

1 **Title: Characterization of the brain functional architecture of psychostimulant withdrawal**
2 **using single-cell whole brain imaging**

3

4 **Abbreviated Title: Brain architecture of psychostimulant withdrawal**

5

6 **Adam Kimbrough, Ph.D.,¹ Lauren C. Smith,^{1,2} Marsida Kallupi, Ph.D.,¹**

7 **Sierra Simpson,^{1,2} Andres Collazo, Ph.D.,³ Olivier George, Ph.D.^{1,*}**

8

9 ¹School of Medicine, Department of Psychiatry, University of California San Diego, MC 0667,
10 La Jolla, California, 92093. ²Department of Neuroscience, The Scripps Research Institute, La
11 Jolla, California 92037. ³Beckman Institute, Cal-Tech, MC 139-74, Pasadena, California, 91125.

12

13 *Correspondence:

14 Dr. Olivier George, School of Medicine, Department of Psychiatry, University of California San
15 Diego, 9500 Gilman Drive, MC 0714, La Jolla, CA 92093-0737. E-mail: olgeorge@ucsd.edu.

16 Telephone: +1-858-822-0323

17

18 Number of pages: 36

19 Number of Figures: 6

20 Number of Tables: 6

21 Number of Words (Abstract): 165

22 Number of Words: (Introduction): 337

23 Number of Words: (Discussion): 1353

24

25 This work was supported by National Institutes of Health grants DA044451, DA043799,
26 DA047113, AA006420, AA020608, AA022977, AA027301, and AA007456, Tobacco-Related
27 Disease Research Program grant 27IR-0047, and the Pearson Center for Alcoholism and
28 Addiction Research. The authors would like to thank Michael Arends for editorial assistance
29 with the manuscript.

30

31 The authors declare no conflicts of interest

32

33 Code and data will be made available upon request.

34

35 Author Contributions: A.K. and O.G. designed the research. A.K., A.C., L.C.S, M.K., and S.S.
36 performed the experiments and collected the data. A.K. and L.C.S. designed the figures. A.K.
37 analyzed the data. A.K. and O.G. wrote the manuscript. All of the authors read and approved the
38 final manuscript.

39

40

41

42 **Abstract**

43 Numerous brain regions have been identified as contributing to addiction-like behaviors, but
44 unclear is the way in which these brain regions as a whole lead to addiction. The search for a
45 final common brain pathway that is involved in addiction remains elusive. To address this
46 question, we used male C57BL/6J mice and performed single-cell whole-brain imaging of neural
47 activity during withdrawal from cocaine, methamphetamine, and nicotine. We used hierarchical
48 clustering and graph theory to identify similarities and differences in brain functional
49 architecture. Although methamphetamine and cocaine shared some network similarities, the
50 main common neuroadaptation between these psychostimulant drugs was a dramatic decrease in
51 modularity, with a shift from a cortical- to subcortical-driven network, including a decrease in
52 total hub brain regions. These results demonstrate that psychostimulant withdrawal produces the
53 drug-dependent remodeling of functional architecture of the brain and suggest that the decreased
54 modularity of brain functional networks and not a specific set of brain regions may represent the
55 final common pathway that leads to addiction.

56

57 **Significance Statement**

58 A key aspect of treating drug abuse is understanding similarities and differences of how drugs of
59 abuse affect the brain. In the present study we examined how the brain is altered during withdrawal
60 from psychostimulants. We found that each drug produced a unique pattern of activity in the brain,
61 but that brains in withdrawal from cocaine and methamphetamine shared similar features.
62 Interestingly, we found the major common link between withdrawal from all psychostimulants,
63 when compared to controls, was a shift in the broad organization of the brain in the form of reduced

64 modularity. Reduced modularity has been shown in several brain disorders, including traumatic
65 brain injury, and dementia, and may be the common link between drugs of abuse.

66

67

68 **Introduction**

69 Psychostimulants are a class of highly addictive and commonly abused drugs that includes
70 cocaine, nicotine, and methamphetamine [1, 2]. A large number of brain regions have been
71 implicated in dependence and addiction-like behaviors that are associated with psychostimulant
72 use [3-9]. However, the complete neural network that is associated with psychostimulant
73 withdrawal remains understudied, and the search for a common brain pathway that is responsible
74 for psychostimulant withdrawal remains elusive. Common features of dependence may not be
75 found at the brain region level but rather at the network level.

76 The identification of changes in neural network structure that are caused by
77 psychostimulant withdrawal may be critical to understanding the ways in which these drugs affect
78 the brain in a common way. Previous studies identified changes in network function after
79 psychostimulant use [10-13], but these analyses focused on macroscale changes and not the
80 mesoscale level, or they focused on preselected regions of interest.

81 The present study sought to identify the ways in which withdrawal from different
82 commonly abused psychostimulants alters functional architecture of the brain. We hypothesized
83 that withdrawal from psychostimulants would result in major changes in functional neural
84 networks and decrease modular structuring of the brain. We further hypothesized that each
85 psychostimulant that was examined herein (i.e., methamphetamine, nicotine, and cocaine) would
86 have a unique neural network that is associated with withdrawal. We measured single-cell whole-
87 brain activity using Fos as a marker for neuronal activation in mice that underwent withdrawal
88 from chronic psychostimulant (cocaine, methamphetamine, and nicotine) administration. The
89 psychostimulant doses were chosen based on previous studies that reported rewarding effects
90 during use and observed withdrawal-like symptoms after the cessation of chronic exposure for

91 each drug [14-20]. We then used single-cell whole-brain activity to identify coactivation patterns
92 of brain regions in the network that was associated with each treatment using hierarchical
93 clustering. The coactivation patterns were used to determine the modular structuring of each
94 network. Graph theory was then used to further characterize each network to determine the brain
95 regions that are responsible for intra- and intermodular connectivity.

96

97 **Results**

98 *Psychostimulant withdrawal induces massive restructuring of the brain*

99 We examined the ways in which withdrawal from different psychostimulants alters neural
100 coactivation and modular structuring of the brain. For an overview of the experimental design, see
101 Fig. 1A. For all of the drugs tested, acute withdrawal produced widespread increases in the
102 coactivation of brain regions compared with saline controls (Fig. 1C-F). Importantly, modular
103 structuring of the brain decreased in response to withdrawal from each psychostimulant compared
104 with controls. When using a threshold of 50% of tree height, saline control mice exhibited a
105 modular structure of the brain that contained seven modules, whereas cocaine mice had four
106 modules, methamphetamine mice had three modules, and nicotine mice had five modules and one
107 isolated brain region that was not grouped with any other region (i.e., interanterodorsal nucleus of
108 the thalamus; Fig. 1B-F). Notably, the decrease in the number of modules during withdrawal was
109 independent of the clustering thresholds that were used (Fig. 1B). These data indicate that
110 psychostimulant withdrawal decreases modularity of the brain functional network compared with
111 controls.

112

113 *Characterization of individual network features*

114 To further characterize the features of each individual network, we used a graph theory
115 approach to identify potential hub brain regions with the most intramodular and intermodular
116 connectivity, which may drive activity within the network and thus be critical for neuronal
117 function in the withdrawal state. We examined positive connectivity (thresholded to a Pearson
118 correlation coefficient > 0.75 [0.75R] for inclusion as a network connection) for the network for
119 each treatment and used the modular organization that was identified by hierarchical clustering
120 to partition the regions of the networks. The 0.75R threshold was chosen because all of the brain
121 regions in each network showed connections to other regions at this threshold. Previous animal
122 model studies used various thresholds, ranging from 0.3R to 0.85R [21, 22], to examine
123 connectivity. Negative network connectivity was not examined herein because the precise
124 meaning of such connectivity is controversial and thus is not often examined in network-based
125 approaches [23-26].

126 We determined the participation coefficient (PC; i.e., a measure of importance for
127 intermodular connectivity) and the within-module degree Z-score (WMDz; i.e., a measure of
128 importance for intramodular connectivity) [27] for all brain regions in the networks. A high PC
129 was considered ≥ 0.30 , and a high WMDz was considered ≥ 0.80 . Overall, the control and
130 nicotine networks showed much greater intermodular connectivity (high PC) and a great number
131 of regions with both high intermodular and intramodular connectivity (high PC and WMDz). The
132 cocaine and methamphetamine networks showed higher levels of intramodular connectivity
133 (high WMDz) and a low number of regions with intermodular connectivity (Fig. 2A-C). We
134 named each module in each network based on the group of brain regions with the highest WMDz
135 score in the module and considered these regions to be drivers of activity within individual
136 modules (see Fig. 3-6 for names).

137

138 *The control network is driven by sensory-motor regions*

139 The saline control network had 3,176 total connections and consisted of seven modules,
140 many of which were heavily driven by sensory-motor brain regions. Of these seven modules, five
141 contained several sensory or motor brain regions that were ranked in the top five for intramodular
142 connectivity (high WMDz). In most cases, a separate set of thalamic brain regions was responsible
143 for intermodular connectivity (high PC; see Table 2 for a full list of values for the network).
144 Overall, the control network had more brain regions with high WMDz, high PC, or both in
145 individual modules compared with other networks. This indicates a more interconnected network
146 with more hub regions (Fig. 2, 3).

