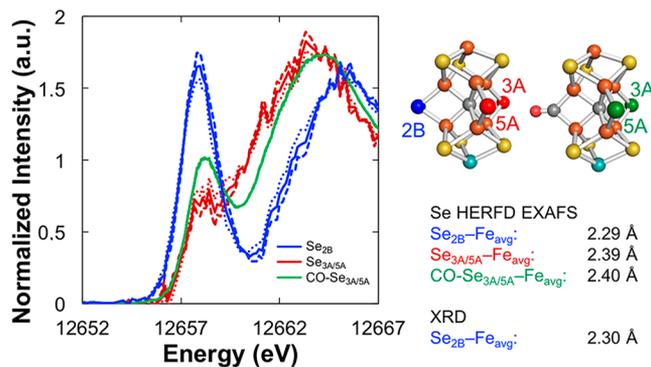


Figure 7. Deconvoluted experimental HERFD Se K-edge XAS spectra (left) of Se_{2B} (blue), Se_{3A/5A} (red), and CO-Se_{3A/5A} (green). The dashed lines are deconvolutions using Av1Se₁₀/Av1Se_{reac}, the dotted lines using Av1Se_{hi}/Av1Se_{reac} and the solid lines the average of the two. Corresponding average Se–Fe distances extracted from Se HERFD EXAFS data are also presented (right).

From the three Se-incorporated MoFe samples investigated (Av1Se₁₀, Av1Se_{hi}, and Av1Se_{reac}), two sets of deconvolutions could be obtained which differ only by minor (<5%)

Table 2. Se HERFD XAS Edge Energies, Pre-edge Energies and Areas, and Relative Fe 3d-Manifold Energies

| sample | edge | | pre-edge | | $ \delta_d $ (eV) |
|--|------------------|------------------|----------|--|-------------------|
| | energy (eV) | energy (eV) | area | | |
| [Et ₄ N] ₂ [Fe ₂ Se ₂ (SPh) ₄] | 12 664.10 | 12 658.05 | 370 | | 6.05 |
| Av1Se ₁₀ | 12 664.80 | 12 657.98 | 330 | | 6.82 |
| Av1Se _{hi} | 12 664.80 | 12 657.92 | 315 | | 6.88 |
| Av1Se _{reac} | 12 664.30 | 12 657.97 | 285 | | 6.33 |
| Av1SeCO | 12 664.30 | 12 658.15 | 260 | | 6.15 |
| Se _{2B} | 12 665.23 ± 0.06 | 12 657.86 ± 0.06 | 385 ± 20 | | 7.38 ± 0.12 |
| Se _{3A/5A} | 12 663.78 ± 0.40 | 12 658.02 ± 0.36 | 130 ± 30 | | 5.76 ± 0.04 |
| CO-Se _{3A/5A} | 12 664.13 ± 0.10 | 12 658.18 ± 0.04 | 245 ± 10 | | 5.95 ± 0.08 |

contributions in 3A or 5A. These deconvolutions were performed by treating the experimental spectra as the sum of the pure $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$ spectra weighted by the relative Se populations at the 2B and 3A/5A positions. Thus, the deconvolutions can be expressed by the following two systems of equations:

$$\begin{bmatrix} \text{Av1Se}_{\text{lo}} \chi_{\text{Se}_{2\text{B}}} & \text{Av1Se}_{\text{lo}} \chi_{\text{Se}_{3\text{A}/5\text{A}}} \\ \text{Av1Se}_{\text{reac}} \chi_{\text{Se}_{2\text{B}}} & \text{Av1Se}_{\text{reac}} \chi_{\text{Se}_{3\text{A}/5\text{A}}} \end{bmatrix} \begin{bmatrix} \text{Se}_{2\text{B}} \\ \text{Se}_{3\text{A}/5\text{A}} \end{bmatrix} = \begin{bmatrix} \text{Av1Se}_{\text{lo}} \\ \text{Av1Se}_{\text{reac}} \end{bmatrix}$$

$$\begin{bmatrix} \text{Av1Se}_{\text{hi}} \chi_{\text{Se}_{2\text{B}}} & \text{Av1Se}_{\text{hi}} \chi_{\text{Se}_{3\text{A}/5\text{A}}} \\ \text{Av1Se}_{\text{reac}} \chi_{\text{Se}_{2\text{B}}} & \text{Av1Se}_{\text{reac}} \chi_{\text{Se}_{3\text{A}/5\text{A}}} \end{bmatrix} \begin{bmatrix} \text{Se}_{2\text{B}} \\ \text{Se}_{3\text{A}/5\text{A}} \end{bmatrix} = \begin{bmatrix} \text{Av1Se}_{\text{hi}} \\ \text{Av1Se}_{\text{reac}} \end{bmatrix}$$

where $\text{Av1Se}_{\text{lo}} \chi_{\text{Se}_{2\text{B}}}$ corresponds to the mole fraction of $\text{Se}_{2\text{B}}$ in sample Av1Se_{lo} , $\text{Se}_{2\text{B}}$ is the pure spectrum of $\text{Se}_{2\text{B}}$, and Av1Se_{lo} is the experimental spectrum of sample Av1Se_{lo} . Solving these two systems of equations results in two sets of pure $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$ spectra, represented as dashed and dotted lines, respectively, in Figure 7, and the average given as a solid line. The two sets of deconvoluted spectra are qualitatively identical and represent an estimate of the combined error over both the Se quantification and HERFD XAS measurements. As such, the data presented in Table 2 correspond to the average of the two deconvolutions mentioned above, with the difference between the two deconvolutions reported as the error. Using the deconvoluted $\text{Se}_{2\text{B}}$ spectra, the 11% occupancy of $\text{Se}_{2\text{B}}$ present in Av1SeCO was subtracted to afford the pure $\text{Se}_{3\text{A}/5\text{A}}$ spectrum of CO-bound FeMoco , CO- $\text{Se}_{3\text{A}/5\text{A}}$ (Figure 7, green spectra). From these deconvolutions, the electronic differences between the $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$ positions, as well as the differences observed at $\text{Se}_{3\text{A}/5\text{A}}$ upon CO binding, can be more clearly visualized and thoroughly analyzed.

As can be seen in Table 2 and Figure 7, the deconvoluted spectra of $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$ are dramatically different, with $\text{Se}_{2\text{B}}$ exhibiting a very intense pre-edge feature at 12 657.9 eV, while $\text{Se}_{3\text{A}/5\text{A}}$ exhibits a significantly less intense pre-edge only one-third the area of the $\text{Se}_{2\text{B}}$ feature (130 ± 30 vs 385 ± 20). This large difference in pre-edge intensity is matched by an equally large shift in the edges of the two spectra, with the $\text{Se}_{3\text{A}/5\text{A}}$ edge shifted 1.5 eV to lower energy. The spectrum of CO- $\text{Se}_{3\text{A}/5\text{A}}$ reveals a pre-edge feature intermediate in intensity between $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$, as well as an edge similarly intermediate between $\text{Se}_{2\text{B}}$ and $\text{Se}_{3\text{A}/5\text{A}}$. Qualitatively, these differences again suggest that $\text{Se}_{2\text{B}}$ engages in more covalent bonding to Fe than $\text{Se}_{3\text{A}/5\text{A}}$ and that the $\text{Se}_{3\text{A}/5\text{A}}$ position is electronically sensitive to CO binding at the distal edge of the cluster.

