

Distinct neurogenetic mechanisms establish the same chemosensory valence state at different life stages in Caenorhabditis elegans

Navonil Banerjee (D),^{1,2} Elisa J. Rojas Palato,¹ Pei-Yin Shih,^{3,4,5} Paul W. Sternberg (D),³ Elissa A. Hallem (D),^{1,2,*}

¹Department of Microbiology, Immunology, and Molecular Genetics, University of California, Los Angeles, CA 90095, USA

²Molecular Biology Institute, University of California, Los Angeles, CA 90095, USA

³Division of Biology and Biological Engineering, California Institute of Technology, Pasadena, CA 91125, USA

⁴Present address: Department of Ecology, Evolution and Environmental Biology, Columbia University, NewYork, NY 10027, USA

⁵Present address: Zuckerman Mind, Brain, Behavior Institute, Columbia University, NewYork, NY 10027, USA

*Corresponding author: Department of Microbiology, Immunology, and Molecular Genetics, University of California, Los Angeles, 237 BSRB, 615 Charles E. Young Dr. S., Los Angeles, CA 90095, USA. Email: ehallem@ucla.edu

An animal's preference for many chemosensory cues remains constant despite dramatic changes in the animal's internal state. The mechanisms that maintain chemosensory preference across different physiological contexts remain poorly understood. We previously showed that distinct patterns of neural activity and motor output are evoked by carbon dioxide (CO₂) in starved adults vs dauers of Caenorhabditis elegans, despite the two life stages displaying the same preference (attraction) for CO₂. However, how the distinct CO2-evoked neural dynamics and motor patterns contribute to CO2 attraction at the two life stages remained unclear. Here, using a CO₂ chemotaxis assay, we show that different interneurons are employed to drive CO₂ attraction at the two life stages. We also investigate the molecular mechanisms that mediate CO₂ attraction in dauers vs adults. We show that insulin signaling promotes CO₂ attraction in dauers but not starved adults and that different combinations of neurotransmitters and neuropeptides are used for CO2 attraction at the two life stages. Our findings provide new insight into the distinct molecular and cellular mechanisms used by C. elegans at two different life stages to generate attractive behavioral responses to CO₂.

Keywords: carbon dioxide response; Caenorhabditis elegans; chemosensory behavior; sensory valence; gas sensing; dauer larva

Introduction

Chemosensation is a critical sensory modality that contributes to locating food, finding mates or hosts, and avoiding pathogens and predators (Hildebrand and Shepherd 1997). In some cases, the valence of a chemosensory stimulus, i.e. whether the stimulus is attractive or aversive, depends on an animal's physiological state (Ribeiro and Dickson 2010; Wasserman et al. 2012; Wasserman et al. 2013; Devineni and Scaplen 2021). However, in other cases, chemosensory valence remains constant despite dramatic changes in internal physiology (Schmidt and Beauchamp 1988; Hansson et al. 2010; Oleszkiewicz et al. 2022; Banerjee et al. 2023). The molecular and cellular mechanisms that enable animals to maintain the same valence state across widely varying physiological conditions remain poorly understood.

The response of the free-living nematode Caenorhabditis elegans to carbon dioxide (CO₂) offers a powerful model system for exploring the mechanisms that determine chemosensory valence (Banerjee and Hallem 2020). While well-fed adults are repelled by CO₂, starved adults are attracted to CO₂ (Bretscher et al. 2008; Hallem and Sternberg 2008; Bretscher et al. 2011; Hallem, Spencer, et al. 2011; Rengarajan et al. 2019). In addition, dauer larvae—long-lived, nonfeeding, stress-resistant larvae that form in response to adverse environmental conditions (Hu 2007)-are attracted to CO₂ (Hallem, Dillman, et al. 2011a; Banerjee et al. 2023). Thus, a comparison of the molecular and cellular mechanisms that drive CO₂ attraction in dauers vs starved adults can provide insight into how different life stages establish similar chemosensory preferences.

We previously showed that although both starved adults and dauers are attracted to CO₂, the neural activity dynamics and the motor outputs evoked by CO₂ differ across the two life stages (Rengarajan et al. 2019; Banerjee et al. 2023). For example, the AIY, RIG, and AIB interneurons show different CO₂-evoked calcium responses in starved adults vs dauers. AIY displays stochastic excitatory CO₂-evoked activity in starved adults (Rengarajan et al. 2019) but inhibitory CO2-evoked activity in dauers (Banerjee et al. 2023), whereas RIG and AIB do not respond to CO_2 in starved adults but display excitatory CO2-evoked activity in dauers (Rengarajan et al. 2019; Banerjee et al. 2023). However, how these distinct CO₂-evoked patterns of neural activity contribute to CO₂ attraction was unclear.

In this study, we correlate the CO₂-evoked activities of these interneurons with their roles in promoting CO₂ attraction in starved adults vs dauers. We show that individual interneurons make distinct contributions to CO2 attraction at the two life stages.

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Whereas AIY promotes CO_2 attraction in starved adults but not dauers, RIG and AIB promote CO_2 attraction in dauers but not starved adults. We also show that insulin signaling, which promotes CO_2 -evoked AIB activity in dauers (Banerjee *et al.* 2023), functions in neurons to selectively drive CO_2 attraction in dauers. In addition, we identify distinct combinatorial codes of neurotransmitters and neuropeptides that promote CO_2 attraction at the two life stages. Our results illuminate the different neurogenetic mechanisms that operate in dauers vs adults to establish the same chemosensory valence state.