147

148 *The cocaine withdrawal network is driven by cortico-thalamo-hypothalamic regions*

149 The cocaine network had 7,127 total connections and consisted of four modules, one with
150 the majority of all brain regions and three others with a small subset of regions. In the large module
151 (module 1; 144 brain regions), nearly one-third (32%) of the total brain regions within the module
152 (i.e., a mixed set of midbrain-cortico-thalamic-hypothalamic-amygdalar brain regions) had high
153 WMDz. The brain regions that drive intramodular connectivity (high WMDz) in this module did
154 not have any intermodular connectivity (PC). Interestingly, only three brain regions in this module
155 (subparaventricular zone, lateral posterior nucleus of the thalamus, and frontal pole cerebral
156 cortex) reached the criterion ($PC \geq 0.30$) for a high level of intermodular connectivity, suggesting
157 sparse communication with other modules.

158 One of the smaller modules, a septal (triangular nucleus of the septum) and cortical (e.g.,
159 secondary motor area and dorsal anterior cingulate area) module (module 3) had a different set of
160 thalamic brain regions that had high PC. The other two smaller modules, a prefrontal-habenular

161 module (module 4; dorsal peduncular area [DP], induseum griseum, and lateral habenula) and a
162 thalamic (parafascicular nucleus, mediodorsal nucleus of the thalamus, and ventral medial nucleus
163 of the thalamus), midbrain (nucleus of the posterior commissure), and striatal (bed nucleus of the
164 accessory olfactory tract) module (module 2) contained regions with both a high WMDz and high
165 PC, suggesting that these regions may be potential hubs within the network. Overall, the cocaine
166 network contained the highest number of connections in any network but had minimal
167 interconnection between modules (Fig. 2, 4; see Table 3 for a full list of values for the network).

168

169 *The methamphetamine withdrawal network is driven by thalamic regions*

170 The methamphetamine network had 3,182 connections and consisted of three modules, one
171 with the majority of all brain regions and two others with a small subset of regions. In the large
172 module (module 1), a group of thalamic (e.g., intermediodorsal nucleus of the thalamus,
173 paraventricular nucleus of the thalamus, intergeniculate leaflet of the lateral geniculate complex,
174 and ventral part of the lateral geniculate complex) and amygdalar (intercalated amygdala, central
175 amygdala, and lateral amygdala) regions had high WMDz, but these brain regions did not have
176 any intermodular connectivity (PC), and a separate set of hypothalamic, cortical, and
177 mid/hindbrain regions was responsible for intermodular connectivity.

178 The second module (module 2) had several hypothalamic (e.g., mammillary body,
179 ventrolateral preoptic nucleus, and tuberal nucleus) and pallidal (globus pallidus and internal
180 segment) brain regions with high WMDz and a separate set of cortical regions (e.g., DP and orbital
181 area, ventral part) and midbrain regions (e.g., posterior pretectal nucleus, nucleus of the posterior
182 commissure, and nucleus of darkschewitsch) that had high interconnectivity with other modules
183 (high PC).

184 The third module (module 3), a thalamic module, had several thalamic regions with high
185 WMDz (e.g., ventral medial nucleus of the thalamus, posterior complex of the thalamus,
186 parafascicular nucleus, and lateral dorsal nucleus of the thalamus). Interestingly, within this
187 module, a separate set of thalamic regions (e.g., paracentral nucleus, ventral anterior-lateral
188 complex of the thalamus, ventral posterior complex of the thalamus, and anterodorsal nucleus) had
189 high PC, indicating that this module is internally directed by thalamic regions and also externally
190 communicates through these regions. Overall, the methamphetamine network had a similar
191 number of total connections to the control network, but it had minimal interconnections between
192 modules (Fig. 2, 5; see Table 4 for a full list of values for the network).

193

194 *The nicotine withdrawal network is driven by cortical and extended amygdalar regions*

195 The nicotine network had 4,957 connections, the second most of all conditions, and
196 consisted of five modules and one brain region (interanterodorsal nucleus of the thalamus) that
197 was disconnected from the entire network. Overall, the nicotine network was relatively
198 interconnected between modules and had two large modules and three medium modules.

199 One of the large modules (module 1) contained midbrain (e.g., pedunculopontine nucleus
200 and periaqueductal gray), hindbrain (e.g., pons and pontine reticular nucleus), cortical (e.g.,
201 perirhinal area, posterior auditory area, ventral anterior cingulate temporal association areas, and
202 visceral area), and subcortical (claustrum) brain regions that had high WMDz. A separate set of
203 cortical (e.g., postsubiculum, lateral visual area, and gustatory areas), thalamic (e.g., anteroventral
204 nucleus of the thalamus and peripeduncular nucleus), hypothalamic (e.g., posterior periventricular
205 nucleus, supramammillary nucleus, and periventricular zone), and midbrain (e.g., midbrain

206 reticular nucleus, ventral tegmental area, and medial pretecal area) brain regions and a few others
207 that included the central amygdala and vestibular nuclei had high PC.

208 In the second large module (module 4), a set of sensory/cortical (e.g., primary
209 somatosensory area, lower limb, ventral agranular insular area [AIV], and primary motor area) and
210 hypothalamic (e.g., parastriatal nucleus, retrochiasmatic area, lateral preoptic area, medial preoptic
211 area, and zona incerta) brain regions had high WMDz. All of the same sensory/cortical and
212 hypothalamic regions had high PC and a number of other thalamic and sensory regions.
213 Additionally, the anterior amygdalar area (AAA) also showed both high WMDz and high PC.

214 One of the smaller modules (module 2) consisted of hippocampal (dentate gyrus) and
215 sensory/cortical (e.g., posterolateral visual area, anteromedial visual area, and subiculum [SUB])
216 regions, along with the nucleus of reuniens (RE) with high WMDz. The SUB and RE also had
217 high PC, along with other thalamic, hypothalamic, and midbrain regions.

218 In another smaller module (module 3), the precommissural nucleus (PRC), medial
219 habenula, and intergeniculate leaflet of the lateral geniculate complex (IGL) had high WMDz and
220 high PC. Other midbrain and thalamic regions also had high PC.

221 In the last small module (module 5), no regions reached the criterion for high WMDz, but
222 the orbitofrontal cortex (lateral and ventrolateral orbital area), bed nucleus of the stria terminalis,
223 and medial amygdalar nucleus were all in the top five values (WMDz = 0.64-0.67). However,
224 every region in this module, with the exception of the bed nucleus of the accessory olfactory tract,
225 reached the criterion for high PC (Fig. 2, 6; see Table 5 for a full list of values for the network).

226

227 **Discussion**

228 The present study used unbiased single-cell whole-brain imaging to identify changes in
229 brain functional architecture after withdrawal from chronic exposure to psychostimulants.
230 Withdrawal from psychostimulants resulted in a massive increase in neural coactivation that was
231 associated with a decrease in modularity with varying degrees of severity, depending on the
232 drug, compared with control mice. This decreased modularity resulted in the emergence of new
233 network architecture and organization of the brain. Using graph theory, we identified brain
234 regions that are most responsible for inter- and intramodular communication within each
235 network. Withdrawal from all of the psychostimulants that were tested in the present study
236 resulted in different network organization than the control network. The methamphetamine and
237 cocaine withdrawal networks closely resembled each other in structural organization, primarily
238 through thalamic motifs, whereas the nicotine withdrawal network shared some similarities with
239 the control network. These unbiased whole-brain analyses demonstrate that psychostimulant
240 withdrawal produces the drug-dependent remodeling of functional architecture of the brain and
241 suggest that decreased modularity of the brain functional network may be a central feature of
242 addiction.

243 We found that cocaine, methamphetamine, and nicotine withdrawal produced major
244 increases in coordinated activity throughout the brain compared with control mice. We further
245 found that withdrawal resulted in a decrease in modular structuring of the brain compared with
246 control mice (seven modules). The decrease in modularity was most evident for methamphetamine
247 withdrawal (three modules) and cocaine withdrawal (four modules), whereas nicotine withdrawal
248 showed a smaller reduction of modularity (five modules). Such reductions of modularity are found
249 in humans who suffer from dementia and traumatic brain injury and are associated with cognitive
250 deficits [28-33]. Changes in network structure/functional connectivity [10-13] and cognitive

251 function [34-36] have been observed after chronic drug use and withdrawal, suggesting that similar
252 mechanisms may be active between these different neural disorders.

253 We examined the components of individual modules within each network and found that
254 the control network was heavily driven by sensory and motor brain regions. This result confers
255 validity to our single-cell whole-brain network analysis approach for characterizing network
256 features because it fits with what might be expected from a normal, awake, behaving animal that
257 explores the environment and relies heavily on sensory/motor systems. Furthermore, the control
258 network was more interconnected between modules overall and contained several regions that
259 could be classified as hubs of each module that are critical for network function, based on high
260 intra- and intermodular connectivity. This suggests that the control brain may be more resilient to
261 the disruption of function because additional hub regions may compensate more easily in
262 response to such disruptions.

263 In the networks that were associated with withdrawal from psychostimulants, a shift was
264 observed from sensory/motor regions to more subcortical (e.g., amygdalar, thalamic,
265 hypothalamic, and midbrain) regions that drive the network. A similar effect was seen in
266 nonhuman primates after cocaine abstinence [37], and alterations of connectivity of the
267 somatosensory cortex are associated with smokers [38]. This may represent a shift from top-
268 down cortical network control [39] to bottom-up subcortical network control and may reflect the
269 greater influence of internal drives that are associated with negative affect during withdrawal in
270 controlling the whole-brain network [40]. This shift may be a major reason why drugs are so
271 addictive because higher cortical connectivity in humans may protect against relapse [41].