3.5. FeMoco Se HERFD EXAFS. The Se-substituted FeMoco samples were also analyzed by Se HERFD EXAFS to complement the local electronic structure information obtained from the Se HERFD XAS experiments with local structural information (Figure 8). It can be immediately observed that the first Se–Fe radial shell (radial distance ~ 1.9 Å, non-phase-corrected) shifts to higher values going from Av1Se_{lo} and Av1Se_{hi} to $\text{Av1Se}_{\text{reac}}$ and Av1SeCO , suggesting the samples with greater $\text{Se}_{3\text{A}/5\text{A}}$ content exhibit longer average Se–Fe distances. The EXAFS data were fit with multiple single-scattering paths including the first Se–Fe radial shell, a Se–S radial shell, and a second Se–Fe' radial shell (Figure S47–50). While fits omitting the long-range Se–S and Se–Fe' scattering paths (Figures S51–54) afforded similar first

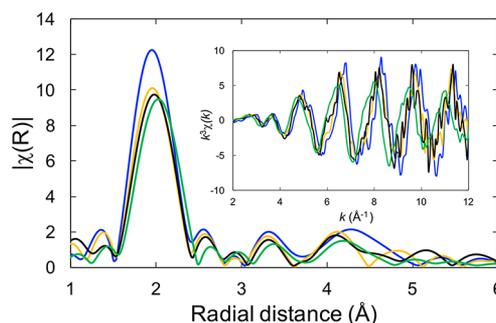


Figure 8. Non-phase-shifted Fourier transform of k^3 -weighted Se HERFD EXAFS (with k^3 -EXAFS inset) of Av1Se_{lo} (blue), Av1Se_{hi} (orange), $\text{Av1Se}_{\text{reac}}$ (black), and Av1SeCO (green).

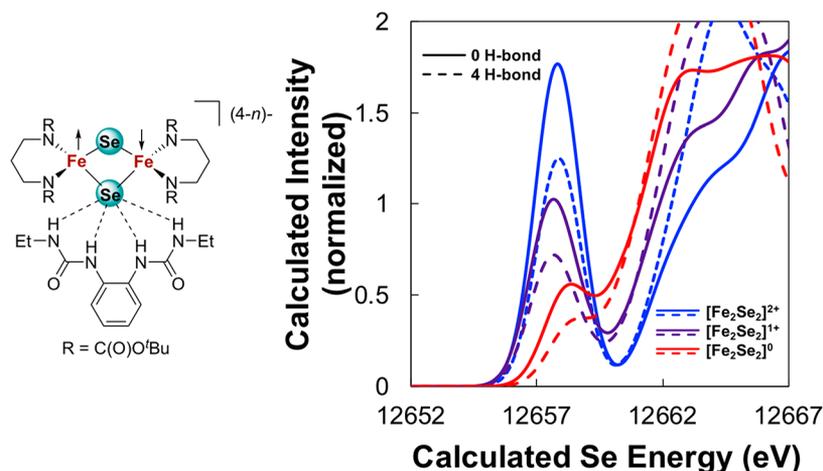
shell Se–Fe distances (± 0.007 Å), their inclusion resulted in improved fits with increased precision and decreased Debye–Waller-like disorder parameters (σ^2) for the first Se–Fe radial shell (Table S6). As shown in Table 3, the fit Se HERFD EXAFS data quantitatively reveal the clear trend observed in the first Se–Fe scattering path, with the average Se–Fe radial shell increasing from 2.314 Å for Av1Se_{lo} to 2.388 Å for Av1SeCO . This observation is again consistent with $\text{Se}_{2\text{B}}$ engaging in a more covalent interaction with Fe compared to $\text{Se}_{3\text{A}/5\text{A}}$.

Similar to the deconvolution of the Se HERFD XAS spectra, the average Se–Fe distances of the 2B and 3A/5A positions were deconvoluted by solving the same systems of equations with the EXAFS-fitted average Se–Fe distance used in place of the Se HERFD XAS spectra (Table 3). Again, the two sets of solutions were averaged and the difference reported as the error. The resultant values reveal a short average Se–Fe distance of 2.29 ± 0.02 Å for $\text{Se}_{2\text{B}}$ and a significantly longer average Se–Fe distance of 2.39 ± 0.02 Å for $\text{Se}_{3\text{A}/5\text{A}}$, which correlate well to the disparate pre-edge intensities observed by HERFD XAS. Noticeably, the average Se–Fe distance of the CO-inhibited CO- $\text{Se}_{3\text{A}/5\text{A}}$ at 2.40 ± 0.01 Å is indistinguishable (within error) from the non-CO-inhibited $\text{Se}_{3\text{A}/5\text{A}}$ distance of 2.39 ± 0.02 Å, suggesting that the observed differences in the $\text{Se}_{3\text{A}/5\text{A}}$ and CO- $\text{Se}_{3\text{A}/5\text{A}}$ Se HERFD XAS spectra (in particular the pre-edge intensities) are dominated by electronic, rather than geometric, effects. The shorter Se–Fe distance of $\text{Se}_{2\text{B}}$ is consistent with the average $\text{Se}_{2\text{B}}$ –Fe distance observed crystallographically in $\text{Se}_{2\text{B}}$ - FeMoco (2.30 ± 0.03 Å), as well as Se–Fe distances crystallographically observed in synthetic $[\text{Fe}_2\text{S}_2]^{2+/+0}$ complexes of ca. 2.30–2.34 Å.^{61–63} The significantly longer Se–Fe distances of $\text{Se}_{3\text{A}/5\text{A}}$ and CO- $\text{Se}_{3\text{A}/5\text{A}}$ suggest the presence of more reduced Fe centers, but secondary interactions from the protein environment could also be affecting the Se–Fe bonding.

3.6. Comparison to S XAS of $[\text{Fe}_2\text{S}_2]^{m+}$ Series. In order to gain a more quantitative estimate of the Fe oxidation states at the Se bridge sites from the HERFD XAS spectra, a comparison was made to the S XAS spectra of the previously reported $[\text{Fe}_2\text{S}_2]^{2+/+0}$ redox series (Table 1 and Figure S4). This series is unique in that it represents the only known, well-characterized synthetic Fe_2S_2 system spanning all three redox states (diferric, mixed-valent, and diferrous), and thus offers the best comparison to the protein data. While the absolute pre-edge and edge energies cannot be directly compared between the S and Se K-edge spectra, the relative metric $|\delta_{\text{p}}|$ and pre-edge areas afford a natural contrast for the two