Materials and methods

C. elegans strains

Worms were cultured on 2% nematode growth media (NGM) plates seeded with *Escherichia coli* OP50 bacteria at ambient temperature (~22°C) and CO₂ (~0.038%) as previously described (Stiernagle 2006; Scott 2011; Banerjee *et al.* 2023). The temperature-sensitive strains CB1370, DR1565, JT191, EAH404, EAH407, and EAH408 were maintained at 15°C but were moved to ambient temperature (~22°C) at least 24 h prior to experiments to minimize any effects of temperature shifts on behavior, as previously described (Banerjee *et al.* 2023). The strains JT709 and BQ1 were treated similarly for direct comparison with strain DR1565 in Fig. 2f. A complete strain list is provided in Supplementary Table 1.

Molecular biology and transgenesis

The strains where specific neurons were genetically ablated or silenced were generated in previous studies (Kunitomo *et al.* 2013; Luo *et al.* 2014; Calhoun *et al.* 2015; Jin *et al.* 2016; Guillermin *et al.* 2017). The *inx*- 6^{AIB} OFF strain, where *inx*-6 function was eliminated specifically in AIB, was generated in a previous study (Bhattacharya *et al.* 2019). For tissue-specific rescue of *daf*-2, the following plasmids containing the *daf*-2 cDNA were obtained from Addgene and individually injected into the CB1370 *daf*-2(*e*1370) strain at 30 ng/µL, along with *Pmyo*-2::*dsRed* (20 ng/µL) as a coinjection marker and pBlueScript (50 ng/µL): pJH4531 (Prgef-1::*daf*-2) for pan-neuronal rescue (Addgene #132366), pJH4723 (Pges-1::*daf*-2) for intestinal rescue (Addgene #178899), and pJH4733 (Pmyo-3:: *daf*-2) for muscle rescue (Addgene #178898) (Hung *et al.* 2014).

CO₂ chemotaxis assays

CO2 chemotaxis assays on starved adults and dauers were performed as previously described (Rengarajan et al. 2019; Banerjee et al. 2023). For assays with starved adults, young adults (~1 day old) were washed in a watch glass and then starved on a 9 cm 2% NGM plate without bacteria for 3 h. Animals were placed within an annular ring of Whatman paper soaked in 20 mM copper chloride (CuCl₂) solution to prevent the worms from migrating off the plate, since copper is aversive to C. elegans (Campbell et al. 2017). After 3 h of starvation, the paper ring was removed. Animals were then washed off the plate into M9 buffer and then washed twice in M9 and once in ddH₂O in a watch glass. For testing, animals were transferred from the watch glass onto the center of a 9 cm 2% NGM plate using a piece of Whatman paper. For assays with dauer larvae, dauers were generated by transferring 8-10 young adults onto 2% NGM plates with a lawn of OP50 bacteria and leaving the plate at room temperature for 10-14 days until the bacterial lawn was consumed. Dauers were isolated from other life stages on the plate using a sodium dodecyl sulfate (SDS) resistance assay (Karp 2016) as previously described

CO2 chemotaxis assays were performed as previously described (Guillermin et al. 2017; Rengarajan et al. 2019; Banerjee et al. 2023). Animals were placed onto the center of a 9 cm 2% NGM plate. The CO_2 stimulus (the test concentration of CO_2 , 21% O_2 , balance N_2) and air stimulus (21% O_2 , balance N_2) were pumped through holes in opposite sides of the plate lid at a rate of 2 mL/min (for adults) or 0.5 mL/min (for dauers) to generate a CO₂ gradient (Supplementary Fig. 1a) using a syringe pump (PHD 2000, Harvard Apparatus), with the syringe output connected to the plate lid via ¼-inch flexible PVC tubing. Assay duration was 20 min (for adults) or 1 h (for dauers). After the assay, the number of animals within a 20-mm diameter circle under each gas inlet (for adults) or within 30-mm segments on the side of the plate (for dauers) were counted (Supplementary Fig. 1b and c). For transgenic strains with extrachromosomal arrays, only animals expressing the fluorescent transgene were counted. A chemotaxis index (CI) was then calculated as

 $CI = \frac{\# \text{ animalsatCO}_2 - \# \text{ animalsataircontrol}}{\# \text{ animalsatCO}_2 + \# \text{ animalsataircontrol}}.$

To account for directional bias due to vibration or other stimuli, assays were conducted in pairs, with the gas gradient in opposite orientations. If the absolute value of the difference in CI between the 2 assays in the pair was ≥ 0.9 , both assays were discarded as behavior was assumed to result from directional bias. Assays were also discarded if fewer than 7 animals navigated into the combined scoring regions. For strain RB2575, which showed decreased motility, single assays within a pair that had more than 7 animals in the combined regions were scored and included in the analysis even if fewer than 7 animals moved in the other assay in the pair, provided there was no directional bias within the pair. In the case of the neuropeptide genes ins-1, flp-2, and flp-17, 2 independent alleles of each gene were tested in CO₂ chemotaxis assays.

Histamine assays with dauers

Transgenic dauers expressing the histamine-gated chloride channel HisCl1 (Pokala *et al.* 2014) were isolated using 1% SDS treatment as described above. Dauers were then incubated in 20 mM histamine solution (in dH₂O) for 1 h prior to assays. For the nohistamine controls, dauers were incubated in dH₂O without histamine for the same duration. Dauers were then transferred onto 2% NGM plates without bacteria, with or without 20 mM histamine, and CO₂ chemotaxis assays were performed as described above.

Microscopy

For starved adults, 1-day-old adults were starved for 3 h prior to imaging. Dauers for imaging were isolated by SDS treatment as described above. Animals were anesthetized using 10 mM levamisole and placed onto 2% agarose pads on glass slides. Imaging was performed with a Zeiss Axio Observer inverted wide-field fluorescent microscope equipped with a Colibri 7 for LED fluorescence illumination, a Plan-APOCHROMAT 20x objective lens, a Hamamatsu ORCA-Flash 4.0 camera, and Zen software (Zeiss). For Supplementary Fig. 3, images were captured as z-stacks and maximum intensity projections were constructed using Fiji (Schindelin *et al.* 2012).