272 Given the modular organization of the different networks, both the control network and
273 nicotine network had a much higher incidence of intermodular connectivity, whereas the

274 methamphetamine and cocaine networks had only a small subset of brain regions that were
275 connected between different modules. Similar changes in neural activity, combined with
276 decreases in interconnectivity and network efficiency, have been observed in humans after
277 psychostimulant use [42-44]. The nicotine network was different from the methamphetamine and
278 cocaine networks and somewhat resembled a slightly altered control network. Similarities and
279 differences in network properties of the three different drugs are likely to be caused by
280 differences in receptor mechanisms and locations where each drug acts throughout the brain.
281 Indeed, both cocaine and methamphetamine target the same dopamine transporter, whereas
282 nicotine acts on nicotinic receptors [45-48]. These results suggest that single-cell whole-brain
283 imaging may be used as a fingerprint or “brainprint” to characterize novel compounds by
284 comparing whole-brain network changes to existing compounds.

285 The interanterodorsal nucleus of the thalamus was disconnected from the nicotine
286 network, suggesting that it may not be involved in controlling the withdrawal network, although
287 we cannot exclude the possibility that its disconnection may instead be a critical feature of
288 nicotine withdrawal. One of the larger modules in the nicotine network was driven by several
289 brain regions, two of which included the AAA and AIv, which have been suggested to be
290 associated with nicotine withdrawal in humans [49, 50]. The methamphetamine and cocaine
291 networks, although having distinctly different features, shared an overall motif of lower
292 modularity and being heavily driven by thalamic brain regions. This suggests that, in a
293 destabilized and less structured neural network, the thalamus becomes more critical to
294 controlling the whole-brain network. The thalamus is thought to play a major role in relaying
295 information, and the reliance of these networks on this group of regions suggests that the
296 thalamus is not simply a relay station but has greater importance in cognitive and emotional

297 function [51, 52]. Substantial evidence corroborates the importance of the thalamus in
298 psychostimulant addiction and withdrawal. In a rat model of cocaine self-administration, the
299 thalamus was found to be heavily involved in network function during acute abstinence, but
300 changes in the network disappeared after 2 weeks [22]. Interestingly, the thalamus in humans has
301 been shown to be hypoactive in cocaine abusers [53], and thalamic connectivity is predictive of
302 cocaine dependence [54] and altered in infants who are exposed to cocaine [55]. Although
303 network changes that are induced by acute withdrawal are reversed over time [22], prolonged use
304 may lead to more permanent restructuring of the brain, and major differences between the
305 nicotine and methamphetamine/cocaine networks may account for differences in the severity of
306 each drug after long-term use [35, 45, 56].

307 In the past 40 years, the addiction field has made tremendous progress by identifying
308 numerous brain regions that are dysregulated after psychostimulant exposure and contribute to
309 addiction-like behaviors [3-9]. Despite this vast knowledge, however, still unclear are the ways
310 in which these neuroadaptations, as a whole, lead to addiction. The identification of a final
311 common brain pathway that is responsible for addiction remains elusive. The present results
312 confirm that a substantial number of brain regions are affected by psychostimulant exposure and
313 suggest that the final common pathway that is responsible for addiction may not reside at the
314 level of brain regions or even single neural circuits. Instead, these results suggest that the main
315 common phenomenon that is observed among all three of these psychostimulants is decreased
316 modularity of whole-brain functional architecture, suggesting that the final common pathway
317 may reside at the whole-network level. This interpretation is consistent with the literature on the
318 modularity of complex systems, including the brain and mind, showing that lower modularity
319 reduces the capacity of the system to adapt to its environment [57]. Such a reduction of the

320 capacity to adapt to the environment is reminiscent of one cardinal symptom of addiction,
321 namely continued drug use despite adverse consequences.

322 In summary, the present study showed that withdrawal from psychostimulants results in
323 changes in neural network structure, including increases in coactivation among brain regions and
324 decreases in modularity. Psychostimulant withdrawal resulted in a shift from a sensory/motor-
325 driven network to a network that is highly driven by subcortical regions. We also found that
326 different psychostimulants do not produce the same neural networks, although methamphetamine
327 and cocaine shared similar properties. These findings shed light on alterations of brain function
328 that are caused by drug exposure and identify potential brain regions that warrant future study. The
329 present study demonstrates that psychostimulant withdrawal produces drug-dependent remodeling
330 of the functional architecture of the brain and suggests that decreased modularity of the brain
331 functional networks and not a specific set of brain regions may represent the final common
332 pathway that leads to addiction. These findings may prove critical to designing future treatment
333 approaches for drug abuse.

334

335 **Materials and Methods**

336 *Animals*

337 Male C57BL/6J mice were bred at The Scripps Research Institute. They were 20-30 g and
338 60 days old at the start of the experiment. The mice were maintained on a 12 h/12 h light/dark
339 cycle with *ad libitum* access to food and water. All of the procedures were conducted in strict
340 adherence to the National Institutes of Health Guide for the Care and Use of Laboratory Animals
341 and approved by The Scripps Research Institute Institutional Animal Care and Use Committee.

342

343 *Drugs*

344 The doses were 4 mg/kg/day for methamphetamine, 24 mg/kg/day for nicotine, and 60
345 mg/kg/day for cocaine. These doses were chosen based on previous studies that indicated
346 rewarding effects during use, resulting in withdrawal-like symptoms after the cessation of chronic
347 use [14-20]. Each drug was dissolved in saline, and the pH was adjusted to 7.4. The drugs were
348 loaded into osmotic minipumps (Alzet; model no. 1002). The minipumps sat overnight in saline
349 before insertion to ensure that drug delivery would begin immediately.

350

351 *Minipump implantation and removal*

352 The mice were split into four groups for the experiment: methamphetamine withdrawal
353 group ($n = 5$), nicotine withdrawal group ($n = 5$), cocaine withdrawal group ($n = 5$), and saline
354 control group ($n = 4$). Each mouse was surgically implanted with an osmotic minipump for
355 methamphetamine, nicotine, cocaine, and saline based on group assignment. The minipumps were
356 implanted in the lower back of each mouse under anesthesia. After brief recovery, the mice were
357 returned to their home cages. The mice remained in their home cages for 1 week to allow for
358 chronic infusion of the drug.

359 After 1 week, the minipumps were surgically removed under anesthesia to allow for drug
360 washout and withdrawal to begin. Mice in the nicotine, cocaine, and saline groups were perfused
361 8 h after removal of the minipumps. Mice in the methamphetamine group were perfused 12 h after
362 removal of the minipumps. These time points were chosen to represent an acute withdrawal period
363 from each drug (e.g., a minimum of 4 h without the drug present) and based on the half-life of each
364 drug in mice [58-62].

365

366 *Tissue collection*

367 The mice were deeply anesthetized and perfused with 15 ml of phosphate-buffered saline
368 (PBS) followed by 50 ml of 4% formaldehyde. The brains were postfixed in formaldehyde
369 overnight. The next day, the brains were washed for 30 min three times with PBS and transferred
370 to a PBS/0.1% azide solution at 4°C for 2-3 days before processing via iDISCO+.

371

372 *iDISCO+*

373 The iDISCO+ procedure was performed as reported by Renier et al. [63, 64].

374

375 *Immunostaining*

376 Fixed samples were washed in 20% methanol (in double-distilled H₂O) for 1 h, 40%
377 methanol for 1 h, 60% methanol for 1 h, 80% methanol for 1 h, and 100% methanol for 1 h
378 twice. The samples were then precleared with overnight incubation in 33% methanol/66%
379 dichloromethane (DCM; Sigma, catalog no. 270997-12X100ML). The next day, the samples
380 were bleached with 5% H₂O₂ (1 volume of 30% H₂O₂ for 5 volumes of methanol, ice cold) at
381 4°C overnight. After bleaching, the samples were slowly re-equilibrated at room temperature and
382 rehydrated in 80% methanol in double-distilled H₂O for 1 h, 60% methanol for 1 h, 40%
383 methanol for 1 h, 20% methanol for 1 h, PBS for 1 h, and PBS/0.2% TritonX-100 for 1 h twice.
384 The samples were then incubated in PBS/0.2% TritonX-100/20% dimethylsulfoxide
385 (DMSO)/0.3 M glycine at 37°C for 2 days and then blocked in PBS/0.2% TritonX-100/10%
386 DMSO/6% donkey serum at 37°C for 2 days. The samples were then incubated in rabbit anti c-
387 fos (1:2000; Synaptic Systems catalog number 226 003) in PBS-0.2% Tween with 10 µg/ml
388 heparin (PTwH)/5% DMSO/3% donkey serum at 37°C for 7 days. The samples were then

389 washed in PTwH for 24 h (five changes of the PTwH solution over that time) and incubated in
390 donkey anti-rabbit Alexa647 (1:500; Invitrogen, catalog no. A31573) in PTwH/3% donkey
391 serum at 37°C for 7 days. The samples were finally washed in PTwH for 1 day before clearing
392 and imaging.