Table 3. Parameters for Multicomponent Single Scattering Path Fits to Se HERFD EXAFS Data and Deconvoluted Average Se–Fe Distances at the 2B and 3A/5A Positions

| sample | path | ΔE_0^a (eV) | N | R (Å) | σ^2 (10^{-3} Å ²) | R-factor | reduced χ^2 |
|-----------------------|--------|---------------------|---|---|---|----------|------------------|
| Av1Se _{lo} | Se–Fe | | 2 | 2.314 (0.004) | 2.24 (0.47) | | |
| | Se–S | 3.403 | 4 | 3.950 (0.020) | 3.29 (4.38) | 0.01716 | 8.63 |
| | Se–Fe' | | 4 | 4.657 (0.013) | 3.90 (2.78) | | |
| Av1Se _{hi} | Se–Fe | | 2 | 2.322 (0.007) | 2.24 (0.47) | | |
| | Se–S | 3.107 | 4 | 3.178 (0.092) | 47.0 (11.6) | 0.04962 | 11.33 |
| | Se–Fe' | | 4 | 4.633 (0.028) | 8.00 (fixed) | | |
| Av1Se _{reac} | Se–Fe | | 2 | 2.338 (0.005) | 2.24 (0.74) | | |
| | Se–S | 3.451 | 4 | 3.950 (0.042) | 3.29 (12.2) | 0.03924 | 2.97 |
| | Se–Fe' | | 4 | 4.657 (0.027) | 3.90 (7.55) | | |
| Av1SeCO | Se–Fe | | 2 | 2.388 (0.004) | 5.06 (0.51) | | |
| | Se–S | 3.710 | 4 | 3.248 (0.052) | 43.6 (9.92) | 0.01833 | 23.93 |
| | Se–Fe' | | 4 | 4.667 (0.014) | 4.87 (3.38) | | |
| position | | | | deconvoluted average Se–Fe distance (Å) | | | |
| Se _{2B} | | | | 2.29 (0.02) | | | |
| Se _{3A/5A} | | | | 2.39 (0.02) | | | |
| COSe _{3A/5A} | | | | 2.40 (0.01) | | | |

 $aE_0 = 12\,661.55$ eV**Figure 9.** TDDFT calculations (right) of antiferromagnetically coupled $[\text{Fe}_2\text{Se}_2]^{n+}$ series (structure on left), where one selenide engages in hydrogen bonding to a bis-urea molecule (plotted dashed lines) and one selenide does not (plotted solid lines).

spectroscopies as supported by our TDDFT calculations, *vide supra*. Through comparison of both the pre-edge area and $|\delta_d|$, it was observed that Se_{2B} is most similar to the diferric $[\text{Fe}_2\text{S}_2]^{2+}$, as both exhibit large pre-edge areas (385 ± 20 vs 350) and large $|\delta_d|$ values (7.38 ± 0.06 and 7.25 eV). Both Se_{3A/5A} and CO-Se_{3A/5A} have smaller relative Fe 3d-manifold energies (5.76 ± 0.04 and 5.95 ± 0.08 eV) more consistent with the diferric $[\text{Fe}_2\text{S}_2]^0$ complex (6.27 eV), though the pre-edge intensities are what most differentiates the two. The greater intensity of CO-Se_{3A/5A} (245 ± 10) is most similar to the mixed-valent $[\text{Fe}_2\text{S}_2]^+$ complex (260), while the smaller pre-edge of Se_{3A/5A} (130 ± 30) better matches that of the diferric $[\text{Fe}_2\text{S}_2]^0$ complex (180). These comparisons suggest that the features of the Se_{2B} spectrum are most consistent with a Se bridging two Fe³⁺ centers, while the CO-Se_{3A/5A} features are more consistent with an average oxidation state of Fe^{2.5+} and the Se_{3A/5A} features suggests an average oxidation state of Fe²⁺. It should be cautioned, however, that these comparisons to synthetic model complexes fail to take into account the effects of electrostatic and hydrogen bonding interactions present in the protein, which may be significant.

3.7. Probing Electrostatic and Hydrogen Bonding Effects through Small Molecule TDDFT Calculations.

In order to quantitatively estimate the effects of hydrogen bonding on the Se XAS spectrum, we turned to TDDFT calculations of a fictitious $[\text{Fe}_2\text{Se}_2]^{n+}$ series in which one of the selenide bridges is hydrogen bonding with a bis-urea small molecule to simulate the interactions of the Se_{3A} position with the neighboring protein residues (Figure 9 and Table 4). As only one of the bridging selenides is engaged in hydrogen

Table 4. Calculated Se XAS Pre-edge Areas and Relative Fe 3d-Manifold Energies in H-Bonded Dimer

| sample | spin-state | H-bonds | calculated pre-edge area (a.u.) | calculated $ \delta_d $ (eV) |
|--|------------|---------|---------------------------------|------------------------------|
| bis-urea + $[\text{Fe}_2\text{Se}_2]^{2+}$ | 0 | 0 | 390 | 10.10 |
| | | 4 | 275 | 6.60 |
| bis-urea + $[\text{Fe}_2\text{Se}_2]^{1+}$ | 1/2 | 0 | 245 | 10.00 |
| | | 4 | 175 | 7.60 |
| bis-urea + $[\text{Fe}_2\text{Se}_2]^0$ | 0 | 0 | 90 | 8.30 |
| | | 4 | 60 | 6.20 |

bonding, the second selenide can be used as a control to directly compare the effects of the hydrogen bonding interaction. Our computations reveal that at every oxidation state (diferric/mixed-valent/diferrous), the hydrogen bonding interaction resulted in a decrease in pre-edge intensity, nearly equivalent to the effect of one-electron reduction of the dimer (a decrease in the average iron oxidation state by 0.5 units). While the presence of the hydrogen bonding interaction results in significant geometric distortions to the $[\text{Fe}_2\text{Se}_2]$ core, we found that the geometric changes do not fully account for the observed changes in pre-edge intensity, as maintaining the geometric distortion while removing the hydrogen bond donor results in regaining a significant portion of the pre-edge intensity lost due to hydrogen bonding. This is consistent with our earlier calculations that reveal negligible geometric effect on ligand pre-edge intensity and further supports our interpretation of the differences between the $\text{Se}_{3\text{A}/5\text{A}}$ and $\text{CO-Se}_{3\text{A}/5\text{A}}$ spectra as largely arising from electronic rather than geometric perturbations.

Furthermore, we sought to briefly explore electrostatic effects on Se XAS. The diferric complex $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$ afforded a convenient combined experimental and computational case study, as the crystal structure exhibits two tetraethylammonium cations in relatively close proximity to each Se atom (Se–N distances of ca. 5 Å). As previously discussed, the experimental spectrum of $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$ reveals an intense pre-edge (area = 385) consistent with our earlier calculations and comparisons to the S XAS data of synthetic analogues, though the $|\delta_d|$ value (6.10 eV) is slightly lower than anticipated for a diferric site when compared to model S XAS data. TDDFT calculations of the $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{2-}$ dianion (Figure 10, solid blue line)

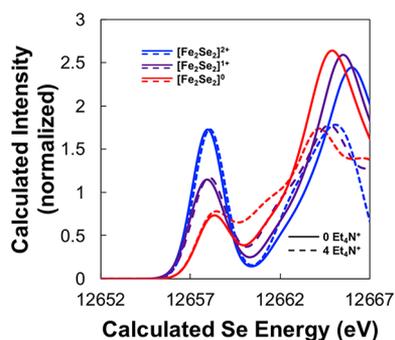


Figure 10. TDDFT calculated Se XAS spectra of antiferromagnetically coupled $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{(4-n)-}$ series ($n = 2, 1, 0$) in the absence (solid lines) and in the presence (dashed lines) of the four crystallographically nearest Et_4N^+ cations observed in $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$, demonstrating the inclusion of the cations decreases $|\delta_d|$ for all oxidation states. All calculations performed on the crystal structure of $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$ with geometry-optimized hydrogen atoms.