Statistical analysis

Statistical tests were performed using GraphPad Prism v9.3.1. Normality was determined using the D'Agostino–Pearson omnibus normality test. If data were not normally distributed, nonparametric tests were used. Power analyses were performed using G*Power v3.1.9.6 (Faul *et al.* 2007).

Results

Distinct sets of interneurons promote CO₂ attraction in dauers vs starved adults

In both starved adults and dauers, CO₂ is detected by the paired BAG sensory neurons in the head (Hallem and Sternberg 2008; Hallem, Spencer, et al. 2011; Smith et al. 2013; Rengarajan et al. 2019; Banerjee et al. 2023). However, many of the interneurons downstream of BAG show different patterns of CO₂-evoked neural activity in starved adults vs dauers (Fig. 1a) (Banerjee et al. 2023), which may in part reflect differences in synaptic connectivity of these interneurons with the BAG neurons at the two life stages (Fig. 1a) (White et al. 1986; Varshney et al. 2011; Bhattacharya et al. 2019; Yim et al. 2023). To understand the contribution of these different interneurons to CO₂ attraction at the two life stages, we used a CO₂ chemotaxis assay (Supplementary Fig. 1) to examine the behavioral responses of strains where individual interneurons downstream of BAG were either genetically ablated or silenced (Carrillo et al. 2013; Guillermin et al. 2017; Katz et al. 2018; Rengarajan et al. 2019). We focused on the RIG, AIY, AVE, AIB, and RIA interneurons; strains lacking RIB and AVA function were not tested because they did not produce enough dauers for chemotaxis assays, indicating a possible role for these neurons in promoting dauer entry.

We found that RIG- and AIB-ablated dauers showed reduced CO₂ attraction compared with wild type, whereas AVE-silenced dauers were repelled by instead of attracted to CO₂ (Fig. 1b, Supplementary Fig. 2a and b). In contrast, the AIY and RIA neurons were not required for CO₂ attraction in dauers across all tested concentrations (Fig. 1b, Supplementary Fig. 2c and d). Thus, RIG, AIB, and AVE promote CO₂ attraction in dauers, whereas AIY and RIA do not. We note that whereas the RIG, AIY, and AIB promoters we used to genetically ablate these interneurons showed the expected cell-specific expression pattern in starved adults and dauers, the promoter used to genetically silence AVE (opt-3) showed faint expression in several neurons in addition to AVE in dauers (Supplementary Fig. 3). Thus, we cannot exclude the possibility that one or more of these opt-3-expressing neurons also contribute to CO₂ response in dauers.

To further investigate the roles of CO₂ microcircuit interneurons in regulating CO2 attraction across life stages, we compared the behavioral responses of interneuron-ablated starved adults and dauers using a CO₂ chemotaxis assay. We found that wildtype and AIB-ablated starved adults were similarly attracted to CO₂, suggesting that these neurons drive CO₂ attraction in dauers but not starved adults (Fig. 1c). RIG is also required for CO₂ attraction in dauers but not starved adults (Fig. 1d). In contrast, AIY is required for CO₂ attraction in starved adults (Rengarajan et al. 2019) but not dauers (Fig. 1e), whereas RIA is not required to establish CO₂ attraction at either life stage (Fig. 1b) (Rengarajan et al. 2019). To confirm the role of AIB in regulating CO_2 response in dauers, we examined the behavioral responses of dauers in which the AIB neurons were chemogenetically silenced using the histamine-gated chloride channel HisCl1 (Pokala et al. 2014). Dauers expressing HisCl1 specifically in AIB showed a significant reduction in CO₂ attraction when treated with exogenous histamine compared with untreated control dauers (Fig. 1f), confirming that AIB promotes CO₂ attraction in dauers. To further confirm the role of RIG in promoting CO₂ attraction in dauers, we hyperactivated RIG by expressing a gain-of-function allele of the protein kinase C (*pkc-1*) gene (Sieburth *et al.* 2005; Sieburth *et al.* 2007) specifically in RIG (Guillermin *et al.* 2017). We found that hyperactivation of RIG results in enhanced CO₂ attraction in dauers (Fig. 1g), providing additional evidence that RIG promotes CO₂ attraction in dauers. Thus, both AIB and RIG promote CO₂ attraction in dauers but not starved adults. Together, our results demonstrate that distinct interneurons are required for establishing the same valence state at 2 different life stages.

We then tested the effects of dauer-specific changes in neuronal connectivity on the life stage-specific requirement for interneurons in establishing CO₂ attraction. AIB was previously shown to form gap junctions with BAG in dauers but not adults-the innexin INX-6 is expressed in AIB specifically in dauers, where it forms a gap junction complex with CHE-7, a gap junction subunit expressed in BAG (Fig. 2a) (Bhattacharya et al. 2019). Consistent with previous results (Bhattacharya et al. 2019), che-7 mutant dauers, as well as dauers where inx-6 expression is eliminated specifically in AIB (inx-6^{AIB OFF}), showed significantly reduced CO₂ attraction compared with wild-type dauers (Fig. 2b). In contrast to inx-6^{AIB OFF} dauers, inx-6^{AIB OFF} starved adults showed no defects in CO₂ attraction (Fig. 2c). Further, ectopic expression of inx-6 in AIB did not affect CO₂ responses in starved adults (Fig. 2d), indicating that additional mechanistic differences underlie CO₂ attraction in dauers vs starved adults.

Insulin signaling promotes CO₂ attraction in dauers but not starved adults

We then investigated the molecular mechanisms that regulate CO₂ attraction in dauers and starved adults. Insulin signaling regulates entry into the dauer state (Hu 2007) as well as a diverse array of chemosensory behaviors in adults (Tomioka *et al.* 2006; Hallem and Sternberg 2008; Adachi *et al.* 2010). Moreover, we previously showed that insulin signaling modulates the CO₂-evoked neuronal activity of AIB in dauers (Banerjee *et al.* 2023). We therefore examined the role of insulin signaling in regulating CO₂ attraction in starved adults and dauers.