393

394 *Sample clearing*

395 Immunolabeled brains were cleared using the procedure of Reiner et al. (2016) [63]. The
396 samples were dehydrated in 20% methanol in double-distilled H₂O for 1 h, 40% methanol for 1
397 h, 60% methanol for 1 h, 80% methanol for 1 h, 100% methanol for 1 h, and 100% methanol
398 again overnight. The next day, the samples were incubated for 3 h in 33% methanol/66% DCM
399 until they sank to the bottom of the incubation tube. The methanol was then washed for 20 min
400 twice in 100% DCM. Finally, the samples were incubated in dibenzyl ether (DBE; Sigma,
401 catalog no. 108014-1KG) until clear and then stored in DBE at room temperature until imaged.

402

403 *Image acquisition*

404 Left hemispheres of cleared samples were imaged in the sagittal orientation (right lateral
405 side up) on a light-sheet microscope (Ultramicroscope II, LaVision Biotec) equipped with an
406 sCMOS camera (Andor Neo) and 2×/0.5 objective lens (MVPLAPO 2×) equipped with a 6 mm
407 working distance dipping cap. Inspector Microscope controller v144 software was used. The
408 microscope was equipped with an NKT Photonics SuperK EXTREME EXW-12 white light laser
409 with three fixed light sheet generating lenses on each side. Scans were made at 0.8×
410 magnification (1.6× effective magnification) with a light sheet numerical aperture of 0.148.
411 Excitation filters of 480/30, 560/40, and 630/30 nm were used. Emission filters of 525/50,

412 595/40, and 680/30 nm were used. The samples were scanned with a step size of 3 μm using
413 dynamic horizontal scanning from one side (the right) for the 560 and 630 nm channels (20
414 acquisitions per plane with 240 ms exposure, combined into one image using the horizontal
415 adaptive algorithm) and without horizontal scanning for the 480 nm channel using two-sided
416 illumination (100 ms exposure for each side, combined into one image using the blending
417 algorithm). To accelerate acquisition, both channels were acquired in two separate scans. To
418 account for micro-movements of the samples that may occur between scans, three-dimensional
419 image affine registration was performed to align both channels using ClearMap [63].

420

421 *Data analysis*

422 *Identification of activated brain regions.* Images that were acquired from the light-sheet
423 microscope were analyzed from the end of the olfactory bulbs (the olfactory bulbs were not
424 included in the analysis) to the beginning of the hindbrain and cerebellum. Counts of Fos-
425 positive nuclei from each sample were identified for each brain region using ClearMap [63].
426 ClearMap uses autofluorescence that is acquired in the 488 nm channel to align the brain to the
427 Allen Mouse Brain Atlas [65] and then registers Fos counts to regions that are annotated by the
428 atlas. The data were normalized to a \log_{10} value to reduce variability and bring brain regions with
429 high numbers (e.g., thousands) and low numbers (e.g., tens to hundreds) of Fos counts to a
430 similar scale.

431 *Identification of co-activation within individual networks.* Separate inter-regional Pearson
432 correlations were then calculated using Statistica software (Tibco) across animals in the saline,
433 cocaine, methamphetamine, and nicotine groups to compare the \log_{10} Fos data from each brain

434 region to each of the other brain regions. See Table 1 for a list of brain regions, their
435 abbreviations, and their Allen atlas grouping.

436

437 *Hierarchical clustering*

438 Previous rat and mouse studies that examined functional connectivity used 5-8 animals
439 [21, 22]. The number of samples that are examined in functional connectivity studies is the
440 number of potential connections (i.e., 178 total brain regions all connecting with each other for
441 each treatment). Furthermore, hierarchical clustering organizes brain regions into modules by
442 grouping regions that show a similar coactivation profile across all other brain regions. Thus,
443 more total connections minimize the effect that an inaccurate brain region-to-brain region
444 connection has on network organization and overall network structure.

445 Inter-regional Fos correlations were then used to calculate complete Euclidean distances
446 between each pair of brain regions in each group of mice. The distance matrices were then
447 hierarchically clustered using R Studio software by both row and column using the complete
448 method to identify modules of coactivation within each treatment group. The hierarchical cluster
449 dendrograms were trimmed at half the height of each given tree to split the dendrogram into
450 specific modules. The result of a decrease in modularity that is attributable to psychostimulant
451 use was consistent across multiple tree-cutting thresholds (Fig. 1B).

452

453 *Graph theory identification of functional networks*

454 We used a graph theory-based approach to identify the functional neural networks that
455 were associated with each treatment condition. Graph theory is a branch of mathematics that is
456 used to analyze complex networks, such as social, financial, protein, and neural networks [21,

457 66-77]. Using graph theory, functional networks can be delineated, and key brain regions of the
458 network can be identified [21, 69, 78, 79].

459 Previous studies of regional connectivity profiles in Fos coactivation networks have
460 focused on global measures of connectivity (e.g., degree) [21]. However, in correlation-based
461 networks, these measures can be strongly influenced by the size of the subnetwork (module) in
462 which a node participates [80]. For the graph theory analyses, we were interested in regional
463 properties and not module size *per se*. Thus, module structure needs to be considered when
464 examining the role that each region plays in the network. To accomplish this, we utilized two
465 widely used centrality metrics that were designed for application to modular systems. The
466 WMDz indexes the relative importance of a region within its own module (e.g., intramodule
467 connectivity), and the PC indexes the extent to which a region connects diversely to multiple
468 modules (e.g., intermodule connectivity) [27].

469 We used the Pearson correlation values that were calculated for the brain regions from
470 each treatment. Prior to plotting and calculating regional connectivity metrics, the network was
471 thresholded to remove any edges that were weaker than $R = 0.75$. As such, visualization and
472 graph theory analyses were performed using only edges with positive weights. Regional
473 connectivity metrics (PC and WMDz) were calculated as originally defined by Guimerà and
474 Amaral (2005) [27], modified for application to networks with weighted edges. PC and WMDz
475 were calculated using a customized version of the bctpy Python package
476 (<https://github.com/aestrivex/bctpy>), which is derived from the MATLAB implementation of
477 Brain Connectivity Toolbox [78].

478 For WMDz, let k_i (within-module degree) be the summed weight of all edges between
479 region i and other regions in module s_i . Then, \bar{k}_{s_i} is the average within-module degree of all

480 regions in module s_i , and $\sigma_{k_{s_i}}$ is the standard deviation of those values. The Z-scored version of
481 within-module degree (WMDz) is then defined as:

482
$$WMDz = \frac{k_i - \bar{k}_{s_i}}{\sigma_{k_{s_i}}}$$

483 This provides a measure of the extent to which each region is connected to other regions in the
484 same module.

485 For PC, let k_{is} (between-module degree) be the summed weight of all edges between
486 region i and regions in module s , and let k_i (total degree) be the summed weight of all edges
487 between region i and all other regions in the network. The PC of each region is then defined as:

488
$$P_i = 1 - \sum_{s=1}^{N_M} \left(\frac{k_{is}}{k_i} \right)^2$$

489 This provides a measure of the extent to which the connections of a region are distributed mostly
490 within its own module (PC approaching 0) or distributed evenly among all modules (PC
491 approaching 1).

492 A high PC was considered ≥ 0.30 , and a high WMDz was considered ≥ 0.80 . Previous
493 studies have used ranges of ≥ 0.30 - 0.80 for high PC and ≥ 1.5 - 2.5 for high WMDz [27, 77].

494 Because of differences in the sizes/types of networks that were examined and the methods that
495 were used (e.g., Fos vs. functional magnetic resonance imaging), we adjusted the range for
496 consideration as having high PC and WMDz accordingly.

497 Network visualization was performed using a combination of Gephi 0.9.2 software [81]
498 and Adobe Illustrator software. Nodes were positioned using the Force Atlas 2 algorithm [82]
499 with a handful of nodes that were repositioned manually for better visual organization.

500

501

502

503

504 **References**

- 505 1. Balfour, D.J.K., *The psychobiology of nicotine dependence*. European Respiratory
506 Review, 2008. **17**: p. 172-181.
- 507 2. Phillips, K.A., D.H. Epstein, and K.L. Preston, *Psychostimulant addiction treatment*.
508 Neuropharmacology, 2014. **87**: p. 150-60.
- 509 3. Kalivas, P.W. and K. McFarland, *Brain circuitry and the reinstatement of cocaine-*
510 *seeking behavior*. Psychopharmacology (Berl), 2003. **168**(1-2): p. 44-56.
- 511 4. Bobadilla, A.C., et al., *Corticostriatal plasticity, neuronal ensembles, and regulation of*
512 *drug-seeking behavior*. Prog Brain Res, 2017. **235**: p. 93-112.
- 513 5. Kalivas, P.W., *Cocaine and amphetamine-like psychostimulants: neurocircuitry and*
514 *glutamate neuroplasticity*. Dialogues Clin Neurosci, 2007. **9**(4): p. 389-97.
- 515 6. Everitt, B.J., et al., *Review. Neural mechanisms underlying the vulnerability to develop*
516 *compulsive drug-seeking habits and addiction*. Philos Trans R Soc Lond B Biol Sci,
517 2008. **363**(1507): p. 3125-35.
- 518 7. Jedynak, J.P., C.M. Cameron, and T.E. Robinson, *Repeated methamphetamine*
519 *administration differentially alters fos expression in caudate-putamen patch and matrix*
520 *compartments and nucleus accumbens*. PLoS One, 2012. **7**(4): p. e34227.
- 521 8. Robinson, T.E. and B. Kolb, *Structural plasticity associated with exposure to drugs of*
522 *abuse*. Neuropharmacology, 2004. **47 Suppl 1**: p. 33-46.
- 523 9. Koob, G.F. and N.D. Volkow, *Neurobiology of addiction: a neurocircuitry analysis*.
524 Lancet Psychiatry, 2016. **3**(8): p. 760-73.
- 525 10. Tomasi, D., et al., *Disrupted functional connectivity with dopaminergic midbrain in*
526 *cocaine abusers*. PLoS One, 2010. **5**(5): p. e10815.