reveal the expected intense pre-edge feature and large $|\delta_d|$ splitting (Table 5) consistent with our earlier calculations for a diferric dimer, suggesting a $|\delta_d|$ somewhat larger than what is observed experimentally. However, when we explicitly include the four nearest $[\text{Et}_4\text{N}]^+$ cations in our TDDFT calculation (Figure 10, dashed blue line), the pre-edge intensity remains relatively unperturbed, while $|\delta_d|$ decreases by ca. 1 eV. This effect is observed across all oxidation states, though it is most pronounced at the diferric level. These relatively simplistic calculations suggest that while electrostatic interactions will have a negligible effect on pre-edge intensity, the perturbation of the rising edge may be significant. Though a more comprehensive experimental and theoretical study on bridging selenide systems will be necessary to better quantify the effects of such secondary interactions on the Se K-edge spectrum, the present results emphasize the utility of Se HERFD XAS for selectively probing the belt positions in FeMoco and further demonstrate the sensitivity of the Se spectra to the bound CO substrate.

3.8. FeMoco Electronic Structure Picture. The results of the Se HERFD XAS experiments on Se-substituted FeMoco reveal the presence of significant asymmetry in the electronic structure of the nitrogenase cofactor, with the Fe2/Fe6 edge exhibiting more local oxidized character while the Fe4/Fe5/Fe3/Fe7 face exhibits more reduced character. This electronic asymmetry is consistent with the structural asymmetry observed crystallographically at the three bridging sulfide positions (Figure 11), wherein protein–cofactor interactions break the formal 3-fold symmetry of the metallocofactor and likely contribute to significant charge localization. Specifically, it has been rationalized that the positive charges of the two conserved arginine residues, R96 and R359, located in close proximity to the 5A bridge position could help to stabilize local ferrous character at Fe3 and Fe7.¹⁶ Similarly, we hypothesize that the four amide groups of R359, L358, G357, and G356 engage in hydrogen bonding to the 3A bridge position and that these interactions could stabilize a more locally reduced state at Fe4 and Fe5. It has previously been shown that hydrogen bonding interactions to thiolate ligands in both protein^{64,65} and synthetic model complexes^{66–68} can significantly shift the reduction potentials of bound Fe centers, with increasing hydrogen bonding interactions resulting in increasing shifts in reduction potentials. Thus, these two sets of interactions (electrostatic and hydrogen bonding) at the 3A and 5A bridge positions are anticipated to localize charge density to the Fe4/Fe5/Fe3/Fe7 face of the cofactor, while the weaker single hydrogen bond interaction at the 2B bridge position with histidine H195 would favor a more locally oxidized Fe2/Fe6 edge. This interpretation is also consistent with the Se HERFD EXAFS data, which reveals a shorter average Se–Fe distance at the 2B position and a significantly longer average Se–Fe distance at the 3A/5A positions.

Table 5. Calculated and Experimental Se XAS Pre-edge Areas and Relative Fe 3d-Manifold Energies for $[\text{Fe}_2\text{Se}_2]^{n+}$

| sample | spin-state | calculated pre-edge area (a.u.) | calculated $ \delta_d $ (eV) | experimental pre-edge area (a.u.) | experimental $ \delta_d $ (eV) |
|--|------------|---------------------------------|------------------------------|-----------------------------------|--------------------------------|
| $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{2-}$ | 0 | 375 | 8.10 | 385 | 6.05 |
| $\{[\text{Et}_4\text{N}]_4[\text{Fe}_2\text{Se}_2(\text{SPh})_4]\}^{2+}$ | 0 | 380 | 7.00 | – | – |
| $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{3-}$ | 1/2 | 285 | 7.60 | – | – |
| $\{[\text{Et}_4\text{N}]_4[\text{Fe}_2\text{Se}_2(\text{SPh})_4]\}^{1+}$ | 1/2 | 280 | 6.70 | – | – |
| $[\text{Fe}_2\text{Se}_2(\text{SPh})_4]^{4-}$ | 0 | 145 | 6.70 | – | – |
| $\{[\text{Et}_4\text{N}]_4[\text{Fe}_2\text{Se}_2(\text{SPh})_4]\}^0$ | 0 | 135 | 6.00 | – | – |

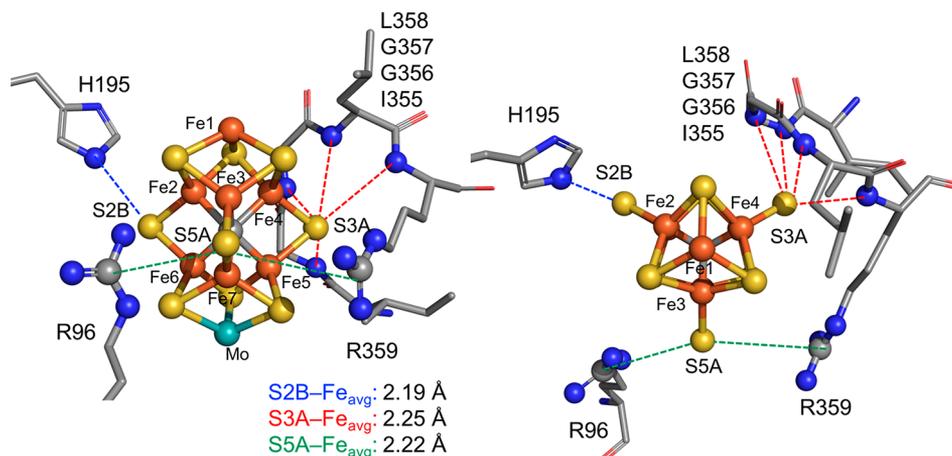


Figure 11. Asymmetric FeMoco-protein interactions in profile (left) and Fe1–Mo top-down view (right) from *A. vinelandii* (PDB-ID 3U7Q). Nearest interactions to the bridging positions 2B (blue), 3A (red), and 5A (green) are emphasized as dashed lines. Reported S–Fe distances are averages of both clusters in the crystallographic asymmetric unit. Cofactor ligands C275, H442, and homocitrate are omitted for clarity. Nitrogen atoms are shown in blue, oxygen in red, and carbon in gray.

The experimental Se HERFD XAS data coupled with the results from TDDFT calculations on small molecule model systems support assignment of Se2B as bridging two ferric Fe centers whose spins are aligned antiparallel with respect to one another. This oxidation state assignment for Fe2/Fe6 is consistent with the previous SpReAD study,¹⁶ while the relative coupling assignment is consistent with recent QM/MM calculations on FeMoco.²⁰ We emphasize that this is the first experimental data to reveal relative coupling between the Fe centers of FeMoco. Additionally, taking into account the relative effects of electrostatic and hydrogen bonding interactions on the Se K-edge absorption features, the experimental Se HERFD XAS data are suggestive of an average Fe oxidation state in the range of ca. +2.25–2.5 for the Fe4/Fe5/Fe3/Fe7 face in the resting state. This oxidation state assignment could be consistent with either the fully localized assignment of the SpReAD study or the more delocalized assignment of the 3A and 5A positions in the present data limits our ability to further distinguish between these two proposals, but in the future such a determination may be feasible if samples with sufficiently disparate 3A and 5A populations can be prepared (the initial Se migration data²¹ suggests this is theoretically possible).