We first examined the CO2 response of animals carrying a loss-of-function mutation in the *daf-2* gene, which encodes the sole C. elegans insulin receptor (Hu 2007). We found that loss of daf-2 function had no effect on CO₂ attraction in starved adults (Fig. 2e). In contrast, daf-2 mutant dauers were unresponsive to CO₂, suggesting that daf-2 promotes CO₂ attraction in dauers (Fig. 2e-f). The daf-2 gene is broadly expressed in multiple tissues in both adults and dauers (Murphy and Hu 2013; Martinez et al. 2020). To identify the tissues where *daf-2* might function to regulate CO₂ attraction in dauers, we performed CO₂ chemotaxis assays on daf-2 mutant dauers in which DAF-2 function was specifically restored in either neurons, intestine, or muscle. Restoring DAF-2 function in neurons, but not intestine or muscle, partially yet significantly rescued the chemotaxis defect of daf-2 mutant dauers (Supplementary Fig. 4). These results suggest that DAF-2 primarily functions in neurons to mediate CO₂ attraction in dauers. In addition, we found that dauers carrying loss-of-function mutations in the 3-phosphoinositide-dependent-kinase-1 gene pdk-1 and the protein kinase B (Akt/PKB) gene akt-1, both of which act downstream of daf-2 (Hu 2007), showed significantly reduced CO₂ attraction (Fig. 2f). Thus, insulin signaling promotes CO₂ attraction in dauers but not starved adults.



Fig. 1. Distinct interneurons establish CO₂ attraction in starved adults vs dauers. a) Synaptic connectivity of the BAG neurons with downstream interneurons and their CO₂-evoked neuronal activity in adults vs dauers (White *et al.* 1986; Rengarajan *et al.* 2019; Banerjee *et al.* 2023; Yim *et al.* 2023). The lighter shade of the BAG neurons in dauers indicates a reduced CO₂-evoked calcium response, as previously reported (Banerjee *et al.* 2023). Adapted from Banerjee *et al.* (2023). b) The interneurons RIG, AIB, and AVE promote CO₂ attraction in dauers. Strains where specific pairs of interneurons were genetically ablated (AIY-, RIG-, RIA-, or AIB-) or silenced with tetanus toxin (AVE::TeTx) were used. ****P < 0.0001, 'P < 0.05, ns = not significant, 1-way ANOVA with Dunnett's posttest. *n* = 14–22 trials per genotype. Responses shown are to 10% CO₂. c-e) Distinct interneurons establish CO₂ attraction in starved adults vs dauers. Behavioral responses of starved adults and dauers with genetically ablated AIB (c), RIG (d), and AIY (e) neurons in CO₂ chemotaxis assays. ****P < 0.0001, **P < 0.001, ns = not significant, 2-way ANOVA with Sidak's posttest. *n* = 12–22 trials per genotype. Responses shown are to 10% CO₂. f) Behavioral responses of wild-type dauers and transgenic dauers with AIB-specific expression of the histamine-gated chloride channel HisCI1 (Pokala *et al.* 2014). Dauers expressing HisCI1 in AIB (AIB::HisCI1) show significantly reduced CO₂ attraction in dauers. ****P < 0.0001, ns = not significant, 2-way ANOVA with Sidak's posttest. *n* = 8–12 assays per genotype and condition. Responses are to 10% CO₂. g) Dauers with RIG-specific expression of a *pkc-1* gain-of-function (*gf*) allele show significantly enhanced CO₂ attraction compared with wild-type dauers. *P < 0.05, Welch's t-test. *n* = 18–20 assays per genotype. Responses are to 1% CO₂. For b–g), each data point represents a single chemotaxis assay; solid lines indicate medians and dashed lines indicate interquar



Fig. 2. A BAG-AIB gap junction complex and insulin signaling are required for CO_2 attraction in dauers but not starved adults. a) A gap junction complex is formed between the BAG sensory neurons and the AIB interneurons in dauers by selective expression of the innexin subunit INX-6 in AIB (Bhattacharya *et al.* 2019). INX-6 partners with the innexin subunit CHE-7, which is expressed in BAG, to form gap junctions (Bhattacharya *et al.* 2019). b) Behavioral responses of AIB-ablated (AIB-) dauers, dauers containing an AIB-specific knockout of *inx*-6 (*inx*-6^{AIB OFF}) (Bhattacharya *et al.* 2019), and *che*-7 mutant dauers. ****P < 0.0001, ***P < 0.001, ***P < 0.01, 1-way ANOVA with Dunnett's posttest. *n* = 14–22 trials per genotype. c) CO₂ attraction in starved adults remains unaffected in *inx*-6 (*inx*-6^{AIB OFF}) mutants. ns = not significant, Mann–Whitney test. *n* = 16 trials per genotype. d) Ectopic expression of INX-6 in AIB does not alter CO₂ responses in starved adults. Behavioral responses of starved adults expressing *inx*-6 specifically in AIB under the control of the *npr*-9 promoter (*Pnpr*-9:*inx*-6) to CO₂ across concentrations. *n* = 12–16 trials per genotype and condition. Lines show medians and interquartile ranges. ns = not significant, 2-way ANOVA with Sidak's posttest. e) Behavioral responses of wild-type vs *daf*-2 mutant starved adults and dauers. DAF-2 promotes CO₂ attraction in dauers but not starved adults. *n* = 12–16 trials per genotype. ****P < 0.001, *n* = not significant, Kruskal–Wallis test with Dunnet's posttest. *g* Behavioral responses of dauers carrying loss-of-function mutations in candidate ILP genes. ***P < 0.001, *n* = not significant, Kruskal–Wallis test with Dunn's posttest. *n* = 8–34 trials per genotype. h) Behavioral responses of starved adults. ***P < 0.001, ns = not significant, 2-way ANOVA with Sidak's posttest. genotype. For b–c) and e–h), each data point represents a single chemotaxis assay; solid lines indicate medians and dauers in the insulin-l