- 527 11. Konova, A.B., et al., *Effects of methylphenidate on resting-state functional connectivity*
528 *of the mesocorticolimbic dopamine pathways in cocaine addiction*. JAMA Psychiatry,
529 2013. **70**(8): p. 857-68.
- 530 12. Ma, L., et al., *Effect of cocaine dependence on brain connections: clinical implications*.
531 Expert Rev Neurother, 2015. **15**(11): p. 1307-19.
- 532 13. Konova, A.B., et al., *Effects of chronic and acute stimulants on brain functional*
533 *connectivity hubs*. Brain Res, 2015. **1628**(Pt A): p. 147-56.
- 534 14. Johnson, P.M., J.A. Hollander, and P.J. Kenny, *Decreased brain reward function during*
535 *nicotine withdrawal in C57BL6 mice: evidence from intracranial self-stimulation (ICSS)*
536 *studies*. Pharmacol Biochem Behav, 2008. **90**(3): p. 409-15.
- 537 15. Zhu, J., et al., *Sodium butyrate modulates a methamphetamine-induced conditioned place*
538 *preference*. J Neurosci Res, 2017. **95**(4): p. 1044-1052.
- 539 16. Eisener-Dorman, A.F., L. Grabowski-Boase, and L.M. Tarantino, *Cocaine locomotor*
540 *activation, sensitization and place preference in six inbred strains of mice*. Behav Brain
541 Funct, 2011. **7**: p. 29.
- 542 17. Stoker, A.K., B. Olivier, and A. Markou, *Involvement of metabotropic glutamate*
543 *receptor 5 in brain reward deficits associated with cocaine and nicotine withdrawal and*
544 *somatic signs of nicotine withdrawal*. Psychopharmacology (Berl), 2012. **221**(2): p. 317-
545 27.
- 546 18. Fish, E.W., et al., *Alcohol, cocaine, and brain stimulation-reward in C57Bl6/J and*
547 *DBA2/J mice*. Alcohol Clin Exp Res, 2010. **34**(1): p. 81-9.

- 548 19. Stoker, A.K. and A. Markou, *Withdrawal from chronic cocaine administration induces*
549 *deficits in brain reward function in C57BL/6J mice*. Behav Brain Res, 2011. **223**(1): p.
550 176-81.
- 551 20. Tracy, M.E., M.L. Banks, and K.L. Shelton, *Negative allosteric modulation of GABAA*
552 *receptors inhibits facilitation of brain stimulation reward by drugs of abuse in C57BL/6/J*
553 *mice*. Psychopharmacology (Berl), 2016. **233**(4): p. 715-25.
- 554 21. Wheeler, A.L., et al., *Identification of a functional connectome for long-term fear*
555 *memory in mice*. PLoS Comput Biol, 2013. **9**(1): p. e1002853.
- 556 22. Orsini, C.A., et al., *Functional Connectivity of Chronic Cocaine Use Reveals Progressive*
557 *Neuroadaptations in Neocortical, Striatal, and Limbic Networks*. eNeuro, 2018. **5**(4).
- 558 23. Giove, F., et al., *Images-based suppression of unwanted global signals in resting-state*
559 *functional connectivity studies*. Magn Reson Imaging, 2009. **27**(8): p. 1058-64.
- 560 24. Chen, G., et al., *Negative functional connectivity and its dependence on the shortest path*
561 *length of positive network in the resting-state human brain*. Brain Connect, 2011. **1**(3): p.
562 195-206.
- 563 25. Murphy, K., et al., *The impact of global signal regression on resting state correlations:*
564 *are anti-correlated networks introduced?* Neuroimage, 2009. **44**(3): p. 893-905.
- 565 26. Meunier, D., et al., *Age-related changes in modular organization of human brain*
566 *functional networks*. Neuroimage, 2009. **44**(3): p. 715-23.
- 567 27. Guimera, R. and L.A. Nunes Amaral, *Functional cartography of complex metabolic*
568 *networks*. Nature, 2005. **433**(7028): p. 895-900.
- 569 28. Gallen, C.L., et al., *Modular Brain Network Organization Predicts Response to Cognitive*
570 *Training in Older Adults*. PLoS One, 2016. **11**(12): p. e0169015.

- 571 29. Bertolero, M.A., et al., *A mechanistic model of connector hubs, modularity and cognition.*
572 *Nature Human Behaviour*, 2018.
- 573 30. de Haan, W., et al., *Disrupted modular brain dynamics reflect cognitive dysfunction in*
574 *Alzheimer's disease.* *Neuroimage*, 2012. **59**(4): p. 3085-93.
- 575 31. Brier, M.R., et al., *Functional connectivity and graph theory in preclinical Alzheimer's*
576 *disease.* *Neurobiol Aging*, 2014. **35**(4): p. 757-68.
- 577 32. Sporns, O. and R.F. Betzel, *Modular Brain Networks.* *Annu Rev Psychol*, 2016. **67**: p.
578 613-40.
- 579 33. Arnemann, K.L., et al., *Functional brain network modularity predicts response to*
580 *cognitive training after brain injury.* *Neurology*, 2015. **84**(15): p. 1568-74.
- 581 34. Sabrini, S., et al., *Methamphetamine use and cognitive function: A systematic review of*
582 *neuroimaging research.* *Drug Alcohol Depend*, 2019. **194**: p. 75-87.
- 583 35. Spronk, D.B., et al., *Characterizing the cognitive effects of cocaine: a comprehensive*
584 *review.* *Neurosci Biobehav Rev*, 2013. **37**(8): p. 1838-59.
- 585 36. Ashare, R.L., M. Falcone, and C. Lerman, *Cognitive function during nicotine*
586 *withdrawal: Implications for nicotine dependence treatment.* *Neuropharmacology*, 2014.
587 **76 Pt B**: p. 581-91.
- 588 37. Murnane, K.S., et al., *Functional connectivity in frontal-striatal brain networks and*
589 *cocaine self-administration in female rhesus monkeys.* *Psychopharmacology (Berl)*, 2015.
590 **232**(4): p. 745-54.
- 591 38. Claus, E.D. and C.R. Weywadt, *Resting-State Connectivity in Former, Current and Never*
592 *Smokers.* *Nicotine Tob Res*, 2018.

- 593 39. Gilbert, C.D. and M. Sigman, *Brain states: top-down influences in sensory processing*.
594 *Neuron*, 2007. **54**(5): p. 677-96.
- 595 40. Koob, G.F., *The dark side of emotion: the addiction perspective*. *Eur J Pharmacol*, 2015.
596 **753**: p. 73-87.
- 597 41. McHugh, M.J., et al., *Executive control network connectivity strength protects against*
598 *relapse to cocaine use*. *Addict Biol*, 2017. **22**(6): p. 1790-1801.
- 599 42. Ahmadlou, M., et al., *Global organization of functional brain connectivity in*
600 *methamphetamine abusers*. *Clin Neurophysiol*, 2013. **124**(6): p. 1122-31.
- 601 43. Wang, Z., et al., *A hyper-connected but less efficient small-world network in the*
602 *substance-dependent brain*. *Drug Alcohol Depend*, 2015. **152**: p. 102-8.
- 603 44. Liang, X., et al., *Interactions between the salience and default-mode networks are*
604 *disrupted in cocaine addiction*. *J Neurosci*, 2015. **35**(21): p. 8081-90.
- 605 45. Nestler, E.J., *The neurobiology of cocaine addiction*. *Sci Pract Perspect*, 2005. **3**(1): p. 4-
606 10.
- 607 46. Rothman, R.B. and M.H. Baumann, *Monoamine transporters and psychostimulant drugs*.
608 *Eur J Pharmacol*, 2003. **479**(1-3): p. 23-40.
- 609 47. Sulzer, D., et al., *Mechanisms of neurotransmitter release by amphetamines: a review*.
610 *Prog Neurobiol*, 2005. **75**(6): p. 406-33.
- 611 48. D'Souza, M.S. and A. Markou, *Neuronal mechanisms underlying development of nicotine*
612 *dependence: implications for novel smoking-cessation treatments*. *Addict Sci Clin Pract*,
613 2011. **6**(1): p. 4-16.