Our results also suggest that upon CO binding to the cluster, the Fe4/Fe5/Fe3/Fe7 face of the cofactor is electronically perturbed, appearing more oxidized. This initially seems counterintuitive, as it is known CO does not react with the cofactor in the resting state, but requires a more reduced state of FeMoco to bind (presumably E_2 ,⁶⁹ where E_0 refers to the resting state and E_n refers to the n -electron reduced state for $n = 1, 2, 3$, etc.). However, it is unclear whether or not following CO binding the cofactor remains in a reduced state or if it can relax back to an E_0 - or E_1 -equivalent state, as proton reduction remains uninhibited by CO. Additionally, while a number of CO-inhibited states have been identified by EPR that are proposed to differ by the mode and number of the Fe–CO interactions (lo-CO, hi-CO, etc.), it has also been observed that EPR-silent species are also present during reaction with CO, suggestive of an E_1 -equivalent state being generated.^{70–72} The observation that CO–Se_{3A/5A} appears more oxidized than Se_{3A/5A} could be explained by a redox reorganization within the

cofactor, whereby electron density is shifted from the Fe4/Fe5/Fe3/Fe7 face to the Fe2/Fe6 edge where CO binds. Recently, redox reorganization upon CO-binding was observed in an electronically localized synthetic $[\text{Fe}_4\text{O}]^{8+}$ cluster, wherein an electron is transferred from a local Fe^{2+} center to a formerly Fe^{3+} center to facilitate CO binding.⁷³ We speculate that such a redox reorganization mechanism in FeMoco would likely be facilitated by the highly covalent carbide, suggesting a plausible role for the central atom during catalysis. Alternatively, a change in the relative coupling across Se3A and Se5A could also account for the observed differences in Se_{3A/5A} and CO–Se_{3A/5A}, as our TDDFT calculations on model systems have shown increased pre-edge intensity for antiferromagnetically coupled centers relative to ferromagnetic centers. Even in the localized limit, these results suggest that the electronic structure of the cluster may be more dynamic than previously assumed, further emphasizing the necessity for development of spectroscopic methods capable of extracting local electronic structure information from FeMoco, particularly under turnover conditions.

4. CONCLUSIONS

The present study demonstrates a new method for probing the local electronic structure of FeMoco and reveals a significant asymmetry with regard to the electronic distribution about the cluster. Analysis of Se HERFD XAS spectra of Se-substituted FeMoco indicate that the 2B and 3A/5A bridging positions of FeMoco are electronically distinct, with the Fe2/Fe6 edge consistent with an antiferromagnetically coupled diferric pair, while the Fe3/Fe4/Fe5/Fe7 face of the cofactor exhibits more localized ferrous character. Additionally, we have demonstrated that this technique can be extended beyond the resting state of nitrogenase through analysis of Se-substituted CO-inhibited FeMoco, and observed that the electronic structure of the Fe3/Fe4/Fe5/Fe7 face is electronically, but not geometrically, sensitive to binding of CO to the distal edge of the cofactor. A redox reorganization mechanism within the cluster has been postulated to account for the observed electronic perturbation upon CO binding. Further studies incorporating synthetic FeSe model complexes and advanced theoretical methods are currently underway in our laboratories to further exploit Se

substitution in elucidating the local electronic structure of FeMoco.

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.9b06988.

Detailed protein preparations, summaries of crystallographic data, fits of experimental spectra, and computational details (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: dcrees@caltech.edu.

*E-mail: serena.debeer@cec.mpg.de.

ORCID

Justin T. Henthorn: 0000-0003-4876-2680

Renee J. Arias: 0000-0002-6505-7513

Uwe Bergmann: 0000-0001-5639-166X

Douglas C. Rees: 0000-0003-4073-1185

Serena DeBeer: 0000-0002-5196-3400

Author Contributions

[#]J.T.H. and R.J.A. contributed equally.

Notes

The authors declare no competing financial interest. Crystallographic data has been uploaded to the PDB.

■ ACKNOWLEDGMENTS

J.T.H. and S.D. thank the Max Planck Society for funding. S.D. acknowledges funding from the European Research Council under the European Union's Seventh Framework Programme (FP/2007-2013)/ERC Grant Agreement No. 615414 and the DFG SPP 1927 "Iron Sulfur for Life" project (Project No. DE 1877/1-1). J.T.H. acknowledges funding from the Alexander von Humboldt Foundation. S.K. would like to thank The Knut and Alice Wallenberg Foundation (KAW 2014.0370) for financial support. We are grateful to Dr. T. Weyhermüller and T. Mußfeldt for supplying $[\text{Et}_4\text{N}]_2[\text{Fe}_2\text{Se}_2(\text{SPh})_4]$. We thank the Gordon and Betty Moore Foundation and the Beckman Institute at Caltech for their generous support of the Molecular Observatory at Caltech, the staff of SSRL BL12-2 for their technical assistance. Support from NIH Grant GM045162 and the Howard Hughes Medical Institute to D.C.R. and from NIH Training Grant T32GM7616 to R.J.A. is gratefully acknowledged. Use of the Stanford Synchrotron Radiation Lightsource, SLAC National Accelerator Laboratory, is supported by the U.S. Department of Energy, Office of Basic Energy Sciences, under Contract No. DE-AC02-76SF00515. The SSRL Structural Molecular Biology Program is supported by the DOE Office of Biological and Environmental Research and by the National Institutes of Health, National Institute of General Medical Sciences (including P41GM103393). The contents of this publication are solely the responsibility of the authors and do not necessarily represent the official views of NIGMS or NIH. We thank Paul Oyala and the Electron Paramagnetic Resonance Facility at Caltech for assistance with EPR measurements.

■ REFERENCES

(1) Canfield, D. E.; Glazer, A. N.; Falkowski, P. G. The Evolution and Future of Earth's Nitrogen Cycle. *Science* **2010**, *330* (6001), 192.

(2) Gruber, N.; Galloway, J. N. An Earth-system perspective of the global nitrogen cycle. *Nature* **2008**, *451*, 293.

(3) Lee, C. C.; Hu, Y.; Ribbe, M. W. Vanadium Nitrogenase Reduces CO. *Science* **2010**, *329* (5992), 642.

(4) Schlögl, R. Catalytic Synthesis of Ammonia—A "Never-Ending Story"? *Angew. Chem., Int. Ed.* **2003**, *42* (18), 2004–2008.

(5) Rofer-DePoorter, C. K. A comprehensive mechanism for the Fischer–Tropsch synthesis. *Chem. Rev.* **1981**, *81* (5), 447–474.