We next sought to identify the DAF-2 ligands that modulate CO_2 response. The *C. elegans* genome encodes 40 insulin-like peptides (ILPs) (Li and Kim 2008). To identify ILPs that may be involved in regulating CO_2 response in dauers, we examined a subset of ILP genes known to be transcriptionally upregulated upon dauer entry (Lee et al. 2017). We found that dauers carrying two independent loss-of-function mutations in the ins-1 gene showed significantly reduced CO_2 attraction compared with wild-type dauers (Fig. 2g and h). In contrast, CO_2 attraction in starved adults was unaffected in ins-1 mutants (Fig. 2h). Thus, the modulatory effects of insulin signaling on CO_2 attraction in dauers are mediated at least in part by INS-1 peptides. Together, our results demonstrate that insulin signaling regulates CO_2 response valence in a life stage-dependent manner.

Both shared and life stage–specific neuropeptides and neurotransmitters regulate CO_2 attraction

Neuropeptide signaling plays a critical role in modulating neuronal function and behavior across species (Li and Kim 2008) and was previously shown to modulate the CO₂-evoked behavioral responses of *C. elegans* adults (Guillermin *et al.* 2017; Rengarajan *et al.* 2019). To examine whether CO₂ responses in dauers are regulated by neuropeptides, we performed a candidate screen of dauers carrying loss-of-function mutations in 16 neuropeptide genes (Fig. 3a). We focused on neuropeptide genes that are either expressed in BAG (Hallem, Spencer, *et al.* 2011), transcriptionally upregulated upon dauer entry (Lee *et al.* 2017), known to regulate CO₂ response in adults (Guillermin *et al.* 2017;



Fig. 3. Both shared and life stage–specific neurotransmitters and neuropeptides shape CO_2 attraction in starved adults vs dauers. a) A reverse genetic screen for neuropeptides that regulate CO_2 attraction in dauers. Behavioral responses of dauers with loss-of-function mutations in neuropeptide genes. **P < 0.01, *P < 0.05, Kruskal–Wallis test with Dunn's posttest. n = 8-42 trials per genotype. The specific loss-of-function (lf) alleles used for flp-2 and flp-17 were ok3351 and ok3587, respectively. Loss-of-function (lf) alleles used for other neuropeptide genes are indicated in the Materials and methods and Supplementary Table 1. b) and c) Behavioral responses of starved adults and dauers containing loss-of-function mutations in the neuropeptide genes flp-2 (b) and flp-17 (c). ****P < 0.0001, ns = not significant, 2-way ANOVA with Sidak's posttest. n = 12-20 trials per genotype and life stage. d–f) Behavioral responses of starved adults and the vesicular glutamate transporter gene *eat*-4 (d), the tyrosine decarboxylase gene *tdc*-1 (e), and the tyramine β -hydroxylase gene *tbh*-1 (f). ****P < 0.001, ***P < 0.001, ns = not significant, 2-way ANOVA with Sidak's posttest. n = 12-18 trials per genotype and life stage. For a-f), each data point represents a single chemotaxis assay; solid lines indicate medians and dashed lines indicate interquartile ranges. Responses shown are to 10% CO₂.

Rengarajan et al. 2019), or known to encode neuropeptides that are dependent on the neuropeptide-processing gene *sbt*-1, since *sbt*-1 promotes dauer entry and regulates CO₂ attraction in dauers (Lee et al. 2017).

We found that the FMRFamide-like neuropeptide gene flp-16 and the neuropeptide-like protein gene nlp-1, which were previously shown to regulate CO₂ attraction in starved adults (Rengarajan et al. 2019), are not required to promote CO₂ attraction in dauers (Fig. 3a). Thus, NLP-1 and FLP-16 neuropeptides play adult-specific roles in regulating CO2 response. However, dauers lacking the FMRFamide-like neuropeptide gene flp-2 showed significantly reduced CO₂ attraction (Fig. 3a and b), whereas starved adults lacking flp-2 responded normally to CO₂ (Fig. 3b). Thus, FLP-2 neuropeptides are required to establish CO₂ attraction in dauers but not starved adults. In addition, we found that both dauers and starved adults carrying loss-of-function mutations in the BAG-expressed neuropeptide gene *flp-17* showed significantly reduced CO₂ attraction (Fig. 3a and c). Thus, our results demonstrate that FLP-17 promotes $\ensuremath{\text{CO}}_2$ attraction in both dauers and starved adults.

We also investigated the role of neurotransmitters in regulating CO₂ attraction in starved adults vs dauers. We first examined the role of glutamate signaling, since the CO₂-detecting BAG neurons express the vesicular glutamate transporter EAT-4 (Serrano-Saiz et al. 2013) and well-fed adults carrying a loss-of-function mutation in the eat-4 gene show dramatically reduced CO₂ repulsion (Guillermin et al. 2017). We found that starved adults lacking eat-4 showed significantly reduced CO₂ attraction (Fig. 3d). In contrast, CO2 attraction remained unaffected in eat-4 mutant dauers (Fig. 3d). Thus, glutamate signaling is required for CO_2 attraction in starved adults but not dauers. Finally, we examined the role of biogenic amines in regulating CO₂ attraction. Both the tyrosine decarboxylase gene tdc-1, which is required for the biosynthesis of tyramine and octopamine (Chase and Koelle 2007), and the tyramine β -hydroxylase gene tbh-1, which is required for the conversion of tyramine to octopamine (Chase and Koelle 2007), promote CO₂ attraction in starved adults (Fig. 3e and f) (Rengarajan et al. 2019). We found that both tdc-1 and tbh-1 also promote CO₂ attraction in dauers, since tdc-1 and tbh-1 mutant dauers show decreased CO₂ attraction (Fig. 3e