- 614 49. Sutherland, M.T., et al., *Down-regulation of amygdala and insula functional circuits by*
615 *varenicline and nicotine in abstinent cigarette smokers*. Biol Psychiatry, 2013. **74**(7): p.
616 538-46.
- 617 50. Naqvi, N.H., et al., *Damage to the insula disrupts addiction to cigarette smoking*.
618 Science, 2007. **315**(5811): p. 531-4.
- 619 51. Ahissar, E. and T. Oram, *Thalamic relay or cortico-thalamic processing? Old question,*
620 *new answers*. Cereb Cortex, 2015. **25**(4): p. 845-8.
- 621 52. Sherman, S.M., *The thalamus is more than just a relay*. Curr Opin Neurobiol, 2007.
622 **17**(4): p. 417-22.
- 623 53. Tomasi, D., et al., *Thalamo-cortical dysfunction in cocaine abusers: implications in*
624 *attention and perception*. Psychiatry Res, 2007. **155**(3): p. 189-201.
- 625 54. Zhang, S., et al., *Cocaine dependence and thalamic functional connectivity: a*
626 *multivariate pattern analysis*. Neuroimage Clin, 2016. **12**: p. 348-58.
- 627 55. Salzwedel, A.P., et al., *Thalamocortical functional connectivity and behavioral*
628 *disruptions in neonates with prenatal cocaine exposure*. Neurotoxicol Teratol, 2016. **56**:
629 p. 16-25.
- 630 56. Grant, K.M., et al., *Methamphetamine-associated psychosis*. J Neuroimmune Pharmacol,
631 2012. **7**(1): p. 113-39.
- 632 57. Kashtan, N. and U. Alon, *Spontaneous evolution of modularity and network motifs*. Proc
633 Natl Acad Sci U S A, 2005. **102**(39): p. 13773-8.
- 634 58. Siu, E.C. and R.F. Tyndale, *Characterization and comparison of nicotine and cotinine*
635 *metabolism in vitro and in vivo in DBA/2 and C57BL/6 mice*. Mol Pharmacol, 2007.
636 **71**(3): p. 826-34.

- 637 59. Benuck, M., A. Lajtha, and M.E. Reith, *Pharmacokinetics of systemically administered*
638 *cocaine and locomotor stimulation in mice*. J Pharmacol Exp Ther, 1987. **243**(1): p. 144-
639 9.
- 640 60. Shabani, S., et al., *Profound reduction in sensitivity to the aversive effects of*
641 *methamphetamine in mice bred for high methamphetamine intake*. Neuropharmacology,
642 2012. **62**(2): p. 1134-41.
- 643 61. Cho, A.K., et al., *Relevance of pharmacokinetic parameters in animal models of*
644 *methamphetamine abuse*. Synapse, 2001. **39**(2): p. 161-6.
- 645 62. Norman, A.B., et al., *A chimeric human/murine anticocaine monoclonal antibody inhibits*
646 *the distribution of cocaine to the brain in mice*. J Pharmacol Exp Ther, 2007. **320**(1): p.
647 145-53.
- 648 63. Renier, N., et al., *Mapping of Brain Activity by Automated Volume Analysis of Immediate*
649 *Early Genes*. Cell, 2016. **165**(7): p. 1789-802.
- 650 64. Renier, N., et al., *iDISCO: a simple, rapid method to immunolabel large tissue samples*
651 *for volume imaging*. Cell, 2014. **159**(4): p. 896-910.
- 652 65. *Allen Mouse Brain Atlas*. 2004; Available from: <http://mouse.brain-map.org/>.
- 653 66. Jeong, H., et al., *Lethality and centrality in protein networks*. Nature, 2001. **411**(6833): p.
654 41-2.
- 655 67. Barabasi, A.L., *Scale-free networks: a decade and beyond*. Science, 2009. **325**(5939): p.
656 412-3.
- 657 68. Babu, M., et al., *Interaction landscape of membrane-protein complexes in*
658 *Saccharomyces cerevisiae*. Nature, 2012. **489**(7417): p. 585-9.

- 659 69. Vetere, G., et al., *Chemogenetic Interrogation of a Brain-wide Fear Memory Network in*
660 *Mice*. Neuron, 2017. **94**(2): p. 363-374 e4.
- 661 70. Jarrell, T.A., et al., *The connectome of a decision-making neural network*. Science, 2012.
662 **337**(6093): p. 437-44.
- 663 71. Oh, S.W., et al., *A mesoscale connectome of the mouse brain*. Nature, 2014. **508**(7495):
664 p. 207-14.
- 665 72. Varshney, L.R., et al., *Structural properties of the Caenorhabditis elegans neuronal*
666 *network*. PLoS Comput Biol, 2011. **7**(2): p. e1001066.
- 667 73. Markov, N.T., et al., *A weighted and directed interareal connectivity matrix for macaque*
668 *cerebral cortex*. Cereb Cortex, 2014. **24**(1): p. 17-36.
- 669 74. Chiang, A.S., et al., *Three-dimensional reconstruction of brain-wide wiring networks in*
670 *Drosophila at single-cell resolution*. Curr Biol, 2011. **21**(1): p. 1-11.
- 671 75. Bullmore, E. and O. Sporns, *The economy of brain network organization*. Nat Rev
672 Neurosci, 2012. **13**(5): p. 336-49.
- 673 76. Bargmann, C.I. and E. Marder, *From the connectome to brain function*. Nat Methods,
674 2013. **10**(6): p. 483-90.
- 675 77. Cohen, J.R. and M. D'Esposito, *The Segregation and Integration of Distinct Brain*
676 *Networks and Their Relationship to Cognition*. J Neurosci, 2016. **36**(48): p. 12083-
677 12094.
- 678 78. Rubinov, M. and O. Sporns, *Complex network measures of brain connectivity: uses and*
679 *interpretations*. Neuroimage, 2010. **52**(3): p. 1059-69.
- 680 79. Sporns, O., C.J. Honey, and R. Kotter, *Identification and classification of hubs in brain*
681 *networks*. PLoS One, 2007. **2**(10): p. e1049.

- 682 80. Power, J.D., et al., *Evidence for hubs in human functional brain networks*. Neuron, 2013.
683 **79**(4): p. 798-813.
- 684 81. Bastian, M., S. Heymann, and M. Jacomy, *Gephi: an open source software for exploring*
685 *and manipulating networks.*, in *International AAAI Conference on Weblogs and Social*
686 *Media*. 2009.
- 687 82. Jacomy, M., et al., *ForceAtlas2, a continuous graph layout algorithm for handy network*
688 *visualization designed for the Gephi software*. PLoS One, 2014. **9**(6): p. e98679.
- 689
690
691

692 **Figure Legends**

693

694 **Figure 1. A.** Experimental design. Mice were surgically implanted with an osmotic minipump that
695 contained either saline or a psychostimulant (60 mg/kg/day cocaine, 4 mg/kg/day
696 methamphetamine, or 24 mg/kg/day nicotine). They were then returned to their home cage for 1
697 week. After 1 week, the minipumps were surgically removed, and the mice were returned to their
698 home cage until brain tissue was collected 8 h later (saline, cocaine, nicotine) or 12 h later
699 (methamphetamine). Brains were then processed for whole-brain Fos immunohistochemistry and
700 clearing via iDISCO+ and then imaged on a light-sheet microscope. Fos values were detected and
701 registered to the Allen Brain Atlas using ClearMap [63]. Pearson correlations were then calculated
702 to determine functional coactivation among brain regions. Brain regions were then grouped into
703 modules based on their coactivation patterns through hierarchical clustering. Graph theory
704 analyses was then performed to identify brain regions that are heavily involved in intra- and
705 intermodular connectivity. **B-F.** Hierarchical clustering of complete Euclidean distance matrices
706 for each treatment. Modules were determined by cutting each dendrogram at half of the maximal
707 tree height. **B.** Number of modules in each treatment condition after cutting the hierarchical
708 clustered dendrogram at different percentages of tree height. In all cases (except at extreme cutoff
709 values; e.g., 90-100%), the psychostimulant networks showed lower modularity compared with
710 the control network. **C.** Relative distance of each brain region relative to the others that were
711 examined in control mice. In control mice, seven distinct modules of coactivation were identified.
712 **D.** Relative distance of each brain region relative to the others that were examined in cocaine mice.
713 In cocaine mice, four distinct modules of coactivation were identified. **E.** Relative distance of each
714 brain region relative to the others that were examined in methamphetamine mice. In

715 methamphetamine mice, three distinct modules of coactivation were identified. **F.** Relative
716 distance of each brain region relative to the others that were examined in nicotine mice. In nicotine
717 mice, five distinct modules of coactivation were identified. For all distance matrices, each module
718 is boxed in purple. For the individual brain regions that are listed in panels C-F, see Table 6.

719
720 **Figure 2.** Intramodular (WMDz) and intermodular (PC) network features of each treatment. A
721 high PC was considered ≥ 0.30 , and a high WMDz was considered ≥ 0.80 . **A.** Highlights of several
722 regions with high PC in each module of each network network (see Table 1 for names of
723 abbreviations). **B.** Highlights of several regions with high WMDz (red = higher, blue = lower) in
724 each module of each network. Note that the WMDz color intensity is only relative to the other
725 regions within the same network and not other networks (see Table 1 for names of abbreviations).
726 **C.** Total number of brain regions that accounted for high PC, high WMDz, or both in each network.
727 The control and nicotine networks showed much greater intermodular connectivity and a greater
728 number of regions with both high intermodular and intramodular connectivity. The cocaine and
729 methamphetamine networks showed higher levels of intramodular connectivity and a low number
730 of regions with intermodular connectivity.

731
732 **Figure 3.** Neural network of control mice thresholded to 0.75R. Nodes/brain regions of the
733 network are represented by circles. The size of the node represents the participation coefficient
734 (smaller = lower PC; larger = higher PC). The internal color of each circle represents the within-
735 module degree Z-score (dark blue = lowest; dark red = highest). The color of the modules that are
736 identified in Fig. 1C are represented by different colored edges. See figure key for examples of
737 each representative component of the figure.