(6) Hoffman, B. M.; Lukoyanov, D.; Yang, Z.-Y.; Dean, D. R.; Seefeldt, L. C. Mechanism of Nitrogen Fixation by Nitrogenase: The Next Stage. *Chem. Rev.* **2014**, *114* (8), 4041–4062.

(7) Yoo, S. J.; Angove, H. C.; Papaefthymiou, V.; Burgess, B. K.; Münck, E. Mössbauer Study of the MoFe Protein of Nitrogenase from *Azotobacter vinelandii* Using Selective ^{57}Fe Enrichment of the M-Centers. *J. Am. Chem. Soc.* **2000**, *122* (20), 4926–4936.

(8) Kowalska, J.; DeBeer, S. The role of X-ray spectroscopy in understanding the geometric and electronic structure of nitrogenase. *Biochim. Biophys. Acta, Mol. Cell Res.* **2015**, *1853* (6), 1406–1415.

(9) Lima, F. A.; Bjornsson, R.; Weyhermüller, T.; Chandrasekaran, P.; Glatzel, P.; Neese, F.; DeBeer, S. High-resolution molybdenum K-edge X-ray absorption spectroscopy analyzed with time-dependent density functional theory. *Phys. Chem. Chem. Phys.* **2013**, *15* (48), 20911–20920.

(10) Kowalska, J. K.; Henthorn, J. T.; Van Stappen, C.; Trncik, C.; Einsle, O.; Keavney, D.; DeBeer, S. X-ray Magnetic Circular Dichroism Spectroscopy Applied to Nitrogenase and Related Models: Experimental Evidence for a Spin-Coupled Molybdenum(III) Center. *Angew. Chem., Int. Ed.* **2019**, *58* (28), 9373–9377.

(11) Rees, J. A.; Bjornsson, R.; Kowalska, J. K.; Lima, F. A.; Schlesier, J.; Sippel, D.; Weyhermüller, T.; Einsle, O.; Kovacs, J. A.; DeBeer, S. Comparative electronic structures of nitrogenase FeMoco and FeVco. *Dalton Transactions* **2017**, *46* (8), 2445–2455.

(12) Hoffman, B. M. ENDOR and ESEEM of a Non-Kramers Doublet in an Integer-Spin System. *J. Phys. Chem.* **1994**, *98* (45), 11657–11665.

(13) Lukoyanov, D.; Yang, Z.-Y.; Barney, B. M.; Dean, D. R.; Seefeldt, L. C.; Hoffman, B. M. Unification of reaction pathway and kinetic scheme for N_2 reduction catalyzed by nitrogenase. *Proc. Natl. Acad. Sci. U. S. A.* **2012**, *109* (15), 5583.

(14) Sturgeon, B. E.; Doan, P. E.; Liu, K. E.; Burdi, D.; Tong, W. H.; Nocek, J. M.; Gupta, N.; Stubbe, J.; Kurtz, D. M.; Lippard, S. J.; Hoffman, B. M. Non-Kramers ESEEM of Integer-Spin Diferrous Carboxylate-Bridged Clusters in Proteins. *J. Am. Chem. Soc.* **1997**, *119* (2), 375–386.

(15) Spatzal, T.; Einsle, O.; Andrade, S. L. A. Analysis of the Magnetic Properties of Nitrogenase FeMo Cofactor by Single-Crystal EPR Spectroscopy. *Angew. Chem., Int. Ed.* **2013**, *52* (38), 10116–10119.

(16) Spatzal, T.; Schlesier, J.; Burger, E.-M.; Sippel, D.; Zhang, L.; Andrade, S. L. A.; Rees, D. C.; Einsle, O. Nitrogenase FeMoco investigated by spatially resolved anomalous dispersion refinement. *Nat. Commun.* **2016**, *7*, 10902.

(17) Bjornsson, R.; Neese, F.; DeBeer, S. Revisiting the Mössbauer Isomer Shifts of the FeMoco Cluster of Nitrogenase and the Cofactor Charge. *Inorg. Chem.* **2017**, *56* (3), 1470–1477.

(18) Beinert, H.; Holm, R. H.; Münck, E. Iron-Sulfur Clusters: Nature's Modular, Multipurpose Structures. *Science* **1997**, *277* (5326), 653.

(19) Noodleman, L.; Peng, C. Y.; Case, D. A.; Mouesca, J. M. Orbital interactions, electron delocalization and spin coupling in iron-sulfur clusters. *Coord. Chem. Rev.* **1995**, *144*, 199–244.

(20) Benediktsson, B.; Bjornsson, R. QM/MM Study of the Nitrogenase MoFe Protein Resting State: Broken-Symmetry States, Protonation States, and QM Region Convergence in the FeMoco Active Site. *Inorg. Chem.* **2017**, *56* (21), 13417–13429.

(21) Spatzal, T.; Perez, K. A.; Howard, J. B.; Rees, D. C. Catalysis-dependent selenium incorporation and migration in the nitrogenase active site iron-molybdenum cofactor. *eLife* **2015**, *4*, No. e11620.