(a)

Life stage / Condition	Sensory neuron		Valence state			
Starved adults	BAG	RIG	AIY	RIA	AIB	CO ₂ attraction
Dauers	BAG	RIG	AIY	RIA	AIB	CO ₂ attraction

(b)

Life stage / Condition		Behavior							
Starved adults	Glutamate	FLP-17	Biogenic amines	FLP-16	NLP-1	FLP-2	INS-1	DAF-2	CO ₂ attraction
Dauers	Glutamate	FLP-17	Biogenic amines	FLP-16	NLP-1	FLP-2	INS-1	DAF-2	CO ₂ attraction

Fig. 4. Distinct mechanisms shape CO_2 response valence across life stages. a) Table showing the requirement for the BAG sensory neurons and downstream interneurons in establishing CO_2 response valence in starved adults vs dauers. Neurons involved in establishing CO_2 attraction are shaded; neurons not required for CO_2 attraction are unshaded. b) Table showing the requirement for different molecular signals in establishing CO_2 response valence across life stages. Neurotransmitters and neuropeptides required to establish CO_2 attraction are indicated by shaded boxes; the lack of requirement for a particular molecular signal in establishing CO_2 attraction is indicated by an unshaded box.

and f). Thus, while glutamate signaling regulates CO_2 attraction specifically in starved adults, biogenic amine signaling regulates CO_2 attraction in both starved adults and dauers. Together, our results identify both shared and life stage–specific neuropeptides and neurotransmitters that regulate CO_2 attraction in starved adults vs dauers, suggesting that distinct combinations of neuropeptides and neurotransmitters regulate CO_2 attraction at the two life stages.

Discussion

We have shown that the same valence state, CO₂ attraction, is established in dauers and starved adults by distinct cellular and molecular mechanisms. The same CO₂-detecting sensory neurons are required for CO₂ attraction in both dauers and starved adults, but distinct interneurons mediate CO₂ attraction at the two life stages (Fig. 4a) (Hallem, Dillman, et al. 2011a; Rengarajan et al. 2019; Banerjee et al. 2023). The requirement for different interneurons at the dauer vs adult stage may arise from anatomical as well as functional differences in neuronal connectivity at the two life stages. For example, the AIY neurons promote CO₂ attraction in starved adults but not dauers (Fig. 4a), consistent with the dauer-specific loss of chemical synapses between BAG and AIY (Yim et al. 2023). However, AIY still displays CO₂-evoked activity in dauers (Banerjee et al. 2023), suggesting that this activity is independent of BAG-AIY chemical synapses. In addition, the AIB and RIG neurons promote CO₂ attraction in dauers but not starved adults despite forming chemical synapses with the BAG neurons at both life stages (Fig. 1a) (White et al. 1986; Yim et al. 2023). The distinct requirement for AIB in mediating dauer CO2 attraction arises due to dauer-specific BAG-AIB gap junctions (Bhattacharya et al. 2019). The role of RIG in selectively promoting CO₂ attraction in dauers may arise from changes in functional connectivity between BAG and RIG, although differences in anatomical connectivity between RIG and other neurons may also contribute. For example, RIG receives dauer-specific synaptic input from the IL2 neurons, which are required for nictation, a dauer-specific behavior that promotes dispersal by facilitating phoretic interactions between dauers and larger invertebrates (Lee *et al.* 2012) (Yim *et al.* 2023). Our results raise the possibility that RIG activity drives CO_2 attraction specifically in dauers to enable dauers to coordinate CO_2 response with nictation.

Our findings also demonstrate that distinct combinations of neurotransmitters and neuropeptides are required for CO₂ attraction at the two life stages (Fig. 4b). Further studies of the effects of these molecular signals on CO₂ microcircuit function will reveal the regulatory principles that enable the worm to establish the same valence state under distinct physiological conditions. For instance, biogenic amine signaling promotes CO₂ attraction in starved adults by regulating the CO2-evoked activity of AIY (Rengarajan et al. 2019). In dauers, biogenic amine signaling is also required for CO₂ attraction despite the lack of requirement for AIY (Fig. 4b), suggesting that biogenic amine signaling promotes CO₂ attraction via distinct neural mechanisms at the two life stages. Taken together, our results demonstrate that different mechanisms may operate at different life stages to promote the same valence state. In future studies, it will be interesting to explore whether life stage-specific chemosensory mechanisms function to couple chemosensory responses to other life stagespecific, ethologically relevant behavioral programs.

Data availability

Strains and plasmids are available upon request. All raw data and statistical analysis from this study are included in Supplementary File S1.

Supplemental material available at G3 online.

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Conflicts of interest

The authors declare no conflict of interest.

Author contributions

N.B., P.-Y.S., P.W.S., and E.A.H. conceived the study. N.B. and E.J.R.P. performed experiments. N.B. and E.J.R.P. analyzed the data. N.B. and E.A.H. wrote the manuscript. All authors read and approved the final manuscript.