738

739 **Figure 4.** Neural network of cocaine mice during withdrawal thresholded to 0.75R. Nodes/brain
740 regions of the network are represented by circles. The size of the node represents the participation
741 coefficient (smaller = lower PC; larger = higher PC). The internal color of each circle represents
742 the within-module degree Z-score (dark blue = lowest; dark red = highest). The color of the
743 modules that are identified in Fig. 1D are represented by different colored edges. See figure key
744 for examples of each representative component of the figure.

745

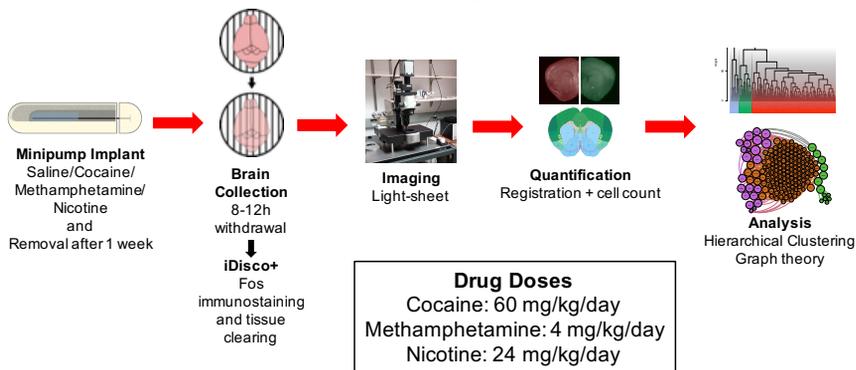
746 **Figure 5.** Neural network of methamphetamine mice during withdrawal thresholded to 0.75R.
747 Nodes/brain regions of the network are represented by circles. The size of the node represents the
748 participation coefficient (smaller = lower PC; larger = higher PC). The internal color of each circle
749 represents the within-module degree Z-score (dark blue = lowest; dark red = highest). The color
750 of the modules that are identified in Fig. 1E are represented by different colored edges. See figure
751 key for examples of each representative component of the figure.

752

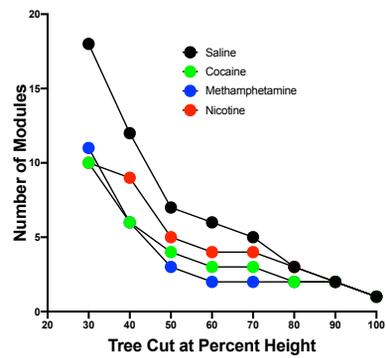
753 **Figure 6.** Neural network of nicotine mice during withdrawal thresholded to 0.75R. Nodes/brain
754 regions of the network are represented by circles. The size of the node represents the participation
755 coefficient (smaller = lower PC; larger = higher PC). The internal color of each circle represents
756 the within-module degree Z-score (dark blue = lowest; dark red = highest). The color of the
757 modules that are identified in Fig. 1F are represented by different colored edges. See figure key
758 for examples of each representative component of the figure.