- (22) Perez, K. A. Ph.D. Thesis, California Institute of Technology, Pasadena, 2016.
- (23) Spatzal, T.; Perez, K. A.; Einsle, O.; Howard, J. B.; Rees, D. C. Ligand binding to the FeMo-cofactor: Structures of CO-bound and reactivated nitrogenase. *Science* **2014**, *345* (6204), 1620–1623.
- (24) Sokaras, D.; Weng, T. C.; Nordlund, D.; Alonso-Mori, R.; Velikov, P.; Wenger, D.; Garachtchenko, A.; George, M.; Borzenets, V.; Johnson, B.; Rabedeau, T.; Bergmann, U. A seven-crystal Johann-type hard x-ray spectrometer at the Stanford Synchrotron Radiation Lightsource. *Rev. Sci. Instrum.* **2013**, *84* (5), No. 053102.
- (25) Ravel, B.; Newville, M. ATHENA, ARTEMIS, HEPHAESTUS: data analysis for X-ray absorption spectroscopy using IFEFFIT. *J. Synchrotron Radiat.* **2005**, *12* (4), 537–541.
- (26) DeBeer George, S.; Neese, F. Calibration of Scalar Relativistic Density Functional Theory for the Calculation of Sulfur K-Edge X-ray Absorption Spectra. *Inorg. Chem.* **2010**, *49* (4), 1849–1853.
- (27) DeBeer George, S.; Petrenko, T.; Neese, F. Time-dependent density functional calculations of ligand K-edge X-ray absorption spectra. *Inorg. Chim. Acta* **2008**, *361* (4), 965–972.
- (28) Neese, F. Prediction of molecular properties and molecular spectroscopy with density functional theory: From fundamental theory to exchange-coupling. *Coord. Chem. Rev.* **2009**, *253* (5), 526–563.
- (29) Neese, F. The ORCA program system. *Wiley Interdisciplinary Reviews: Computational Molecular Science* **2012**, *2* (1), 73–78.
- (30) Staroverov, V. N.; Scuseria, G. E.; Tao, J.; Perdew, J. P. Comparative assessment of a new nonempirical density functional: Molecules and hydrogen-bonded complexes. *J. Chem. Phys.* **2003**, *119* (23), 12129–12137.
- (31) Tao, J.; Perdew, J. P.; Staroverov, V. N.; Scuseria, G. E. Climbing the Density Functional Ladder: Nonempirical Meta-Generalized Gradient Approximation Designed for Molecules and Solids. *Phys. Rev. Lett.* **2003**, *91* (14), 146401.
- (32) Grimme, S.; Antony, J.; Ehrlich, S.; Krieg, H. A consistent and accurate ab initio parametrization of density functional dispersion correction (DFT-D) for the 94 elements H-Pu. *J. Chem. Phys.* **2010**, *132* (15), 154104.
- (33) Grimme, S.; Ehrlich, S.; Goerigk, L. Effect of the damping function in dispersion corrected density functional theory. *J. Comput. Chem.* **2011**, *32* (7), 1456–1465.
- (34) Lenthe, E. v.; Baerends, E. J.; Snijders, J. G. Relativistic regular two-component Hamiltonians. *J. Chem. Phys.* **1993**, *99* (6), 4597–4610.
- (35) van Wüllen, C. Molecular density functional calculations in the regular relativistic approximation: Method, application to coinage metal diatomics, hydrides, fluorides and chlorides, and comparison with first-order relativistic calculations. *J. Chem. Phys.* **1998**, *109* (2), 392–399.
- (36) Pantazis, D. A.; Chen, X.-Y.; Landis, C. R.; Neese, F. All-Electron Scalar Relativistic Basis Sets for Third-Row Transition Metal Atoms. *J. Chem. Theory Comput.* **2008**, *4* (6), 908–919.
- (37) Weigend, F.; Ahlrichs, R. Balanced basis sets of split valence, triple zeta valence and quadruple zeta valence quality for H to Rn: Design and assessment of accuracy. *Phys. Chem. Chem. Phys.* **2005**, *7* (18), 3297–3305.
- (38) Izsák, R.; Neese, F. An overlap fitted chain of spheres exchange method. *J. Chem. Phys.* **2011**, *135* (14), 144105.
- (39) Neese, F.; Wennmohs, F.; Hansen, A.; Becker, U. Efficient, approximate and parallel Hartree–Fock and hybrid DFT calculations. A ‘chain-of-spheres’ algorithm for the Hartree–Fock exchange. *Chem. Phys.* **2009**, *356* (1), 98–109.
- (40) Kowalska, J. K.; Hahn, A. W.; Albers, A.; Schiewer, C. E.; Bjornsson, R.; Lima, F. A.; Meyer, F.; DeBeer, S. X-ray Absorption and Emission Spectroscopic Studies of [L₂Fe₂S₂]_n Model Complexes: Implications for the Experimental Evaluation of Redox States in Iron–Sulfur Clusters. *Inorg. Chem.* **2016**, *55* (9), 4485–4497.
- (41) Glaser, T.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. Ligand K-Edge X-ray Absorption Spectroscopy: A Direct Probe of Ligand–Metal Covalency. *Acc. Chem. Res.* **2000**, *33* (12), 859–868.
- (42) Solomon, E. I.; Hedman, B.; Hodgson, K. O.; Dey, A.; Szilagy, R. K. Ligand K-edge X-ray absorption spectroscopy: covalency of ligand–metal bonds. *Coord. Chem. Rev.* **2005**, *249* (1), 97–129.
- (43) Dey, A.; Jenney, F. E.; Adams, M. W. W.; Johnson, M. K.; Hodgson, K. O.; Hedman, B.; Solomon, E. I. Sulfur K-Edge X-ray Absorption Spectroscopy and Density Functional Theory Calculations on Superoxide Reductase: Role of the Axial Thiolate in Reactivity. *J. Am. Chem. Soc.* **2007**, *129* (41), 12418–12431.
- (44) Dey, A.; Jiang, Y.; Ortiz de Montellano, P.; Hodgson, K. O.; Hedman, B.; Solomon, E. I. S K-edge XAS and DFT Calculations on Cytochrome P450: Covalent and Ionic Contributions to the Cysteine-Fe Bond and Their Contribution to Reactivity. *J. Am. Chem. Soc.* **2009**, *131* (22), 7869–7878.
- (45) Dey, A.; Peng, Y.; Broderick, W. E.; Hedman, B.; Hodgson, K. O.; Broderick, J. B.; Solomon, E. I. S K-edge XAS and DFT Calculations on SAM Dependent Pyruvate Formate-Lyase Activating Enzyme: Nature of Interaction between the Fe₄S₄ Cluster and SAM and its Role in Reactivity. *J. Am. Chem. Soc.* **2011**, *133* (46), 18656–18662.
- (46) Dey, A.; Roche, C. L.; Walters, M. A.; Hodgson, K. O.; Hedman, B.; Solomon, E. I. Sulfur K-Edge XAS and DFT Calculations on [Fe₄S₄]²⁺ Clusters: Effects of H-bonding and Structural Distortion on Covalency and Spin Topology. *Inorg. Chem.* **2005**, *44* (23), 8349–8354.
- (47) Glaser, T.; Rose, K.; Shadle, S. E.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. S K-edge X-ray Absorption Studies of Tetranuclear Iron–Sulfur Clusters: μ -Sulfide Bonding and Its Contribution to Electron Delocalization. *J. Am. Chem. Soc.* **2001**, *123* (3), 442–454.
- (48) Ha, Y.; Arnold, A. R.; Nuñez, N. N.; Bartels, P. L.; Zhou, A.; David, S. S.; Barton, J. K.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. Sulfur K-Edge XAS Studies of the Effect of DNA Binding on the [Fe₄S₄] Site in EndoIII and MutY. *J. Am. Chem. Soc.* **2017**, *139* (33), 11434–11442.
- (49) Sun, N.; Dey, A.; Xiao, Z.; Wedd, A. G.; Hodgson, K. O.; Hedman, B.; Solomon, E. I. Solvation Effects on S K-Edge XAS Spectra of Fe–S Proteins: Normal and Inverse Effects on WT and Mutant Rubredoxin. *J. Am. Chem. Soc.* **2010**, *132* (36), 12639–12647.
- (50) Bjornsson, R.; Delgado-Jaime, M. U.; Lima, F. A.; Sippel, D.; Schlesier, J.; Weyhermüller, T.; Einsle, O.; Neese, F.; DeBeer, S. Molybdenum L-Edge XAS Spectra of MoFe Nitrogenase. *Z. Anorg. Allg. Chem.* **2015**, *641* (1), 65–71.
- (51) Hedman, B.; Frank, P.; Gheller, S. F.; Roe, A. L.; Newton, W. E.; Hodgson, K. O. New structural insights into the iron-molybdenum cofactor from *Azotobacter vinelandii* nitrogenase through sulfur K and molybdenum L x-ray absorption edge studies. *J. Am. Chem. Soc.* **1988**, *110* (12), 3798–3805.
- (52) Kvashnina, K. O.; Butorin, S. M.; Cui, D.; Vegelius, J.; Puranen, A.; Gens, R.; Glatzel, P. Electron transfer during selenium reduction by iron surfaces in aqueous solution: High resolution X-ray absorption study. *Journal of Physics: Conference Series* **2009**, *190* (1), No. 012191.
- (53) Krause, M. O.; Oliver, J. H. Natural widths of atomic K and L levels, $K\alpha$ X-ray lines and several KLL Auger lines. *J. Phys. Chem. Ref. Data* **1979**, *8* (2), 329–338.
- (54) Pickering, I. J.; George, G. N.; Van Fleet-Stalder, V.; Chasteen, T. G.; Prince, R. C. X-ray absorption spectroscopy of selenium-containing amino acids. *JBIC, J. Biol. Inorg. Chem.* **1999**, *4* (6), 791–794.
- (55) Castillo, R. G.; Banerjee, R.; Allpress, C. J.; Rohde, G. T.; Bill, E.; Que, L.; Lipscomb, J. D.; DeBeer, S. High-Energy-Resolution Fluorescence-Detected X-ray Absorption of the Q Intermediate of Soluble Methane Monooxygenase. *J. Am. Chem. Soc.* **2017**, *139* (49), 18024–18033.
- (56) Hämäläinen, K.; Siddons, D. P.; Hastings, J. B.; Berman, L. E. Elimination of the inner-shell lifetime broadening in x-ray-absorption spectroscopy. *Phys. Rev. Lett.* **1991**, *67* (20), 2850–2853.
- (57) Kowalska, J. K.; Lima, F. A.; Pollock, C. J.; Rees, J. A.; DeBeer, S. A Practical Guide to High-resolution X-ray Spectroscopic Measurements and their Applications in Bioinorganic Chemistry. *Isr. J. Chem.* **2016**, *56* (9–10), 803–815.