Literature cited

- Adachi T, Kunitomo H, Tomioka M, Ohno H, Okochi Y, Mori I, Iino Y. 2010. Reversal of salt preference is directed by the insulin/PI3K and Gq/PKC signaling in *Caenorhabditis elegans*. Genetics. 186(4): 1309–1319. doi:10.1534/genetics.110.119768.
- Banerjee N, Hallem EA. 2020. The role of carbon dioxide in nematode behavior and physiology. Parasitology 147(8):841–854. doi:10. 1017/S0031182019001422.
- Banerjee N, Shih P-Y, Rojas Palato EJ, Sternberg PW, Hallem EA. 2023. Differential processing of a chemosensory cue across life stages sharing the same valence state in *Caenorhabditis elegans*. Proc Natl Acad Sci U S A. 120(19):e2218023120. doi:10.1073/pnas. 2218023120.
- Bhattacharya A, Aghayeva U, Berghoff EG, Hobert O. 2019. Plasticity of the electrical connectome of *C. elegans*. Cell 176(5):1174–1189. doi:10.1016/j.cell.2018.12.024.
- Bretscher AJ, Busch KE, de Bono M. 2008. A carbon dioxide avoidance behavior is integrated with responses to ambient oxygen and food in *Caenorhabditis elegans*. Proc Natl Acad Sci U S A. 105(23): 8044–8049. doi:10.1073/pnas.0707607105.
- Bretscher AJ, Kodama-Namba E, Busch KE, Murphy RJ, Soltesz Z, Laurent P, de Bono M. 2011. Temperature, oxygen, and saltsensing neurons in *C. elegans* are carbon dioxide sensors that control avoidance behavior. Neuron 69(6):1099–1113. doi:10.1016/j. neuron.2011.02.023.
- Calhoun AJ, Tong A, Pokala N, Fitzpatrick JA, Sharpee TO, Chalasani SH. 2015. Neural mechanisms for evaluating environmental variability in *Caenorhabditis elegans*. Neuron 86(2):428–441. doi:10. 1016/j.neuron.2015.03.026.
- Campbell JC, Chin-Sang ID, Bendena WG. 2017. A Caenorhabditis elegans nutritional-status based copper aversion assay. J Vis Exp. 125(e55939). doi:10.3791/55939.
- Carrillo MA, Guillermin ML, Rengarajan S, Okubo RP, Hallem EA. 2013. O₂-sensing neurons control CO₂ response in *C. elegans*. J Neurosci. 33(23):9675–9683. doi:10.1523/JNEUROSCI.4541-12. 2013.
- Chase DL, Koelle MR. 2007. Biogenic amine neurotransmitters in C. elegans, pp. 1–15 in WormBook, www.wormbook.org.

- Devineni AV, Scaplen KM. 2021. Neural circuits underlying behavioral flexibility: insights from *Drosophila*. Front Behav Neurosci. 15: 821680. doi:10.3389/fnbeh.2021.821680.
- Faul F, Erdfelder E, Lang AG, Buchner A. 2007. G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. Behav Res Methods. 39(2):175–191. doi:10.3758/BF03193146.
- Guillermin ML, Carrillo MA, Hallem EA. 2017. A single set of interneurons drives opposite behaviors in C. elegans. Curr Biol. 27(17):2630–2639. doi:10.1016/j.cub.2017.07.023.
- Hallem EA, Dillman AR, Hong AV, Zhang Y, Yano JM, DeMarco SF, Sternberg PW. 2011. A sensory code for host seeking in parasitic nematodes. Curr Biol. 21(5):377–383. doi:10.1016/j.cub.2011.01.048.
- Hallem EA, Spencer WC, McWhirter RD, Zeller G, Henz SR, Rätsch G, Miller DM III, Horvitz HR, Sternberg PW, Ringstad N. 2011. Receptor-type guanylate cyclase is required for carbon dioxide sensation by *Caenorhabditis elegans*. Proc Natl Acad Sci U S A. 108(1):254–259. doi:10.1073/pnas.1017354108.
- Hallem EA, Sternberg PW. 2008. Acute carbon dioxide avoidance in Caenorhabditis elegans. Proc Natl Acad Sci U S A. s105(23): 8038–8043. doi:10.1073/pnas.0707469105.
- Hansson BS, Knaden M, Sachse S, Stensmyr MC, Wicher D. 2010. Towards plant-odor-related olfactory neuroethology in Drosophila. Chemoecology 20(2):51–61. doi:10.1007/s00049-009-0033-7.
- Hildebrand JG, Shepherd GM. 1997. Mechanisms of olfactory discrimination: converging evidence for common principles across phyla. Annu Rev Neurosci. 20(1):595–631. doi:10.1146/annurev. neuro.20.1.595.

Hu PJ. 2007. Dauer, pp. 1–19 in WormBook. www.wormbook.org.

- Hung WL, Wang Y, Chitturi J, Zhen M. 2014. A Caenorhabditis elegans developmental decision requires insulin signaling-mediated neuron-intestine communication. Development 141(8): 1767–1779. doi:10.1242/dev.103846.
- Jin X, Pokala N, Bargmann CI. 2016. Distinct circuits for the formation and retrieval of an imprinted olfactory memory. Cell 164(4): 632–643. doi:10.1016/j.cell.2016.01.007.
- Karp X. 2016. Working with dauer larvae, pp. 1–19 in WormBook. www.wormbook.org. doi:10.1534/g3.116.030163.
- Katz M, Corson F, Iwanir S, Biron D, Shaham S. 2018. Glia modulate a neuronal circuit for locomotion suppression during sleep in C. elegans. Cell Rep. 22(10):2575–2583. doi:10.1016/j.celrep.2018.02.036.
- Kunitomo H, Sato H, Iwata R, Satoh Y, Ohno H, Yamada K, Iino Y. 2013. Concentration memory-dependent synaptic plasticity of a taste circuit regulates salt concentration chemotaxis in *Caenorhabditis elegans*. Nat Commun. 4(1):2210. doi:10.1038/ncomms3210.
- Lee H, Choi MK, Lee D, Kim HS, Hwang H, Kim H, Park S, Paik YK, Lee J. 2012. Nictation, a dispersal behavior of the nematode *Caenorhabditis elegans*, is regulated by IL2 neurons. Nat Neurosci. 15(1):107–112. doi:10.1038/nn.2975.
- Lee JS, Shih PY, Schaedel ON, Quintero-Cadena P, Rogers AK, Sternberg PW. 2017. FMRFamide-like peptides expand the behavioral repertoire of a densely connected nervous system. Proc Natl Acad Sci U S A. 114(50):E10726–E10735. doi:10.1073/pnas.1710374114.
- Li C, Kim K. 2008. Neuropeptides, pp. 1–36 in WormBook. www. wormbook.org.
- Luo L, Cook N, Venkatachalam V, Martinez-Velazquez LA, Zhang X, Calvo AC, Hawk J, MacInnis BL, Frank M, Ng JHR, et al. 2014. Bidirectional thermotaxis in *Caenorhabditis elegans* is mediated by distinct sensorimotor strategies driven by the AFD thermosensory neurons. Proc Natl Acad Sci U S A. 111(7):2776–2781. doi:10. 1073/pnas.1315205111.
- Martinez BA, Reis Rodrigues P, Nunez Medina RM, Mondal P, Harrison NJ, Lone MA, Webster A, Gurkar AU, Grill B, Gill MS.