759

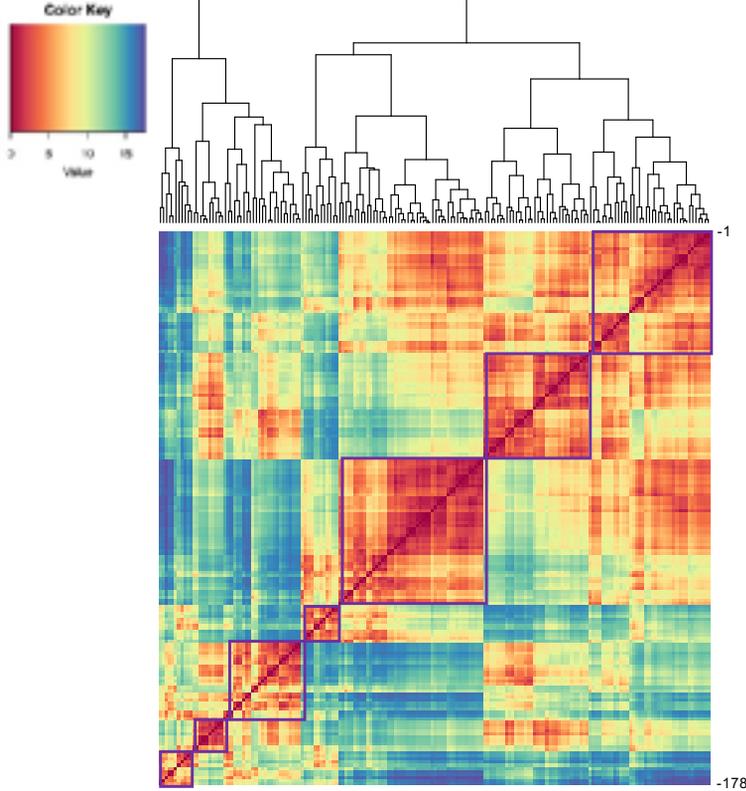
A. Experimental Design



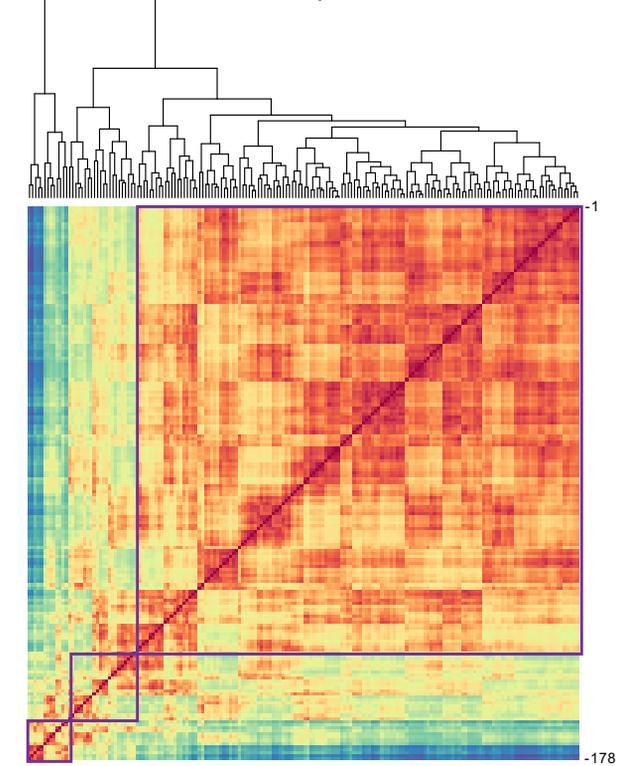
B. Modules



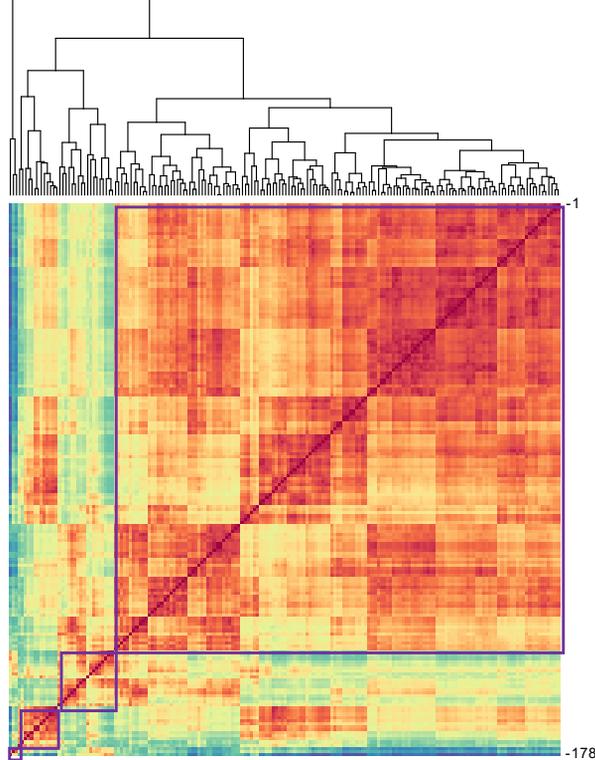
C. Saline



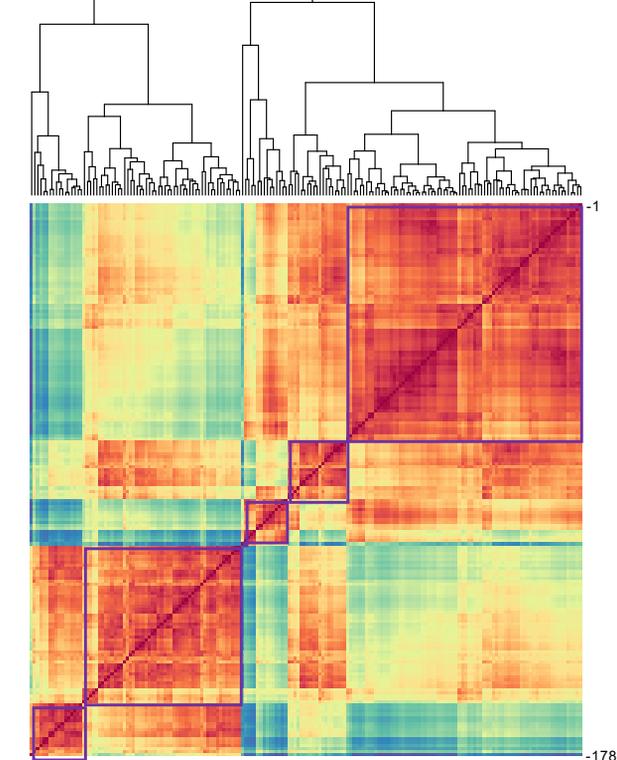
E. Methamphetamine



D. Cocaine



F. Nicotine



Midbrain-Amygdalo-Hypothalamic

Sensory-Hippocampal

Cerebellar-Thalamic

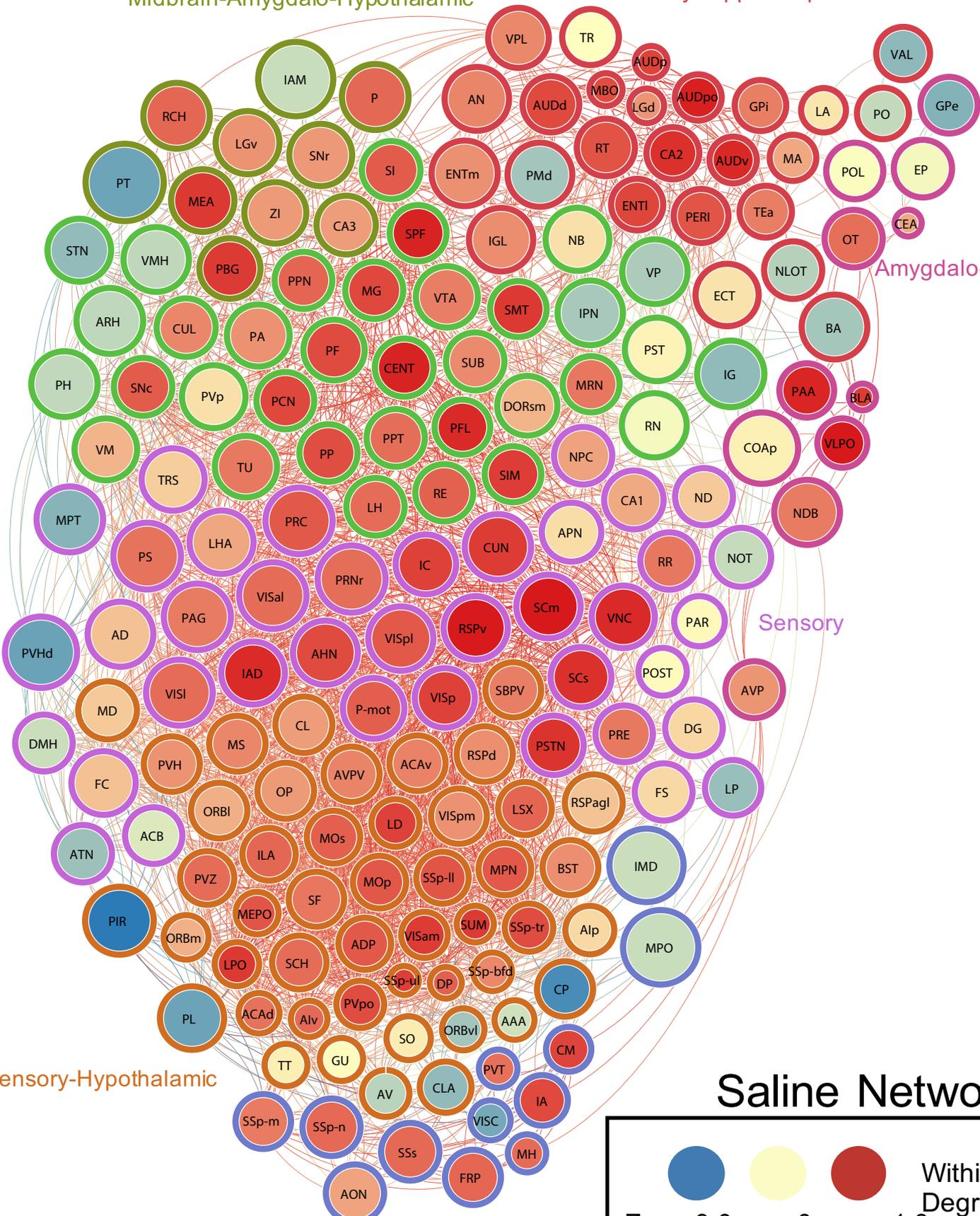
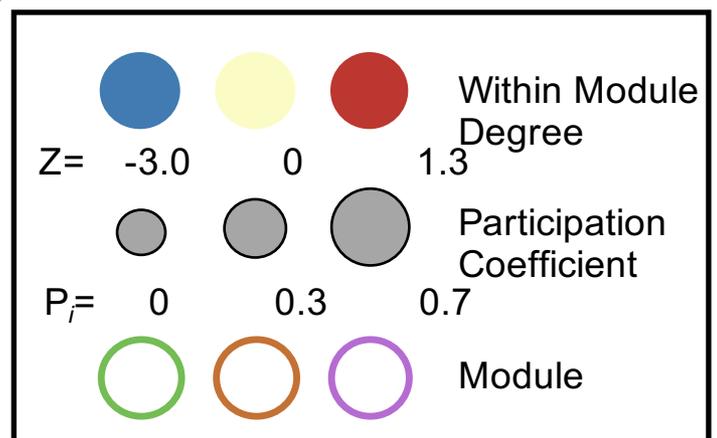
Amygdalo-Hypothalamic

Sensory

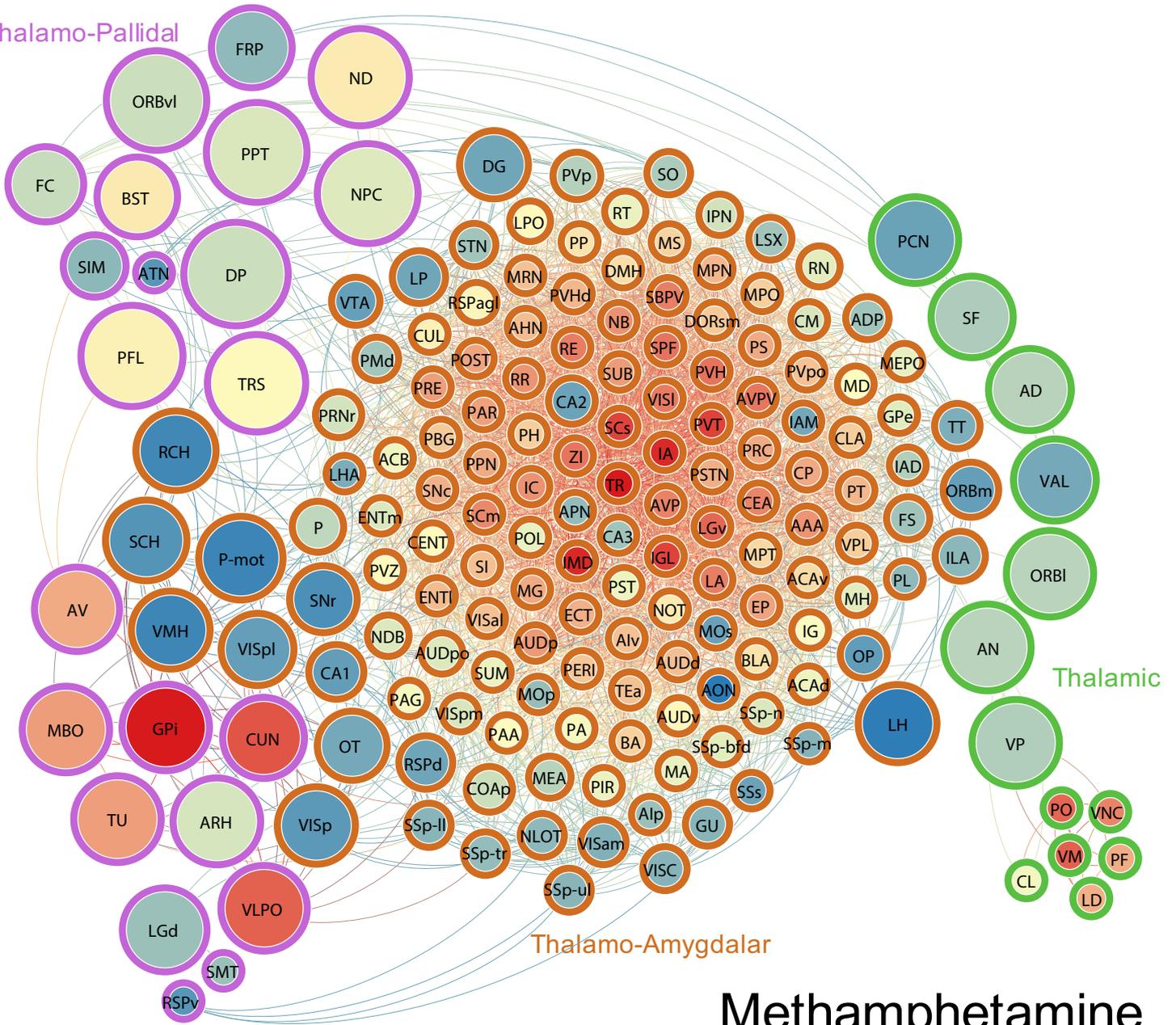
Sensory-Hypothalamic

Amygdalo-Thalamic

Saline Network



Hypothalamo-Pallidal



Methamphetamine Network