(58) Gaillard, J.; Moulis, J. M.; Auric, P.; Meyer, J. High-multiplicity spin states of $2[4\text{Fe-4Se}]^+$ clostridial ferredoxins. *Biochemistry* **1986**, *25* (2), 464–468.

(59) Carney, M. J.; Papaefthymiou, G. C.; Spartalian, K.; Frankel, R. B.; Holm, R. H. Ground spin state variability in $[\text{Fe}_4\text{S}_4(\text{SR})_4]^{3-}$. Synthetic analogs of the reduced clusters in ferredoxins and other iron-sulfur proteins: cases of extreme sensitivity of electronic state and structure to extrinsic factors. *J. Am. Chem. Soc.* **1988**, *110* (18), 6084–6095.

(60) Davis, L. C.; Henzl, M. T.; Burris, R. H.; Orme-Johnson, W. H. Iron-sulfur clusters in the molybdenum-iron protein component of nitrogenase. Electron paramagnetic resonance of the carbon monoxide inhibited state. *Biochemistry* **1979**, *18* (22), 4860–4869.

(61) Fohlmeister, L.; Vignesh, K. R.; Winter, F.; Moubaraki, B.; Rajaraman, G.; Pottgen, R.; Murray, K. S.; Jones, C. Neutral diiron(III) complexes with $\text{Fe}_2(\mu_2\text{-E})_2$ (E = O, S, Se) core structures: reactivity of an iron(I) dimer towards chalcogens. *Dalton Transactions* **2015**, *44* (4), 1700–1708.

(62) Panda, C.; Menezes, P. W.; Walter, C.; Yao, S.; Miehlich, M. E.; Gutkin, V.; Meyer, K.; Driess, M. From a Molecular 2Fe-2Se Precursor to a Highly Efficient Iron Diselenide Electrocatalyst for Overall Water Splitting. *Angew. Chem., Int. Ed.* **2017**, *56* (35), 10506–10510.

(63) Yu, S. B.; Papaefthymiou, G. C.; Holm, R. H. Comprehensive iron-selenium-thiolate cluster chemistry. *Inorg. Chem.* **1991**, *30* (18), 3476–3485.

(64) Anxolabéhère-Mallart, E.; Glaser, T.; Frank, P.; Aliverti, A.; Zanetti, G.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. Sulfur K-Edge X-ray Absorption Spectroscopy of 2Fe-2S Ferredoxin: Covalency of the Oxidized and Reduced 2Fe Forms and Comparison to Model Complexes. *J. Am. Chem. Soc.* **2001**, *123* (23), 5444–5452.

(65) Yoshioka, S.; Tosha, T.; Takahashi, S.; Ishimori, K.; Hori, H.; Morishima, I. Roles of the Proximal Hydrogen Bonding Network in Cytochrome P450cam-Catalyzed Oxygenation. *J. Am. Chem. Soc.* **2002**, *124* (49), 14571–14579.

(66) Dey, A.; Okamura, T.-a.; Ueyama, N.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. Sulfur K-Edge XAS and DFT Calculations on P450 Model Complexes: Effects of Hydrogen Bonding on Electronic Structure and Redox Potentials. *J. Am. Chem. Soc.* **2005**, *127* (34), 12046–12053.

(67) Ueno, T.; Nishikawa, N.; Moriyama, S.; Adachi, S.; Lee, K.; Okamura, T.-a.; Ueyama, N.; Nakamura, A. Role of the Invariant Peptide Fragment Forming $\text{NH}\cdots\text{S}$ Hydrogen Bonds in the Active Site of Cytochrome P-450 and Chloroperoxidase: Synthesis and Properties of Cys-Containing Peptide Fe(III) and Ga(III) (Octaethylporphinato) Complexes as Models. *Inorg. Chem.* **1999**, *38* (6), 1199–1210.

(68) Ueyama, N.; Terakawa, T.; Nakata, M.; Nakamura, A. Positive shift of redox potential of $[\text{Fe}_4\text{S}_4(\text{Z-cys-Gly-Ala-OMe})_4]^{2-}$ in dichloromethane. *J. Am. Chem. Soc.* **1983**, *105* (24), 7098–7102.

(69) Lowe, D. J.; Fisher, K.; Thorneley, R. N. F. *Klebsiella pneumoniae* nitrogenase. Mechanism of acetylene reduction and its inhibition by carbon monoxide. *Biochem. J.* **1990**, *272* (3), 621.

(70) Cameron, L. M.; Hales, B. J. Investigation of CO Binding and Release from Mo-Nitrogenase during Catalytic Turnover. *Biochemistry* **1998**, *37* (26), 9449–9456.

(71) Yan, L.; Dapper, C. H.; George, S. J.; Wang, H.; Mitra, D.; Dong, W.; Newton, W. E.; Cramer, S. P. Photolysis of Hi-CO Nitrogenase – Observation of a Plethora of Distinct CO Species Using Infrared Spectroscopy. *Eur. J. Inorg. Chem.* **2011**, *2011* (13), 2064–2074.

(72) Yan, L.; Pelmenschikov, V.; Dapper, C. H.; Scott, A. D.; Newton, W. E.; Cramer, S. P. IR-Monitored Photolysis of CO-Inhibited Nitrogenase: A Major EPR-Silent Species with Coupled Terminal CO Ligands. *Chem. - Eur. J.* **2012**, *18* (51), 16349–16357.

(73) Arnett, C. H.; Chalkley, M. J.; Agapie, T. A Thermodynamic Model for Redox-Dependent Binding of Carbon Monoxide at Site-Differentiated, High Spin Iron Clusters. *J. Am. Chem. Soc.* **2018**, *140* (16), 5569–5578.