2020. An alternatively spliced, non-signaling insulin receptor modulates insulin sensitivity via insulin peptide sequestration in *C. elegans*. eLife 9:e49917. doi:10.7554/eLife.49917.

- Murphy CT, Hu PJ. 2013. Insulin/insulin-like growth factor signaling in C. elegans, pp. 1–43 in WormBook. www.wormbook.org.
- Oleszkiewicz A, Schriever VA, Valder C, Agosin E, Altundag A, AvniH, Cao Van H, Cornejo C, Fishman G, Guarneros M, et al. 2022. Hedonic perception of odors in children aged 5–8 years is similar across 18 countries: preliminary data. Int J Pediatr Otorhinolaryngol. 157:111129. doi:10.1016/j.ijporl.2022.111129.
- Pokala N, Liu Q, Gordus A, Bargmann CI. 2014. Inducible and titratable silencing of *Caenorhabditis elegans* neurons in vivo with histamine-gated chloride channels. Proc Natl Acad Sci U S A. 111(7):2770–2775. doi:10.1073/pnas.1400615111.
- Rengarajan S, Yankura KA, Guillermin ML, Fung W, Hallem EA. 2019. Feeding state sculpts a circuit for sensory valence in *Caenorhabditis elegans*. Proc Natl Acad Sci U S A. 116(5): 1776–1781. doi:10.1073/pnas.1807454116.
- Ribeiro C, Dickson BJ. 2010. Sex peptide receptor and neuronal TOR/S6K signaling modulate nutrient balancing in Drosophila. Curr Biol. 20(11):1000–1005. doi:10.1016/j.cub.2010.03.061.
- Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S, Schmid B, et al. 2012. Fiji: an open-source platform for biological-image analysis. Nat Methods. 9(7):676–682. doi:10.1038/nmeth.2019.
- Schmidt HJ, Beauchamp GK. 1988. Adult-like odor preferences and aversions in three-year-old children. Child Dev. 59(4): 1136–1143. doi:10.2307/1130280.
- Scott K. 2011. Out of thin air: sensory detection of oxygen and carbon dioxide. Neuron 69(2):194–202. doi:10.1016/j.neuron.2010.12.018.
- Serrano-Saiz E, Poole RJ, Felton T, Zhang F, De La Cruz ED, Hobert O. 2013. Modular control of glutamatergic neuronal identity in *C. elegans* by distinct homeodomain proteins. Cell 155(3):659–673. doi: 10.1016/j.cell.2013.09.052.

- Sieburth D, Ch'ng Q, Dybbs M, Tavazoie M, Kennedy S, Wang D, Dupuy D, Rual J-F, Hill DE, Vidal M, *et al.* 2005. Systematic analysis of genes required for synapse structure and function. Nature 436(7050):510–517. doi:10.1038/nature03809.
- Sieburth D, Madison JM, Kaplan JM. 2007. PKC-1 regulates secretion of neuropeptides. Nat Neurosci. 10(1):49–57. doi:10.1038/nn1810.
- Smith ES, Martinez-Velazquez L, Ringstad N. 2013. A chemoreceptor that detects molecular carbon dioxide. J Biol Chem. 288(52): 37071–37081. doi:10.1074/jbc.M113.517367.
- Stiernagle T. 2006. Maintenance of C. *elegans*, pp. 1–11 in WormBook. www.wormbook.org.
- Tomioka M, Adachi T, Suzuki H, Kunitomo H, Schafer WR, Iino Y. 2006. The insulin/PI 3-kinase pathway regulates salt chemotaxis learning in *Caenorhabditis elegans*. Neuron 51(5):613–625. doi:10. 1016/j.neuron.2006.07.024.
- Varshney LR, Chen BL, Paniagua E, Hall DH, Chklovskii DB. 2011. Structural properties of the Caenorhabditis elegans neuronal network. PLoS Comput Biol. 7(2):e1001066. doi:10.1371/journal.pcbi.1001066.
- Wasserman S, Lu P, Aptekar JW, Frye MA. 2012. Flies dynamically anti-track, rather than ballistically escape, aversive odor during flight. J Exp Biol. 215(16):2833–2840. doi:10.1242/jeb.072082.
- Wasserman S, Salomon A, Frye MA. 2013. Drosophila tracks carbon dioxide in flight. Curr Biol. 23(4):301–306. doi:10.1016/j.cub.2012.12.038.
- White JG, Southgate E, Thomson JN, Brenner S. 1986. The structure of the nervous system of the nematode *Caenorhabditis elegans*. Philos Trans R Soc Lond B Biol Sci. 314(1165):1–340. doi:10.1098/rstb.1986.0056.
- Yim H, Choe DT, Bae JA, Kang H-M, Nguyen KCQ, Choi MK, Ahn S, Bahn SK, Yang H, Hall DH, et al. 2023. Mind of a dauer: comparative connectomics reveals developmental plasticity. bioRxiv 533915. https://doi.org/10.1101/2023.03.23.533915, 23 March 2023, preprint: not peer reviewed.

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